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Persistence of microbial and chemical pig manure markers as compared to faecal indicator bacteria survival in freshwater and seawater microcosms

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Abstract :

Natural seawater and freshwater microcosms inoculated with pig manure were set up to determine the persistence of pig faecal microbial and chemical markers in these two types of surface water. The concentrations of *Lactobacillus amylovorus*, the *Bacteroidales* Pig-2-Bac 16S rRNA genetic marker, five stanols and the evolution of two ratios of stanols, R_1 (coprostanol to the sum of coprostanol and 24-ethylcoprostanol) and R_2 (sitostanol to coprostanol) were analyzed during two months along with the concentration of Faecal Indicator Bacteria (FIB). Pig manure was inoculated to unfiltered water microcosms incubated aerobically at 18 °C in the dark. The faecal contamination load represented by the concentrations of culturable *Escherichia coli* and/or enterococci remained for two months in the freshwater and seawater microcosms water column. These concentrations followed a biphasic decay pattern with a 97% reduction of the initial amount during a first rapid phase (<6 days) and a remaining proportion undergoing a slower or null second decline. The *L. amylovorus* marker and five stanols persisted as long as the indicators in both treatments. The Pig-2-Bac marker persisted 20 and 27 days in seawater and freshwater, respectively. The ratios R_1 and R_2 were in the range specific to pig manure until day 6 in both types of water. These results indicate that Pig-2-Bac, *L. amylovorus* and stanol ratios might be used in combination to complement FIB testing to determine the pig source of fecal pollution. However, stanol ratios are to be used when the time point of the discharge is known.

Highlights

- ▶ *Escherichia coli* and enterococci from pig manure followed a biphasic decay in water microcosms.
- ▶ The marker *Lactobacillus amylovorus* followed the same trend as enterococci in fresh and seawater.
- ▶ The marker Pig-2-Bac followed the same trend as *E. coli* in seawater.
- ▶ Pig-specific stanols and *L. amylovorus* persisted for two months.
- ▶ Stanols and *L. amylovorus* detection and persistence was not affected by salinity.

Keywords : Faecal source tracking; Decay rates; Faecal stanols; Biphasic kinetic; *Lactobacillus amylovorus*; Bacteroidales marker

1. Introduction

Each year, the microbial analyses of the 540 freshwater and seawater bathing areas located in Brittany, North West of France, reveal that these areas might be sporadically contaminated by high faecal loads. Among the three main sources of faecal contamination (human, pig and cow) which can lead to the downgrading of these areas (Soller et al., 2010), pig manure spreading is considered as a potential factor of water pollution (Thurston-Enriquez et al., 2005). Brittany supports more than half of the national pigs' livestock while representing only 5% of the French territory and this high concentration generates from 8 to 10 million tons of pig manure each year. To protect public health whilst bathing and to improve management practices at bathing waters, the revised Bathing Water European Directive (2006/7/EC) requires to establish bathing water profiles to identify the source(s) of a faecal pollution upstream a targeted water body.

To track pollution to livestock facilities or diffuse non point sources, animal-specific markers have been proposed. They include DNA molecules from intestinal bacteria (Dick et al., 2005; Ufnar et al., 2007), endogenous eukaryotic cells (Balleste et al., 2010) and faecal stanols (Tyagi et al., 2009). Among pig genetic markers, studies have focused essentially on bacteria from the *Bacteroidales* order as potential markers preferred for their abundance in the gastrointestinal tract and host specificity occurrence (Dick et al., 2005; Mieszkin et al., 2009). Among chemical markers, faecal stanols are considered as direct markers because they occur in the faeces (Leeming et al., 1996). The distribution of those compounds in animal faeces depends on three host factors: (i) the animal's diet, (ii) the ability to biosynthesize

54 endogenous sterols and (iii) the occurrence of anaerobic bacteria able to biohydrogenate
55 sterols to stanols of various isomeric configurations. The combination of those three factors
56 determines the “sterol fingerprint” that is characteristic of each animal faeces (Leeming et al.,
57 1996).

58 In a previous study from our teams, several microbial and chemical markers detected in river
59 waters impacted by faecal pollutions were proposed to be used in a toolbox as Faecal Source
60 Tracking methods (FST) (Gourmelon et al., 2010). Two of which were microbial pig-specific
61 markers previously validated for their host specificity, the genetic *Bacteroidales* marker Pig-
62 2-Bac and the bacterial species *Lactobacillus amylovorus* (Marti et al., 2010; Mieszkin et al.,
63 2009). Five faecal stanols found in pig faeces and two steroids ratios were also selected to be
64 part of the toolbox. Specific markers should exhibit high host-specificity and represent the
65 load of faecal pollution (Field and Samadpour, 2007). In this scope, the evaluation of markers
66 to be used as FST must consider whether the degradation and transport characteristics of the
67 markers are similar to that of one or several pathogens or to the traditional faecal indicator
68 bacteria (FIB) such as *Escherichia coli* (*E. coli*) and enterococci.

69 The present study examines the detection of Pig-2-Bac marker, *L. amylovorus* and faecal
70 stanols as compared to the detection of FIB in natural surface water microcosm. The faecal
71 stanols selected comprised coprostanol (5 β -cholestan-3 β -ol) and 24-ethylcoprostanol (24-
72 ethyl-5 β -cholestan-3 β -ol) dominant in fresh pig manure (Leeming et al., 1996; Shah et al.,
73 2007). We also included epicoprostanol (5 β -cholestan-3 α -ol) and campestanol (24-methyl-
74 5 α -cholestan-3 β -ol) since the land spreading of pig manure as a soil fertilizer results in high
75 concentration of those compounds (Jarde et al., 2009). Two ratios of concentrations of
76 steroids were calculated along the experiment: coprostanol to the sum of coprostanol and 24-
77 ethylcoprostanol (R_1) and sitostanol to coprostanol (R_2). In the previous study to this work,
78 pig manure was characterized by a R_1 ratio of 0.57 ± 0.02 and a R_2 ratio of 0.3 ± 0.1
79 (Gourmelon et al., 2010).

80 The objective of this study was to estimate the persistence and decay rates of pig genetic
81 markers, faecal stanols, and FIB in fresh and marine water during two months. The evaluation
82 to which extent the detection of tested markers and ratios correlates with that of FIB and
83 hence, represents faecal load contamination should allow to validate their usefulness as
84 markers of pig faecal pollution in these two types of surface water.

85

86 **2. MATERIALS AND METHODS**

87 **2.1. Microcosms design**

88 Microcosms consisted of 6 one hundred-litres inert glass aquariums placed in a dark room,
89 protected from sunlight and fluctuating temperature. Three of which were filled with seawater
90 and the remaining aquariums with freshwater. Both waters were not filtered to study the
91 persistence of FIB and specific markers in presence of protozoa. Waters were seeded with pig
92 liquid manure. Constant mixing was achieved with the aid of a helix agitator and oxygen
93 saturation with air pumped in throughout the experiment. A plastic film cover limited
94 evaporation of water. The type of surface water was the changing parameter. Ambient
95 temperature (around 18 °C) corresponded with surface water temperature during the warmer
96 months in Brittany (France). Nine hundred mL of untreated liquid pig manure was added to
97 90 L of water (1:100 dilution). This ratio was chosen to represent a high faecal load
98 contamination likely to remain for two months. However, the turbidity resulting from this
99 parameter hindered light treatment.

100 Sampling took place on the starting day, then on day 2, 6, 13, 20, 27, 34, 41, 48 and 55. Both
101 unseeded initial types of water were kept in the same conditions during the whole experiment
102 to use as controls, sampling took place on the starting day and on day 55. Culturable *E. coli*
103 (cEC), culturable enterococci (cENT), bacterial genetic markers and stanol concentrations
104 were measured at each sampling point. Dissolved O₂ concentration and temperature were
105 measured every 3 to 4 days. Although constant mixing was achieved in the middle of the

106 aquarium, sedimentation occurred during the course of the experiment and biofilms formation
107 occurred on the walls. Samples were drawn from the water column.

108

109 **2.2. Water and pig manure samples**

110 The seawater was collected in the end of January 2010 in the Atlantic Ocean from Landunvez,
111 in the NW Brittany region of France (lat × long: 48.540819 × -4.751587). Salinity was 33 g/L,
112 total dissolved organic carbon measurement was 0.4 mg C/L, total dissolved nitrogen was 4.0
113 mg N/L. The freshwater was sampled from a lake in Commana in Brittany (lat × long:
114 48.3887488 × -4.0177564) on the same day, the total dissolved organic carbon was 2.7 mg
115 C/L, the total dissolved nitrogen was 9.3 mg N/L. Pig manure was collected from a farm
116 located in Brittany and samples were taken from a storage tank after homogenisation with
117 propeller agitator for 20 minutes, the total dissolved carbon and total dissolved nitrogen were
118 6.5 g C/L, and 2.9 g N/L, respectively.

119

120 **2.3. Enumeration of FIB**

121 Depending on sample turbidity, FIB counts were achieved either by serial dilution in buffered
122 peptoned water (Oxoid, Basingstoke, England) or by filtration of 100 mL of sample on a 0.45
123 µm cellulose membrane (Whatman, Dassel, Germany). Filters or 0.1 mL of the dilution was
124 plated on TBX agar (Oxoid) and on Slanetz and Bartley agar (Biokar Diagnostics, Beauvais,
125 France). TBX plates were incubated for 24 h at 44 °C. Blue colonies (glucuronidase positive)
126 were counted to determine the concentration of *E. coli*. After incubation at 37 °C for 48 h,
127 membranes on Slanetz and Bartley agar were transferred onto Bile-Esculin-Azide agar (BEA)
128 (Biokar Diagnostics) and incubated for 2 h at 44 °C. Black colonies on BEA were counted as
129 enterococci. The detection limit of both methods was 1 colony forming units (CFU) per 100
130 mL.

131

132 **2.4. Microbial markers analyses**

133 **2.4.1. Samples preparation and DNA extraction**

134 Two hundreds mL of samples were either centrifuged (9000 g for 15 min) or filtered onto 0.2
135 μm polycarbonate membrane (Sartorius, Goettingen, Germany) depending on suspended
136 matter density. Filtration was the preferred method to recover DNA. However, filtration was
137 not possible until day 13. On that day, samples from one microcosm of both types of water
138 were treated by both methods. Since qPCR results were similar, it was decided to carry on
139 with filtration. From 0.30 to 250 mg of solid matter were recovered. DNA extraction was
140 performed on sample solid matter with the aid of the FastDNA® SPIN for Soil kit (MP
141 Biomedicals, Illkirch, France), following manufacturer's instructions. The elution volume was
142 100 μL .

143

144 **2.4.2. Real-time PCR**

145 *2.4.2.1. Oligonucleotide primers and probes*

146 The pig-specific *Bacteroidales* 16S rRNA gene marker (Pig-2-Bac) and the *L. amylovorus*
147 marker were quantified with the primers and probe described by Mieszkin et al. (2009) and
148 Konstantinov et al. (2005), respectively.

149 *2.4.2.2. DNA standard curves*

150 For the quantification of the *Bacteroidales* marker Pig-2-Bac, standard curves were generated
151 from serial dilutions of known concentration of plasmid DNA ranging from 5×10^7 to 5×10^0
152 copies per reaction. Linear plasmids were extracted with the QIAquick Miniprep Extraction
153 Kit (Qiagen), following the manufacturer's instructions. The linear form of plasmid was
154 obtained with *NotI* enzyme (Roche Diagnostics, Meylan, France) in a final volume of 50 μL
155 for 3 h at 37 °C. The PCR standard curve for the *L. amylovorus* markers was prepared by 10-
156 fold dilution of bacterial genomic DNA extracted from one mL of a pure culture of *L.*
157 *amylovorus* DSM16698, with the Wizard genomic DNA purification Kit (Promega, Madison,

158 USA) according to the manufacturer's instructions. Dilutions ranged from 4.5×10^6 to $4.5 \times$
159 10^0 CFU equivalent from direct plating count considering that 100 % of the DNA from the
160 culture was recovered. Standard curves were generated by plotting threshold cycles (Ct)
161 against 16S rRNA genes or CFU equivalent, depending on the marker. Standard curves were
162 obtained by means of 3 replicates per point.

163 2.4.2.3. Real-time PCR assays

164 For Pig-2-Bac marker, amplification was performed using the Chromo4 real-time detection
165 system associated with Bio-Rad Opticon Manager software version 3.1 (Bio-Rad, Hercules,
166 CA). Real-time PCR was performed using the TaqMan[®] Brilliant II QPCR Master Mix kit
167 (Agilent technologies, Massy, France). Each reaction was run in triplicate. The cycle
168 conditions were 1 cycle at 95 °C for 10 min, followed by 40 cycles at 95 °C for 15 s and 60
169 °C for 1 min. Reactions were carried out in a final volume of 25 µL with primers and probe
170 final concentration being 300 nM and 200 nM respectively. Quantification limit was 1250
171 16S rRNA gene copies per 100 mL. The presence/absence of PCR inhibitors was verified
172 using an Internal Positive Control (IPC; AppliedBiosystem, France). Samples were diluted if
173 inhibitors were present.

174 Concerning the *L. amylovorus* marker, PCR was performed on the CFX96 real time system
175 (Bio-Rad), with the software Opticon Monitor version 3.1.32 and CFX manager version 1.1
176 (Bio-Rad), using the IQ SYBR-Green Supermix (Bio-Rad). The cycle conditions were 1 cycle
177 at 95 °C for 3 min, followed by 40 cycles at 95 °C for 15 s and 60 °C for 45 s. Reactions were
178 carried out in a final volume of 25 µL with primers final concentration being 200 nM.
179 Quantification limit was 112.5 CFU equivalents per 100 mL.

180 No filtration, extraction and template positive controls from a known concentration of a
181 control DNA fragment were included in the method to evaluate the yield of DNA recovery.

182

183 2.5. Faecal stanols analysis

184 Five faecal stanols, namely coprostanol, epicoprostanol, 24-ethylcoprostanol, campestanol
185 and sitostanol were investigated in this study. Analyses were performed on 1 L of non seeded
186 initial waters and 500 mL of microcosm waters. Faecal stanols were extracted from the
187 dissolved phase ($< 0.7 \mu\text{m}$) by solid phase extraction and quantified by gas chromatography –
188 mass spectrometry (GC-MS) according to the protocol developed by Jeanneau et al. (2011).
189 Cholesterol d6 (2,2,3,4,4,6- $^2\text{H}_6$ -5-cholesten-3 β -ol) was used as a recovery standard and was
190 added to water samples prior to the extraction step. 5 α -cholestane was used as an internal
191 standard and was added prior to the GC-MS analysis. Faecal stanols were quantified by the
192 internal standard method using a five-point calibration curve. The resulting calibration ranged
193 from 0.01 to 40 $\mu\text{g/L}$ for stanols in water.

194

195 **2.6. Decay rate calculations**

196 The decay rates of faecal stanols were calculated based on a first order decay model (Chick
197 model):

$$198 C_{(t)} = C_0 \times e^{-kt} \text{ or } \ln(C_{(t)}/C_0) = -kt$$

199 Where C_0 is the average initial concentration of the target in $\mu\text{g/L}$, $C_{(t)}$ is the target average
200 concentration at time t in $\mu\text{g/L}$, t is the time in days and k the decay constant or rate in days^{-1} .

201 The model describes a linear regression, k is the slope of the regression line and R^2 the
202 regression coefficient.

203 The biphasic model described by Lee et al. (2001) was used to calculate decay rates for every
204 microbial target in both treatments. Two constants were calculated from a biphasic first order
205 decay model (Cerf model):

$$206 C_{(t)} = C_0 \times (f \times e^{-k_1 t} + (1-f) \times e^{-k_2 t}) \text{ or } \ln(C_{(t)}/C_0) = \ln(f \times e^{-k_1 t} + (1-f) \times e^{-k_2 t})$$

207 Where f is the proportion of C_0 that declined during the first phase, k_1 is the decay constant of
208 the first phase and k_2 the decay constant of the second phase. $C_{(t)}$ and C_0 are expressed in
209 CFU/100 mL for FIB, in CFU equivalent/100 mL for *L. amylovorus* and in DNA copies/100

210 mL for Pig-2-Bac. The biphasic model and associated parameters were obtained with the aid
211 of XLSTAT 2010.4 using the nonlinear regression modelling.

212 Decay rates were calculated until the day concentrations were below quantification limit or
213 until day 55 when the detection limit was not reached.

214 The length of time (expressed in days) needed to obtain a 90% reduction in initial
215 concentration of stanols or bacteria was calculated as follows: $T_{90} = -\ln(0.1)/k$ (Chick model)
216 or, if f was $> 90\%$, $T_{90} = -\ln(0.1)/kI$ (Cerf Model)

217

218 **2.7. Regression tests**

219 In order to compare occurrence and concentration of FIB and stanols with time, regressions
220 were performed by plotting average concentrations of each stanol against average
221 concentrations of FIB. Regression lines were drawn between four or several time points. The
222 regression coefficient R^2 values illustrate the relationship between the two variables
223 compared.

224

225 **2.8. Statistical analysis**

226 **2.8.1. Decay rates distribution**

227 The validity of parametric tests is limited to samples following a normal distribution. When
228 the distribution is unknown non parametric tests should be preferred. To infer on normality, a
229 large number of samples is required, therefore to check whether the decay rates obtained in
230 this study were normally distributed, we performed an additional experiment where 20
231 microcosms were investigated. These latter consisted of 2 L-polypropylene bottles placed in
232 the same conditions as in the study, filled with the same freshwater and the same pig manure
233 added to 1:100. For practical and economical reasons, only, culturable *E. coli* concentrations
234 were measured. The concentrations of cEC were followed every week for 6 weeks. Decay
235 constants $k1$ and $k2$ were found to be normally distributed with the Normality tests from

236 XLSTAT 2010.4 (n=20) (data not shown). As a result parametric tests were performed to
237 compare decay constants of FIB and microbial markers. However, regression coefficients R^2
238 were found not to follow a normal distribution, hence all R^2 were compared with the Mann
239 and Whitney test.

240

241 **2.8.2. Tests of significance**

242 Two null hypotheses were posed (i) no difference exists between decay rates of one target in
243 either treatments, (ii) no difference exists between decay rates of either indicator (cEC or
244 cENT) and marker in a particular treatment (seawater or freshwater). The risk α to reject the
245 null hypothesis while it might be true was set at 0.05.

246 All statistical analyses were calculated from the regression coefficients (R^2) and decay rates
247 (k) of three independent experimental replicates. Concerning microbial markers, an F-test was
248 performed to determine variance equality between set of samples prior to a two tailed
249 Student's t -test assuming equal or unequal variance depending on the F-test results. Analyses
250 were achieved by Microsoft Office Excel 2003.

251 It was not possible to compare decay rates from FIB and faecal stanols because the former
252 decay was described with a non linear regression model whilst the latter with a linear model.
253 Only the first null hypothesis could be considered. The distribution of stanols decay rates
254 could not be determined. As a consequence, to compare decay rates of stanols in freshwater
255 and seawater microcosms; a non parametric test for small samples of unknown distribution
256 (Mann-Whitney) was performed on XLSTAT 2010.4.

257 Decay rates were compared only if the R^2 of the linear regressions they originated from were
258 not statistically different. Otherwise, the test is stated to be non applicable (NA).

259

260 **3. RESULTS**

261 In this study, we examined the persistence of pig faecal markers in three independent
262 controlled unfiltered water microcosms under aerobic condition. Temperature oscillated
263 between 16 and 20 °C and dissolved O₂ ranged from 8.3 to 9.7 mg/L. The waters used to
264 constitute microcosms were free or slightly contaminated with FIB and stanols. In the
265 seawater, FIB were not detected. Among stanols, only coprostanol and sitostanol were
266 quantifiable with a concentration of 0.02 and 0.05 µg/L, respectively. In the freshwater, cEC
267 concentration was 60 CFU/100 mL and cENT were not detected. The concentrations of
268 coprostanol, epicoprostanol, 24-ethylcoprostanol and sitostanol were 0.04, 0.02, 0.02 and 0.07
269 µg/L, respectively whereas campestanol was not detected. Pig-specific DNA markers Pig-2-
270 Bac and *L. amylovorus* were not detected in both types of water. At the end of the experiment
271 FIB were not detected in both types of water controls in 100 mL. We thus inferred that
272 microflora and stanols present in the microcosms at day 0 arose from the pig manure.

273

274 **3.1. Decay curves**

275 **3.1.1 FIB and microbial markers**

276 FIB and microbial markers followed a biphasic first order decay kinetic (Fig. 1). We,
277 therefore, used the Cerf biphasic decay model to determine the decay rates of the first and
278 second phase. An example of this model, illustrated by the behaviour of cENT in seawater is
279 shown on Fig. A.2. The model shows a sharper slope for the regression of the first phase than
280 for the second phase. Hence k_1 is always higher than k_2 in this model (Table 1). All decay
281 curves fitted to the model ($R^2 > 0.90$). As indicated by the value of the f parameter –
282 proportion of the initial concentration of the target that declined during the first phase-, more
283 than 97% of the initial inoculum was lost during this phase. The decimal reduction time
284 occurred thus during the first phase soon after the beginning of the experiment and was
285 reached in less than 6 days independently of the bacteria or the microcosm conditions.
286 However, in many instances a small remaining proportion persisted until the end of the

287 experiment. Culturable *E. coli* were recovered until day 20 in seawater whilst it was still
288 detected on day 55 in freshwater microcosms. Culturable enterococci and *L. amylovorus*
289 concentrations were over the limit of quantification in both types of water on the last day of
290 the experiment. In seawater, Pig-2-Bac marker followed the same trend as *E. coli*, it reached
291 the quantification limit by day 13. On day 20, it was detected just below quantification limit
292 in the three microcosms. It was decided to take this point into account for calculations (Fig.
293 A.1). The week after Pig-2-Bac was still detected in two microcosms. In freshwater, it was
294 found over quantification limit until day 27 in the three microcosms (Fig. B.1), but remained
295 detectable until day 34 in two microcosms.

296

297 **3.1.2. Faecal stanols**

298 Faecal stanols followed a monophasic first order decay kinetic as illustrated by Fig. B.2. In
299 seawater microcosms, initial concentrations of coprostanol, episcoprostanol, 24-
300 ethylcoprostanol, campestanol and sitostanol were 16.7, 3.1, 12.1, 2.6 and 4.7 $\mu\text{g/L}$,
301 respectively. Their final concentrations reached 0.15, 0.05, 0.18, 0.03 and 0.10 $\mu\text{g/L}$,
302 respectively (Fig A.3). The concentrations observed after 55 days represented a degradation
303 of 98 to 99% of the initial amounts of the five stanols. Nevertheless, they remained higher
304 than the initial concentrations in seawater before the addition of pig manure. In freshwater
305 microcosms, initial concentrations of coprostanol, epicoprostanol, 24-ethylcoprostanol,
306 campestanol and sitostanol were higher than in seawater microcosms and were 33.4, 9.3, 25.5,
307 6.9 and 11.8 $\mu\text{g/L}$, respectively whereas their final concentrations were 0.49, 0.15, 0.53, 0.10
308 and 0.20 $\mu\text{g/L}$, respectively. At the end of the experiment, they were in average 13 ± 6 times
309 higher than the initial concentrations in freshwater before the addition of pig manure.
310 Furthermore, as observed in the seawater microcosms, they represented a degradation of 98 to
311 99% of the initial amount of the five stanols (Fig. B.3).

312

313 **3.2. Progression of stanol ratios**

314 At the beginning of the experiment, the initial values of R_1 (coprostanol/ (coprostanol + 24-
315 ethylcoprostanol)) and R_2 (sitostanol/coprostanol) ratios ranged between 0.57 and 0.58 and
316 between 0.20 and 0.23, respectively. Regardless the microcosms, the ratio R_1 progressively
317 decreased to reach a value of 0.47 (Fig. A.4), due to the difference of decay rate between
318 coprostanol and 24-ethylcoprostanol (Table 1). The changes in the values of R_2 were also
319 similar in freshwater and in seawater. They were most marked than those of R_1 ratio as R_2
320 ranged mainly from 0.20 to 0.91 with however a maximum value of 1.55 on day 20 (Fig.
321 B.4).

322

323 **3.3. Decay rates statistical analysis**

324 **3.3.1. Decay rates comparisons from seawater and freshwater microcosms**

325 The regression coefficients from non linear regressions were not statistically different. As a
326 consequence every k_1 and k_2 drawn from microbial markers kinetic models could be
327 compared. The results concerning the first null hypothesis are shown in Table 1. *L.*
328 *amylovorus* decay first constants (k_1) were statistically different whilst their seconds (k_2)
329 were not. The opposite was noted for Pig-2-Bac, its first decay constant was not different in
330 seawater or freshwater microcosms whilst they were significantly different during the second
331 phase.

332 For stanols, the regression coefficients from linear regressions were also not statistically
333 different. Thus, every k drawn from faecal stanols kinetics could be compared. According to
334 the Mann and Whitney non parametric test, stanols decay rates were not significantly different
335 in freshwater or seawater microcosms (Table 1).

336

337 **3.3.2. Decay rates comparisons from FIB and microbial markers**

338 In seawater, the first phase constant from Pig-2-Bac decay did not significantly differed from
339 that of FIB. Contrary to this pig-specific marker, the decrease of *L. amylovorus* was slower
340 than the ones of cEC ($p < 0.0001$) and of cENT ($p = 0.027$) during the first phase. During the
341 second phase, the decays of both genetic markers were not significantly different from that of
342 cENT whilst they were lower than that of cEC ($p < 0.001$). In freshwater, Pig-2-Bac and *L.*
343 *amylovorus* decays did not significantly differed from that of cENT in the first phase while
344 they were faster than cEC ($p = 0.001$). During the second phase, decay rates of both markers
345 were not statistically different from those of FIB.

346

347 **3.4. Regression tests**

348 In seawater microcosms, the changes in concentrations of cEC from day 0 to day 13 were
349 well correlated with those of coprostanol ($R^2 = 0.92$) and 24-ethylcoprostanol ($R^2 = 0.91$).
350 However the decrease in concentrations of cEC was not correlated to concentrations of
351 epicoprostanol, campestanol and sitostanol ($R^2 < 0.40$) due to their increases between day 6
352 and day 13. The correlation between the decrease of concentrations of cENT and stanols
353 showed a same trend with $R^2 > 0.95$ for coprostanol and 24-ethylcoprostanol and $R^2 < 0.60$ for
354 epicoprostanol, campestanol and sitostanol. In freshwater microcosms, the five stanols were
355 better correlated to the change in concentrations of cEC ($R^2 > 0.85$) than to cENT with R^2
356 ranging from 0.73 (sitostanol) to 0.78 (epicoprostanol).

357

358 **4. DISCUSSION**

359 The objective of this research was to evaluate the persistence of bacterial and chemical
360 markers as compared to the survival of indicator organisms that are measured currently to
361 assess water microbial quality. It is expected that a microbial load added to a water body by a
362 faecal pollution for instance would decline with time due to the effects of several parameters
363 including sunlight, sedimentation, dilution, transport or grazing by biological agents (Barcina

364 et al., 1997; Easton et al., 1999). Although here, conditions of natural water bodies were not
365 fulfilled, since the experiment was performed in a closed environment, a decline was
366 observed for every target: molecular and living organisms. This observation is consistent with
367 other recent microcosm studies in freshwater and seawater (Dick et al., 2010; Walters et al.,
368 2009).

369

370 **4.1. Decay curves**

371 The kinetics of the FIB and microbial markers followed a biphasic curve (Fig. 1). The first
372 phase occurred within 6 days on average. During this phase, a high proportion of the starting
373 inoculum decayed. The remaining proportion persisted in the water column and seemed to be
374 more resistant to decline, since the second phase decline was slow or even null. This was
375 reported before by Easton et al. (1999) who showed, using in-situ chambers that faecal
376 microorganisms did not die-off at a constant rate, and this was only true for the initial decline.
377 Their experiment demonstrated that the die-off rate slowed down as the organism level
378 approached equilibrium with the environment. They found that the initial rapid die-off
379 occurred, generally during the first seven days of the experiment which is consistent with our
380 findings. They proposed two hypotheses to explain this observation (1) the microorganism
381 die-off at a rapid rate until the carrying capacity of the environment is reached, (2)
382 microorganisms would use quorum sensing to regulate their numbers and adapt to their new
383 environment. Although here other parameters than the genetic programming of organisms
384 might have triggered the decline such as sedimentation (Hartz et al., 2008), grazing by
385 protozoa (Bell et al., 2009) or loss of culturability (Barcina et al., 1997), it seems that bacteria
386 can regulate their numbers in a microcosm. Thus, Hellweger et al. (2009) established the
387 biphasic decay kinetic of a pure strain of culturable *E. coli* inoculated in sterile phosphate
388 buffered saline. They observed a decline of the initial inoculum during two days followed by
389 a slight increase of *E. coli* densities. Their experiment established that the resistant fraction

390 was not a population or strain dependent parameter since they used a pure strain. They
391 proposed as possible explanation that this resistant fraction was made of mutants growing on
392 nutrients released by dead cells.

393 In agreement with our results, Bae and Wuertz (2009) demonstrated the biphasic persistence
394 of *Bacteroidales* gene markers and *Enterococcus* 23S rRNA gene from human, cattle and dog
395 faecal samples in seawater. Furthermore, Dick et al. (2010) also observed a decay with a
396 biphasic pattern for cEC and *Bacteroidales* genetic markers from human wastewater in
397 freshwater. They also noted that 99% of the initial inoculum was inactivated during the first
398 phase regardless the microcosm conditions. It is noteworthy that in the mentioned studies,
399 although the type of water, the polluting matrix and the physical conditions differed, the
400 biphasic pattern of every different microbial target was observed. This is consistent with our
401 results as each bacterial target in both treatments followed a biphasic decay trend.
402 Furthermore, it has been reported that a high level of prey would be reduced by predators to
403 an equilibrium density that would ensure the survival of the predators (Marino and Gannon,
404 1991; Menon et al., 2003). It appears that numerous factors are involved in the biphasic decay
405 and the mechanisms responsible for this trend. Results from our study would probably depend
406 on both predation, as the waters were not filtered, and on intrinsic characteristic of the studied
407 bacteria.

408 The persistence pattern of the DNA markers tended to follow the survival pattern of the living
409 organisms. We could then speculate that the DNA we quantified arose from living cells. This
410 is somehow illustrated by the positive second decay rate from the *L. amylovorus* marker
411 (Table 1). This very low rate could be due to sampling or measurement variations or to a
412 multiplication of the marker . However, it was not possible to verify this hypothesis because
413 no medium enables the isolation of *L. amylovorus* from a complex matrix.

414 Stanol decay results were concordant with the evolution of coprostanol in seawater during
415 microcosm experiment performed in darkness at 19 °C (Thoumelin et al., 1990). The increase

416 of the concentration of sitostanol, campestanol and epicoprostanol between day 6 and day 13
417 could be due to the death of living organisms inherited from the pig manure. As a
418 consequence the sterols that constitute those organisms were liberated in the dissolved phase
419 (Marty et al., 1996) and further hydrogenated into sitostanol, campestanol and epicoprostanol
420 (Pratt et al., 2008). This explanation would agree with the observed microbial decay.

421

422 **4.2. FIB survival and markers persistence**

423 In seawater, Pig-2-Bac presented a rapid decay rate close to that of cEC (1.1 and 1.3 d⁻¹,
424 respectively) during the first phase, twice faster than that of cENT. It has been described that
425 protozoa eliminate gram positive bacteria at lower rates than gram negative bacteria (Barcina
426 et al., 1997; Davies et al., 1995). Furthermore, in a recent study Balleste and Blanch (2010)
427 proved that *Bacteroides fragilis* survival was highly hindered by grazing predators in warm
428 conditions in a river. Additionally, Jin et al. (2005) established in a natural slightly salted
429 water storm event experiment that the percentage of *E. coli* attached to suspended particles
430 was 21.8 to 30.4 % compared to 8.3 to 11.5 % for enterococci. However, in salty water, Gram
431 positive bacteria would try to protect themselves from the osmotic pressure by attaching to
432 organic matter (Hartz et al., 2008). In this experiment, turbidity was not measured, but it was
433 clear since we could filtered the water from day 13, that suspended organic matter had settled
434 and thus the water column was poorer in organic matter from that day. As a consequence, the
435 concentration of organic matter might have accounted for in the survival of *E. coli* and Pig-2-
436 Bac marker in seawater. Predation and sedimentation might explain the more rapid decay of
437 Gram negative compared to Gram positive markers in seawater. Another factor that might
438 explain the sensibility of the *Bacteroidales* marker compared to the three other organisms in
439 freshwater, is their sensibility to oxygen. It is well established that *Bacteroidales* cells are
440 negatively affected by increased dissolved oxygen in water (Bae and Wuertz, 2009; Balleste
441 and Blanch, 2010). In freshwater, the *Bacteroidales* marker was the only target not detected

442 until the end of the experiment although, Pig-2-Bac decay constant was not different from
443 cENT decay constants (Table 1), thus the shorter relative persistence time period might also
444 be explained by the higher detection limit of the method. This could be improved by filtering
445 higher volumes of water especially, this would be possible in natural environment normally
446 less concentrated in suspended organic matter, or improving DNA extraction yield.

447 No microcosm studies have yet reported on the persistence of *L. amylovorus*. It is thus
448 interesting to note that the pig-specific *L. amylovorus* marker followed the same trend as
449 cENT until day 55 in seawater and in freshwater, although the detection limit was elevated
450 (112 CFU equivalent/100 mL) compared to that of FIB (1 CFU/100 mL). However, the
451 marker follows cEC evolution only in freshwater as it was inactivated more rapidly in
452 seawater. This is not surprising as it has been demonstrated that unlike *Enterococcus faecalis*
453 or *Lactobacillus casei*, *E. coli* does not harbour resistance mechanisms to high osmotic
454 pressure (Lee et al., 1977). However, as stated before, attachment to organic matter aids the
455 bacteria to resist to this pressure.

456 The progressions of the amount of FIB and of the concentration of stanols followed the same
457 trends except for cEC in seawater. Coprostanol and ethylcoprostanol were correlated with
458 cEC until it was no more detected in seawater. Regression coefficients were higher between
459 the five stanols and cENT than with cEC in seawater. On the other hand in freshwater
460 microcosms the five stanols concentrations were more closely related to the amount of cEC
461 than of cENT. However the regression coefficients were still high *i.e.* $R^2 > 0.70$ and defined a
462 close relationship between the occurrence and concentrations of stanols and FIB.

463

464 **4.3. Effect of the type of water**

465 To determine whether the type of water influenced the persistence of markers in the
466 conditions of the experiment, we compared decay rates observed from both microcosms for
467 each target. Pig-2-Bac was not significantly influenced by the type of water during the first

468 phase maybe due to the presence of suspended particles but salinity or other factors from the
469 seawater accelerated the decay during the second phase (Table 1) when the particles had
470 settled. As a consequence, the marker was detected one more week in freshwater microcosms.
471 These results are somehow different from those from Okabe and Shimazu (2007) who
472 exposed their *Bacteroidales* pig marker Pig-Bac2 to different salinities and observed no real
473 difference in the persistence or decay of the marker to 0, 10, 20 and 30 g/L under dark
474 conditions; however their experiments were performed at 10°C which decreases the decay.
475 The lactobacillus marker was affected by the type of water at the beginning of the experiment,
476 but the remaining proportion of the population was not affected and persisted for the same
477 length of time in freshwater and seawater microcosms. It has been shown that *L. amylovorus*
478 can multiply in NaCl concentration of 5-10 g/L and can survive in concentrations from 20 to
479 40 g/L (Neysens et al., 2003). This new genetic marker, which belongs to a genus
480 phylogenetically close to the *Enterococcus* genus, might be useful to monitor marine waters.
481 The concentrations of stanols after the addition of pig manure were twice to three times
482 higher in the dissolved phase ($< 0.7\mu\text{m}$) of freshwater than of seawater. This observation was
483 probably due to the increased salinity in seawater that induced an aggregation of dissolved
484 macromolecules instead of allowing them to suspend in the dissolved phase as colloids. In
485 spite of slight differences observed on decay curves (Fig. 3), the decay rates calculated for
486 stanols in seawater and freshwater microcosms did not exhibit significant differences (Table
487 1).

488

489 **4.4. MST toolbox validation**

490 Pig-2-Bac and *L. amylovorus* markers were selected to be part of a toolbox to identify sources
491 of faecal pollution in water in our previous study (Gourmelon et al., 2010), we wanted to
492 estimate how their detection is representative of a faecal load. In seawater and freshwater, in
493 the conditions of the study, according to the concentrations in culturable enterococci, the

494 faecal contamination was present until the end of the experiment. *L. amylovorus* followed
495 globally the same trend as cENT and its detection was thus representative of the faecal load
496 during two months. Pig-2-Bac followed also the same trend as cENT but only until day 20
497 and day 27 in seawater and freshwater, respectively. Thus it was no longer detected whilst the
498 faecal contamination was still present. However, although Pig-2-Bac appeared less persistent
499 than *L. amylovorus*, both markers were detected for a long period of time (at least 20 days)
500 that would allow water managers to take necessary actions in cases of important discharges.
501 The development of the MST toolbox in our previous study has highlighted two ratios
502 allowing the discrimination between human, pig and cow faeces. Thus, coprostanol to the sum
503 of coprostanol and 24-ethylcoprostanol (R_1) allows the differentiation between human ($>$
504 0.71), porcine (0.55-0.59) and herbivore ($<$ 0.41) contributions and the second ratio, sitostanol
505 to coprostanol (R_2) exhibits values $>$ 1 for bovine manures and $<$ 0.4 for pig manures and
506 waste water treatment plant effluents (Gourmelon et al., 2010). In seawater and freshwater
507 microcosms, between day 0 and day 13, R_1 exhibited values characteristic of pig manure.
508 However, over the course of the experiment, R_1 remained above the specific values of bovine
509 manure, suggesting that slight changes in R_1 values over time do not lead to misinterpretation
510 of the origin of the contamination. In both microcosms, R_2 was characteristic of pig manure
511 from day 0 to day 6. As a consequence, the combination of stanol ratios R_1 and R_2 can be
512 investigated in order to indicate a faecal contamination from pig manure up to six days from
513 the beginning of the discharge, which allows time to enumerate FIB in order to determine if
514 further analyses are necessary.

515

516 **5. CONCLUSION**

517 This laboratory microcosm study aimed to compare the decay rates of FIB, *Bacteroidales* Pig-
518 2-Bac and *L. amylovorus* pig genetic markers and stanol ratios in both freshwater and
519 seawater inoculated with pig manure.

520 Regardless the microcosm conditions and the target, the persistence or survival profiles of the
521 two genetic markers and of the FIB followed a biphasic curve whereas the five stanols
522 followed a monophasic first order decay kinetic.

523 The persistence of genetic and chemical markers were similar in freshwater and in seawater.
524 According to the values of the ratio R_1 and R_2 , and as the persistence of *L. amylovorus* and
525 Pig-2-Bac *Bacteroidales* marker was relatively close to that of cultivable *E. coli* and
526 enterococci, all these markers can be used to complement *E. coli* and enterococcus detection
527 method to identify a source of pig pollution at least during 6 days when the time point of the
528 contaminating discharge is known. Furthermore, they should prove useful to trace a pig faecal
529 pollution from bathing areas and shellfish farming waters that are sporadically classified as
530 “non satisfactory” in terms of microbial quality.

531

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539

540 **REFERENCES**

541 Bae, S. and Wuertz, S. (2009) Rapid decay of host-specific faecal *Bacteroidales* cells in
542 seawater as measured by quantitative PCR with propidium monoazide. *Water Research*
543 43, 4850-4859.

544 Balleste, E. and Blanch, A. R. (2010) Persistence of Bacteroides Species Populations in a
545 River as Measured by Molecular and Culture Techniques. *Applied and Environmental*
546 *Microbiology* 76, 7608-7616.

547 Balleste, E., Bonjoch, X., Belanche, L.A. and Blanch, A.R. (2010) Molecular Indicators Used
548 in the Development of Predictive Models for Microbial Source Tracking. *Applied and*
549 *Environmental Microbiology* 76, 1789-1795.

550 Barcina, I., Lebaron, P., VivesRego, J. (1997) Survival of allochthonous bacteria in aquatic
551 systems: A biological approach. *FEMS Microbiology Ecology* 23, 1-9.

552 Bell, A., Layton, A. C., Mckay, L., Williams, D., Gentry, R., Sayler, G. S. (2009) Factors
553 Influencing the Persistence of Fecal Bacteroides in Stream Water. *Journal of*
554 *Environmental Quality* 38, 1224-1232.

555 Davies, C. M., Long, J. A. H., Donald, M., Ashbolt, N. J. (1995) Survival of Fecal
556 Microorganisms in Marine and Fresh-Water Sediments. *Applied and Environmental*
557 *Microbiology* 61, 1888-1896.

558 Dick, L.K., Bernhard, A.E., Brodeur, T.J., Domingo, J.W.S., Simpson, J.M., Walters, S.P. and
559 Field, K.G. (2005) Host distributions of uncultivated faecal Bacteroidales bacteria
560 reveal genetic markers for faecal source identification. *Applied and Environmental*
561 *Microbiology* 71, 3184-3191.

562 Dick, L.K., Stelzer, E.A., Bertke, E.E., Fong, D.L. and Stoeckel, D.M. (2010) Relative Decay
563 of Bacteroidales Microbial Source Tracking Markers and Cultivated Escherichia coli in
564 Freshwater Microcosms. *Applied and Environmental Microbiology* 76, 3255-3262.

565 Easton, J. H., Lalor, M., Gauthier, J. J., Pitt, R., Newman, D. E., Meyland, S. (1999)
566 Determination of Survival Rates for Selected Bacterial and Protozoan Pathogens from
567 Wet Weather Discharges. WEFTEC 1999. Water Environment Federation 72nd Annual
568 Conference and Exposition. New Orleans.

569 Field, K. G. and Samadpour, M. (2007) Fecal source tracking, the indicator paradigm, and
570 managing water quality. *Water Research* 41, 3517-38.

571 Gourmelon, M., Caprais, M.P., Mieszkin, S., Marti, R., Wéry, N., Jardé, E., Derrien, M.,
572 Jadas-Hecart, A., Communal, P.Y., Jaffrezic, A. and Pourcher, A.M. (2010)
573 Development of microbial and chemical MST tools to identify the origin of the faecal
574 pollution in bathing and shellfish harvesting waters in France. *Water Research* 44,
575 4812-4824.

576 Hartz, A., Cuvelier, M., Nowosielski, K., Bonilla, T. D., Green, M., Esiobu, N.,
577 McCorquodale, D. S., Rogerson, A. (2008) Survival potential of *Escherichia coli* and
578 enterococci in subtropical beach sand: Implications for water quality managers. *Journal*
579 *of Environmental Quality* 37, 898-905.

580 Hellweger, F.L., Bucci, V., Litman, M.R., Gu, A.Z. and Onnis-Hayden, A. (2009) Biphasic
581 Decay Kinetics of Faecal Bacteria in Surface Water Not a Density Effect. *Journal of*
582 *Environmental Engineering-Asce* 135, 372-376.

583 Jarde, E., Gruau, G. and Jaffrezic, A. (2009) Tracing and Quantifying Sources of Fatty Acids
584 and Steroids in Amended Cultivated Soils. *Journal of Agricultural and Food Chemistry*
585 57, 6950-6956.

586 Jeanneau, L., Jarde, E., Gruau, G. (2011) Influence of salinity and natural organic matter on
587 the solid phase extraction of sterols and stanols: Application to the determination of the
588 human sterol fingerprint in aqueous matrices. *Journal of Chromatography A* 1218, 2513-
589 2520.

590 Jin, G., Jeng, H. W., Bradford, H., Englande, A. J. (2005) Comparison of *E. coli*, Enterococci,
591 and fecal coliform as indicators for brackish water quality assessment (vol 76, pg 245,
592 2004). *Water Environment. Research.* 77, 433-433.

593 Konstantinov, S.R., Smidt, H. and de Vos, W.M. (2005) Representational difference analysis
594 and real-time PCR for strain-specific quantification of *Lactobacillus sobrius* sp nov.
595 *Applied and Environmental Microbiology* 71, 7578-7581.

596 Lee, D.U., Heinz, V. and Knorr, D. (2001) Biphasic inactivation kinetics of *Escherichia coli*
597 in liquid whole egg by high hydrostatic pressure treatments. *Biotechnology progress* 17,
598 1020-1025.

599 Lee, S.K., Calcott, P.H. and MacLeod, R.A. (1977) Relationship of cytochrome content to the
600 sensitivity of bacteria to NaCl on freezing and thawing. *Canadian journal of*
601 *microbiology* 23, 413-419.

602 Leeming, R., Ball, A., Ashbolt, N. and Nichols, P. (1996) Using faecal sterols from humans
603 and animals to distinguish faecal pollution in receiving waters. *Water Research* 30,
604 2893-2900.

605 Marino, R. P. and Gannon, J. J. (1991) Survival of faecal coliforms and faecal streptococci in
606 storm drain sediment. *Water Research* 25, 1089-1098.

607 Marti, R., Dabert, P., Ziebal, C. and Pourcher, A.M. (2010) Evaluation of *Lactobacillus*
608 *sobrius*/*L. amylovorus* as a New Microbial Marker of Pig Manure. *Applied and*
609 *Environmental Microbiology* 76, 1456-1461.

610 Marty, Y., Quemeneur, M., Aminot, A., LeCorre, P. (1996) Laboratory study on degradation
611 of fatty acids and sterols from urban wastes in seawater. *Water Research* 30, 1127-1136.

612 Menon, P., Billen, G., Servais, P. (2003) Mortality rates of autochthonous and fecal bacteria
613 in natural aquatic ecosystems. *Water Research*. 37, 4151-4158.

614 Mieszkin, S., Furet, J.P., Corthier, G. and Gourmelon, M. (2009) Estimation of Pig Faecal
615 Contamination in a River Catchment by Real-Time PCR Using Two Pig-Specific
616 *Bacteroidales* 16S rRNA Genetic Markers. *Applied and Environmental Microbiology*
617 75, 3045-3054.

618 Neysens, P., Messens, W. and De Vuyst, L. (2003) Effect of sodium chloride on growth and
619 bacteriocin production by *Lactobacillus amylovorus* DCE 471. *International Journal of*
620 *Food Microbiology* 88, 29-39.

621 Okabe, S. and Shimazu, Y. (2007) Persistence of host-specific *Bacteroides-Prevotella* 16S
622 rRNA genetic markers in environmental waters: effects of temperature and salinity.
623 *Applied Microbiology and Biotechnology* 76, 935-944.

624 Pratt, C., Warnken, J., Leeming, R., Arthur, J. M., Grice, D. I. (2008) Degradation and
625 responses of coprostanol and selected sterol biomarkers in sediments to a simulated
626 major sewage pollution event: A microcosm experiment under sub-tropical estuarine
627 conditions. *Organic Geochemistry* 39, 353-369.

628 Shah, V.G., Dunstan, R.H., Geary, P.M., Coombes, P., Roberts, M.K. and Von Nagy-
629 Felsobuki, E. (2007) Evaluating potential applications of faecal sterols in distinguishing
630 sources of faecal contamination from mixed faecal samples. *Water Research* 41, 3691-
631 3700.

632 Soller, J.A., Schoen, M.E., Bartrand, T., Ravenscroft, J.E. and Ashbolt, N.J. (2010) Estimated
633 human health risks from exposure to recreational waters impacted by human and non-
634 human sources of faecal contamination. *Water Research* 44, 4674-91.

635 Thoumelin, G., Marty, Y., Lecorre, P. and Aminot, A. (1990) Laboratory Investigation of the
636 Degradation of Organic-Matter in Estuarine and Coastal Waters - Sterols Variations.
637 *Oceanologica Acta* 13, 53-60.

638 Thurston-Enriquez, J.A., Gilley, J.E. and Eghball, B. (2005) Microbial quality of runoff
639 following land application of cattle manure and swine slurry. *Journal of Water and*
640 *Health* 3, 157-171.

641 Tyagi, P., Edwards, D. R., Coyne, M. S. (2009) Fecal Sterol and Bile Acid Biomarkers:
642 Runoff Concentrations in Animal Waste-Amended Pastures. *Water Air and Soil*
643 *Pollution* 198, 45-54.

- 644 Ufnar, J.A., Ufnar, D.F., Wang, S.Y. and Ellender, R.D. (2007) Development of a swine-
645 specific faecal pollution marker based on host differences in methanogen *mcrA* genes.
646 *Applied and Environmental Microbiology* 73, 5209-5217.
- 647 Walters, S.P., Yamahara, K.M. and Boehm, A.B. (2009) Persistence of nucleic acid markers
648 of health-relevant organisms in seawater microcosms: Implications for their use in
649 assessing risk in recreational waters. *Water Research* 43, 4929-4939.

650 **Figures Captions:**

651

652 Fig. 1 – Decay curves of pig specific microbial markers in water microcosms seeded with
653 liquid pig manure (1:100 dilution), (A) seawater, (B) freshwater. Conditions: dark, O₂
654 saturation, around 18 °C. The uncertainties represent standard errors. Limit of quantification
655 was 1 CFU/100 mL for FIB, 1250 gene copies/100 mL for Pig-2-Bac and 112 CFU
656 equivalent/100 mL for *L. amylovorus*.

657

658 Fig. 2 - Modelisation of the mean concentration of cENT (A) by Cerf biphasic decay model
659 ($R^2 = 0.97$) and of 24-ethylcoprostanol (B) by Chick first order decay model ($R^2 = 0.98$) from
660 the seawater microcosms

661

662 Fig. 3 - Decay curves of stanols in seawater (A) and freshwater (B) microcosms. The
663 uncertainties represent standard errors. The scale of Y-axes are different

664

665 Fig. 4 - Evolution of stanols ratios R_1 (A; coprostanol/coprostanol+24-ethylcoprostanol) and
666 R_2 (B; sitostanol/coprostanol). Grey areas correspond to the range of values characteristic of
667 pig manure. SWP: seawater, FWP: freshwater.

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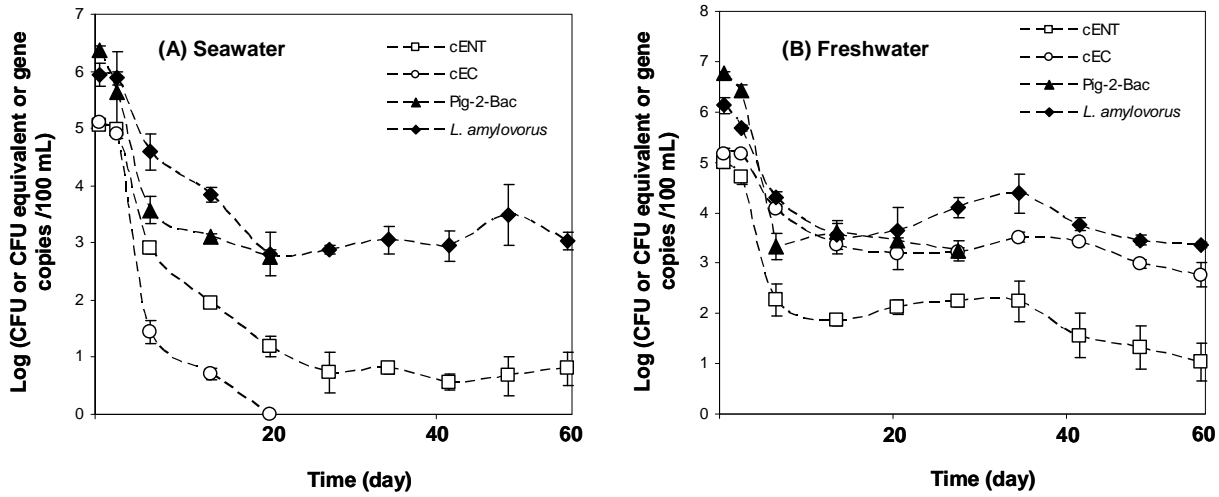
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680 Fig 1

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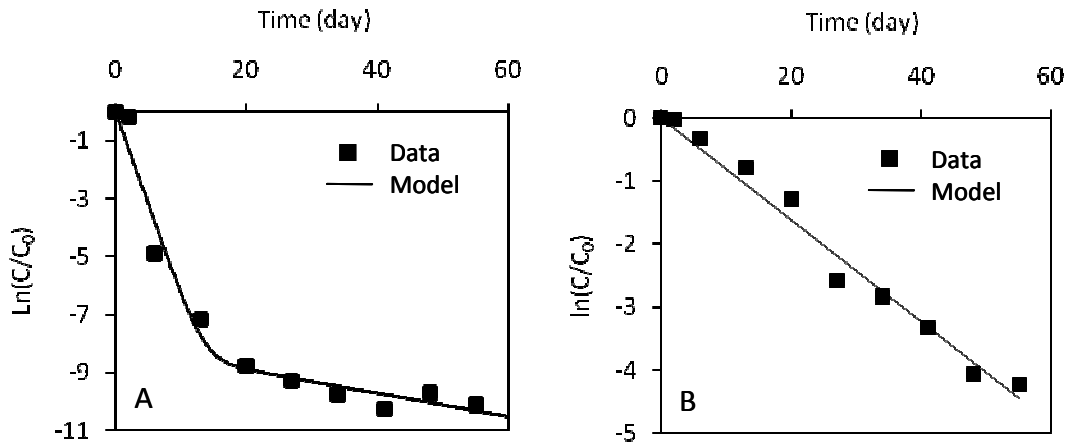
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692 Fig 2

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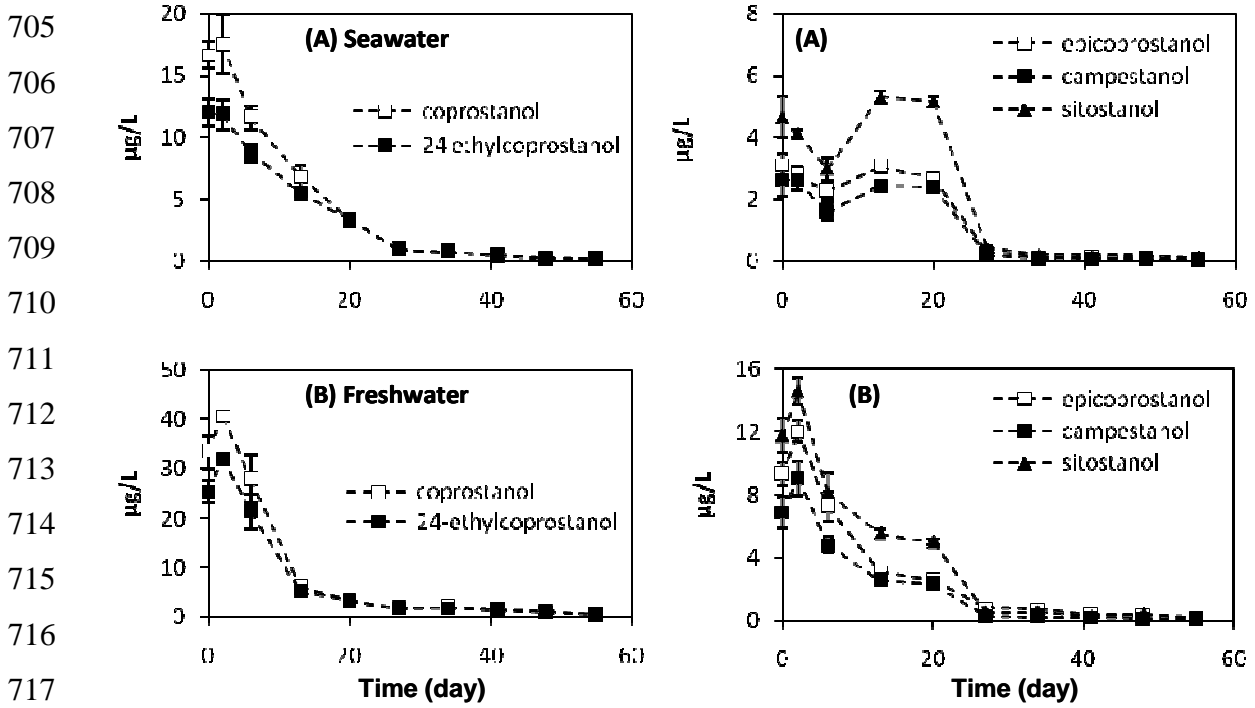


Fig 3

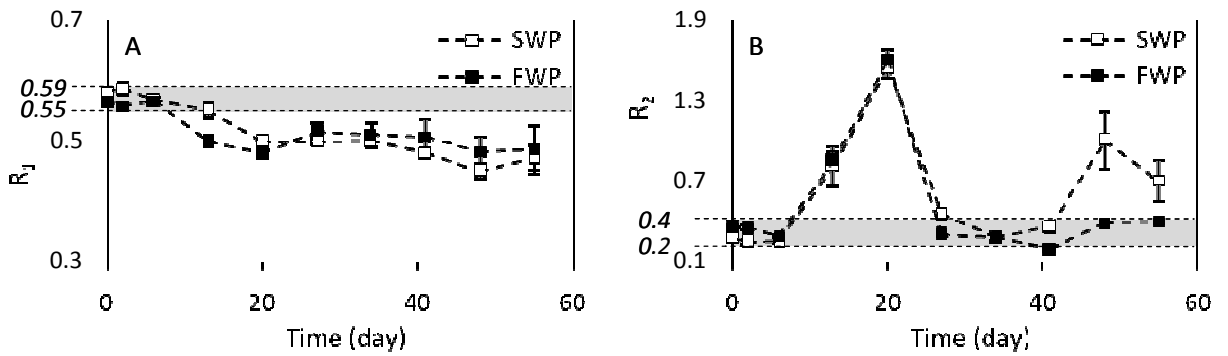


Fig4

725 Table 1 - Decay rates and T90 of FIB and pig specific microbial and chemical markers in water microcosms seeded with liquid pig manure (1:100
 726 dilution). p-values represent comparison between decay rates in seawater and freshwater. Conditions: dark, O₂ saturation, around 18 °C.
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Marker	Seawater					Freshwater					p-value k1	p-value k2
	k1 (d ⁻¹)	k2 (d ⁻¹)	f (%)	T ₉₀ (d)	R ²	k1 (d ⁻¹)	k2 (d ⁻¹)	f (%)	T ₉₀ (d)	R ²		
<i>cE. coli</i>	1.291	0.388	99.7	1.8	0.96	0.428	-0.021	97.8	5.4	0.94	0.0006	0.0004
cENT	0.605	0.016	99.9	3.8	0.97	1.015	-0.048	99.7	2.3	0.94	0.0377	0.6714
Pig-2-Bac	1.100	0.075	99.9	2.1	0.99	1.247	-0.001	99.9	1.9	0.95	0.7488	0.0412
<i>L. amylovorus</i>	0.406	-0.031	99.9	5.7	0.97	0.737	-0.014	99.4	3.1	0.94	0.0295	0.3089
Coprostanol	0,092			23.7	0.98	0,082			29.5	0.92	0,1000	
Ethylcoprostanol	0,086			26.2	0.98	0,075			32.0	0.93	0,2000	
Epicoprostanol	0,078			25.0	0.91	0,075			29.1	0.98	0,3000	
Campestanol	0,078			25.6	0.88	0,083			25.6	0.93	1,0000	
Sitostanol	0,072			32,0	0,82	0,079			29,1	0,91	0,3000	

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