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Mg/Ca in fossil oyster shells as palaeotemperature proxy, an example from the Palaeogene of Central Asia.

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Abstract

Fossil oyster shells are well-suited to provide palaeotemperature proxies from geologic to seasonal timescales due to their ubiquitous occurrence from Triassic to Quaternary sediments, the seasonal nature of their shell growth and their strong resistance to post-mortem alteration. However, the common use to translate calcitic oxygen isotopes into palaeotemperatures is challenged by uncertainties in accounting for past seawater δ^{18} O, especially in shallow coastal environment where oysters calcify. In principle, the Mg/Ca ratio in ovster shells can provide an alternative palaeothermometer. Several studies provided temperature calibrations for this potential proxy based on modern species, nevertheless their application to palaeo-studies remains hitherto unexplored. Here, we show that past temperature variability in seawater can be obtained from Mg/Ca analyses from selected oyster fossil species and specimens. High-resolution Mg/Ca profiles, combined with δ^{18} O, were obtained along 41 fossil oyster shells of seven different species from the Palaeogene Proto-Paratethys sea (Central Asia) found in similar as well as different depositional age and environments providing comparison. Suitable Mg/Ca profiles, defined by continuous cyclicity and reproducibility within one shell, are found to be consistent for specimens of the same species but differ systematically between species, implying a dominant species-specific effect on the Mg/Ca incorporation. Two species studied here (Ostrea (Turkostrea) strictiplicata and Sokolowia buhsii) provide an excellent proxy

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for palaeoclimate reconstruction from China to Europe in Palaeogene marine sediments. More generally, the protocol developed here can be applied to identify other fossil oyster species suitable for palaeoclimate reconstructions.

Keywords: palaeoclimate, oyster, Mg/Ca, sclerochronology, Palaeogene, Central Asia

1. Introduction

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The ability to track quantitatively climate seasonality patterns through the geological record is 2 key to understand past, present and future climate processes (Felis et al., 2004; Licht et al., 2014). However, most palaeoclimate proxies yield only qualitative assessments of seasonality or insufficient resolution averaging across thousands of years (Eldrett et al., 2007; Evans et al., 2013). In contrast, bivalve shells have long been recognized to potentially represent excellent high-6 resolution (i.e. infra-annual) palaeoclimate archives (e.g. Hudson et al., 1976; Richardson, 2001). The shell of bivalves is formed by incremental growth resulting in a record that can be sampled at an intra-annual resolution (Stenzel, 1971; Kirby et al., 1998). Moreover, they live over a large range of climate regimes and habitats (Surge et al., 2001; Mouchi et al., 2013). In addition, their generally large and thick calcitic shells promote good preservation of fossils such that oysters represent one 11 the most abundant macro-fossil groups readily available in the sedimentary records (Stenzel, 1971). 12 When biomineralizing, bivalve species have been shown to build their shell in isotopic equilibrium 13 with sea water and therefore, its stable oxygen isotope ratios (δ^{18} O) reflect seawater temperatures in which the organisms formed its shell (Killingley and Berger, 1979; Wefer and Berger, 1991; Ivany 15 et al., 2000). However, carbonate δ^{18} O also varies with the stable oxygen isotopic composition of 16 seawater ($\delta^{18}O_{sw}$, e.g. Anderson and Arthur, 1983), such that measured isotope ratios corresponds to a combination of past seawater temperature and isotope composition. Estimating seawater 18 isotopic composition and its intra-annual variability in the geologic past remains a major challenge 19 to isolate the temperature signal from shell oxygen isotopes. In principle, seawater oxygen isotope composition can be resolved using a second independent temperature proxy, not primarily depending 21 on salinity or seawater oxygen isotope composition. Such a proxy may be available from minor and 22

trace elements in biominerals, particularly the Mg/Ca and Sr/Ca ratios in carbonates are known

to reflect past seawater temperatures in foraminifera (e.g. Nürnberg et al., 1996; Lear et al., 2000)

and corals (Lough, 2010), respectively. Based on Mg/Ca and Sr/Ca, these temperatures can be

combined with carbonate $\delta^{18}O$ to estimate past seawater $\delta^{18}O$, which in turn reflects seawater salinity (Elderfield and Ganssen, 2000; Lear et al., 2000; Bougeois et al., 2014).

Accurate minor/trace element-temperature calibration for bivalve shells have long been unavail-28 able by a lack of appropriate controlled growth studies (Dodd, 1965). More recently, many studies 29 succeeded in relating seawater temperature to bivalve calcitic Mg/Ca ratio in mussels (Klein et al., 30 1996; Vander Putten et al., 2000; Freitas et al., 2009; Wanamaker et al., 2008), scallop (Freitas 31 et al., 2006, 2009, 2012), fan mussels (Freitas et al., 2005), and ovsters (Surge and Lohmann, 2008; 32 Mouchi et al., 2013). These studies revealed relatively large inter-specific differences in temperature 33 sensitivity (see Table 1 for different calibrations). In addition, environmental conditions exert a 34 strong impact on bivalve Mg/Ca even within one species. For example, calibrations differ for M. 35 edulis when calcifying in estuarine, culture or natural brackish waters (eq. 6, 7 and 8 in Table 1), 36 suggesting that variables other than temperature additionally determine bivalve calcite chemistry 37 (Freitas et al., 2005, 2006, 2009, 2012). These suggested variables include metabolism (Rosenberg 38 and Hughes, 1991; Vander Putten et al., 2000), intra-shell variability (Freitas et al., 2009), salinity 39 and growth rate (Wanamaker et al., 2008) on Mytilus edulis. Studies also revealed an ontogeny-40 related effect in *Pinna nobilis* (Freitas et al., 2005) and an impact of incorporated organic matter 41 (mainly conchiolin) on bivalve aragonite shell chemistry in Arctica islandica (Schöne et al., 2010). 42 These effects are negligible in ovster shells in which incorporation of organic compounds into the 43 carbonate crystal lattice, especially in foliated calcite, is much lower than in other bivalve (e.g. mussels and scallops; Stenzel, 1971). Furthermore, the oyster's conchiolin is relatively easily degraded by bacterial activity, so that even the organic content of older parts of the shell has already 46 decreased during the life of the animal and is absent in fossil oysters (Stenzel, 1971). Recent studies 47 also showed that impacts of salinity, ontogeny and growth rate are modest compared to that of 48 temperature on Mg incorporation into the foliated oyster calcite shell (Surge and Lohmann, 2008; 49 Mouchi et al., 2013). Furthermore, duplicate of Mg/Ca ratio analyses in the ligamental area showed 50 that intra-shell small scale variability for magnesium is negligible (Bougeois et al., 2014). 51

Although these considerations clearly designate oyster shells as prime targets for palaeoclimate reconstructions using Mg/Ca, such analyses have yet to be tested using a thorough analytical protocol. We previously conducted a pilot study on a single fossil oyster shell (*Sokolowia buhsii*) from the Eocene Proto-Paratethys sea in Central Asia suggesting that its Mg/Ca may be used to recon-

struct palaeotemperatures (Bougeois et al., 2014). However, to constrain the reliability of Mg/Ca as a palaeothermometer for the geological past and to quantify inter-specimen variability in element composition, additional investigations on specimens and species from a range of environments and ages are necessary. Here, we extend the approach to various Palaeogene oyster species including different environmental and stratigraphic contexts well-constrained by parallel studies of the western Proto-Paratethys sea (Bosboom et al., 2014a,b,c) (Bosboom et al., 2014a,b,c). We aim to determine whether Mg/Ca ratios yield reliable and reproducible environmental intra-annual proxy data in fossil oyster shells from various species and environments and thereby the reliability of the Mg/Ca-temperature calibration established on modern oysters when applied to fossil specimens.

65 2. Context of sampling

66 2.1. Environmental and stratigraphic setting

During Palaeogene times, a large shallow epicontinental sea belonging to the Tethyan Realm 67 covered the Eurasian continent from the Mediterranean Tethys (west side) to the Tarim Basin (west-68 ern China, east side) (Dercourt et al., 1993; Burtman, 2000). In the Tarim Basin (Xinjiang, China), 69 the Afghan-Tajik depression (Afghanistan-Tajikistan), the Fergana basin and the Alai Valley (Kyr-70 gyzstan) (Figure 1a, b), regional regression-transgression sequences have been established based on 71 inter- and intra- basin stratigraphic correlations of the deposits with distinct lithological facies and fossil assemblages including bivalves, dinoflagellate cysts, foraminifera and ostracods (see Bosboom 73 et al., 2014a,b,c, and references therein). From the Cenomanian to the final sea-retreat dated late 74 Bartonian-early Priabonian, five major second-order marine incursions have been recognized in the 75 sedimentary record in Central Asia. The marine intervals are dominated by bioclastic wackestones to grainstones, evaporites, clay and mudstones containing ostracods, gastropods, bryozoa, serpulids, 77 echinoids, foraminiferas, algae, fish scales and molluscs. Bivalve calcitic shells such as Ostreidae and 78 Pectinidae mainly remained in sedimentary deposits, whereas aragonite molluscs were leached and not preserved. Sedimentary facies and fossils assemblages are characteristic of shallow coastal envi-80 ronments between offshore to coastal plains, and typical of a carbonate-rich neritic ramp (Manceau 81 et al., 2014).. Continental intervals between marine incursions contain fine to middle grained detrital 82 rocks, red clay to siltstones and evaporitic deposits characterised by nodular and massive gypsum. 83 These deposits are indicative of flood plains, alluvial plains and playa environments. Sedimentary

deposits indicate alternation between more humid and dry periods, typical of semi-arid climates with a relatively strong seasonal contrast (Manceau et al., 2014).

87 2.2. Oyster sampling, age and determination

Mollusc macrofauna taxonomic identification largely follows Lan and Wei (1995), Lan (1997) 88 and Bosboom et al. (2014a,c), representing the most recent revision of regional systematic litera-89 ture. Fossil ages were estimated using mollusc biostratigraphy (Lan and Wei, 1995) and adjusted for 90 the two last marine incursions from recent studies combining bio- and magnetostratigraphy in this 91 region (Bosboom et al., 2014a,b) (Figure 1c). Specimens from the oyster species Ostrea bellovacina 92 (Late palaeocene, Thanetian), Flemingostrea hemiglobosa (Early Eocene, Ypresian), Sokolowia orientalis (Middle Eocene, Early Lutetian), Ostrea (Turkostrea) strictiplicata (Middle Eocene, Early 94 to Middle Lutetian), Sokolowia buhsii (Middle Eocene, Middle to Late Lutetian), Platygena asiatica 95 (Middle Eocene, Late Bartonian) and Ferganea bashibulakeensis (Middle Eocene, Early Priabonian) were extracted from marine sediment strata in Central Asia during field excursions in the summers 97 of 2007, 2010, 2011, 2012 and 2013 (Figure 1). All taxa belong to the Superfamily Ostreaoidea and 98 the Families Ostreidae or Flemingostreidae (Carter et al., 2011). Specimens were selected according 99 to the following criteria: (1) good preservation of the shell, particularly of the ligamental area, 100 (2) sufficiently large ligamental surface to contain growth bands spanning more than ten years and 101 providing a high resolution, (3) specimens fossilized in living position attesting of the living envi-102 ronment. Field sedimentological analyses yield characterisation of the depositional environmental 103 for each species (Figure 2, Manceau et al., 2014). 104

3. Material and methods

To determine the applicability of Mg/Ca as a temperature proxy we have combined a number of analyses on specimens from different species and representative of a range of environments. To detect potential diagenetic alteration and identify the presence of seasonal banding, shells were first analysed with cathodoluminescence microscopy. Trace element analyses were then performed at high resolution yielding Mg/Ca variations with varying reliability criteria defined below. Finally, to provide a reference for comparing and assessing the Mg/Ca results, stable isotope analyses were also performed at high incremental resolution and/or on bulk samples (Table 2).

3.1. Shell description and sample preparation

Oyster shells grow by incremental deposition of calcium carbonate (mainly calcite), resulting in alternating dark and light bands corresponding to the colder and warmer seasons respectively (Kirby et al., 1998). The foliated calcite forming the ligamental area (or umbo) is very dense and resistant to post-mortem alteration, resulting in conservation of the primary environmental signal. For this reason, we focused our chemical analyses to the ligamental area, following practice from a number of previous studies (Kirby et al., 1998; Surge and Lohmann, 2008; Lartaud et al., 2010b; Goodwin et al., 2012; Mouchi et al., 2013; Bougeois et al., 2014).

After isolation from the sediment and thorough cleaning of the shell's surface by brushing off 121 loose material and rinsing with deionized water, specimens were prepared for sclerochronological and 122 geochemical analysis by cutting 0.5 cm-thick slices along the maximum growth axis of the left valve 123 in the middle of the resilifer (Figure 3). These sections were subsequently polished and cleaned in 124 an ultrasonic bath with deionized water for ten minutes and dried overnight. Radial sections reveal 125 numerous growth lines providing a continuous record along the oyster's lifetime (Figure 3). Shell 126 slices were then sectioned into slices of maximum 2.5 cm width and 5.0 cm length to fit the sample 127 holder for geochemical analyses. 128

3.2. Cathodoluminescence analyses

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Shell preservation and potential impact of diagenesis are commonly detected in bivalve shells 130 using cathodoluminescence (CL) microscopy (e.g. Langlet et al., 2006). In calcite, Mn²⁺ is the main 131 luminescence activator, causing emission of yellow to orange light of which the intensity is positively 132 correlated with Mn concentration (El Ali et al., 1993; De Rafélis et al., 2000; Habermann, 2002; 133 Langlet et al., 2006). As Mn is also preferentially incorporated in the oyster shell calcite during 134 summer months (Langlet et al., 2006; Lartaud et al., 2010b), this results in high luminescence 135 intensities during summer and conversely relatively low luminescence during winter. Thus variations 136 of the CL intensity following the shell's growth axis in each slab and can thus be used as a rapid 137 and non-invasive method to identify the presence of seasonal banding (Bougeois et al., 2014). CL 138 analyses were performed at the Université Pierre et Marie Curie (Paris, France) on a cold cathode 130 device (Cathodyne OPEA) coupled to an optical microscope and a digital camera. 140

3.3. Trace element analyses

Trace element composition was determined by Laser Ablation-Inductively Coupled-Plasma-Mass 142 Spectrometry (LA-ICP-MS), with a Geolas 200Q Excimer 193 nm laser coupled to a sector field 143 ICP-MS (Element2, Thermo Scientific) at the Utrecht University (The Netherlands). The sectioned oyster shells were ablated at lower energy density ($\sim 1 \text{ J.cm}^{-2}$) by moving the sample in x, y, and z 145 axis at a constant speed underneath the laser beam in a He environment. The path was programmed 146 so that subsequent circular ablation spots (120 µm in diameter) overlapped slightly and resulted 147 in a resolution of one data point every ~ 7 µm leading in more than 100 measurements for each 148 growth band. All transects were ablated perpendicularly to the growth axis to monitor consistency 149 of obtained profiles. To test data consistency and quantify intra-shell variability inside the umbo, 150 each transect was duplicated by a second parallel transect spaced at approximately 0.5 to 1 mm 151 from the first one. Masses monitored by the ICP-MS included 24 Mg, 26 Mg, 43 Ca, 44 Ca, 55 Mn, 88 Sr 152 and ¹³⁸Ba, using the international standard NIST SRM610 (using element concentrations reported 153 in Jochum et al., 2011) and ⁴³Ca as an internal standard (assuming 40% wt in CaCO₃). The glass standard was ablated between measurements of each oyster at a higher energy density ($\sim 5 \text{ J.cm}^{-2}$). 155 Calibration of element/calcium ratios in calcium carbonate samples was obtained with the NIST 156 glass standard yielding accurate values for many elements when using a 193 nm laser as indicated 157 by similar fractionation factors obtained when using glass and carbonate standard material, despite 158 differences in ablation characteristics for these materials (Hathorne et al., 2008). Here, we mainly 159 focus on the obtained patterns in Mg/Ca (for more details about the LA-ICP-MS protocol, see 160 Bougeois et al., 2014). To account for small scale fluctuations in the Mg/Ca ratio, corresponding 161 values from the two parallel transects were averaged so that a single data-point was obtained for 162 each incremental position. On these data-points, a moving average using a Bartlett window was 163 then run through data from every transect as previously described in (Bougeois et al., 2014).

165 3.4. Stable isotopes analyses

Stable oxygen isotopes were analysed (1) at high incremental resolution aiming to resolve seasonal variability and (2) on bulk samples aiming to average the isotopic record throughout the oyster's life. Bulk carbon isotopes (δ^{13} C) were also analysed to assess potential environmental effects on the shell's chemistry. To collect material for these analyses, a high-precision, computerdriven Micromill (New Wave Research) attached to an x, y and z stage was used to follow digitized

milling path positions. Bulk analyses were performed by milling one continuous line following laser 171 transects across the ligamental area. For incremental analyses, parallel paths were milled every 172 100-120 µm following the growth bands (Figure 3). At least 30 to 50 µg of calcitic shell powder 173 was collected for each milled sample and analysed for oxygen and carbon stable isotopes (δ^{18} O and 174 δ¹³C in %₀ VPDB) using a KIEL-III device coupled online to a Finnigan MAT-253 mass spectrom-175 eter at Utrecht University for incrementally resolved data of the specimens AT11-O04, KZ07-O01, 176 KY10-O01. Material from all other specimens and for bulk analyses were analysed using a KIEL-IV 177 carbonate device at the Université Pierre et Marie Curie (UPMC, Paris, France). The international 178 standard NBS-19 and an in-house standard (Naxos marble in Utrecht University and Marceau in 179 UPMC) were used for calibration of all sample sets. For both instruments, long-term analytical 180 precision was better than 0.08% for $\delta^{18}O$ and better than 0.05% for $\delta^{13}C$. 181

For the oysters analysed at the Utrecht University the entire length of each shell specimen was 182 milled for incremental study at high resolution (one sample every 120 μm) totalling ~130 samples 183 per shell for ~20 cycles (or years). Results from this first batch of samples indicate that a slightly 184 lower resolution was sufficient except for intervals with much lower growth rates (and thus thinner 185 bands). This justified a drilling resolution of only 5 to 8 annual cycles into the remaining specimens 186 that all displayed higher growth rates and thereby enabled increasing the total number of specimens 187 analysed. At a resolution of one micro-milled sample every 100 to 200 µm (samples with sufficiently 188 high growth rates were analysed with only one micro-sample every 200 µm) between 42 and 82 189 samples were thus analysed per specimen. This enabled the incremental stable isotope analyses 190 for 20 carefully-chosen shells (out of the 41 shells initially analysed for trace element composition) 191 while bulk stable isotope analyses was performed on 40 shells, to obtain at least one stable isotopic 192 value for each specimen. 193

194 4. Analytical results

195 4.1. Cathodoluminescence results

In total, 38 oyster shells were analysed with cathodoluminescence microscopy. In general, clear and consistent banding in CL intensities show that diagenetic effects potentially affecting the analysed oyster shells were minimal, indicating a good preservation of the carbonaceous skeleton. A cyclic pattern is well defined for the species S. buhsii, O. (T.) strictiplicata and O. bellovacina with

alternations between intense luminescence for the light summer bands and lower luminescence intensities for the dark winter growth bands. The luminescence for *F. hemiglobosa* is very weak and the luminescence for *F. bashibulakeensis* is inverted with thinnest layers corresponding to winter growth deceleration (Richardson et al., 1993; Kirby, 2001) with relatively bright luminescence (Figure 3).

204 4.2. Trace element results

- A total of 41 shells were analysed for trace elements. Mg/Ca values range between 1.06 and 14.33 mmol/mol. Average Mg/Ca values for a single specimen varies from 2.74 to 9.75 mmol/mol (Figures 6 7, 8 and 9e-h).
- The intra-shell patterns in Mg/Ca vary between specimens and we therefore evaluated the quality of the results using two properties: (1) the small-scale intra-shell variability, which is the difference between patterns in Mg/Ca from the two parallel transects; (2) the cyclicity in Mg/Ca reflecting whether a primary seasonal signal has been recorded. According to these criteria we grouped the Mg/Ca results into three categories (Table 2):
- suitable (continuously cyclic and consistent between parallel transects, Figure 6)
- partly suitable (only some parts are cyclic and consistent between transects, Figure 7)
- unsuitable (non cyclic and/or not consistent between parallel transects, Figure 8).
- For example Mg/Ca ratios from specimen KZ12-O05.1 (Figure 8) display a cyclic pattern but 216 the absolute values of the two transects are clearly different and therefore this sample is qualified 217 as unsuitable. Transects from the specimen MS12-O06.5 (Figure 7) display very consistent values 218 from 7 to 19 cm but not from 0 to 7 cm, therefore it is qualified as partly suitable. These reliability 219 assessments based on elemental results provide a useful criterion for selecting promising specimen 220 before engaging into further isotopic analyses. Unsuitable specimens were discarded altogether 221 from further isotopic analyses. In suitable and partly suitable specimens, well-defined continuous 222 parts were selected for further isotopic analyses. Of all shells analysed (n=41), 12 were considered 223 suitable, 13 partly suitable and 16 unsuitable (Table 2). 224

225 4.3. Stable isotopes results

All isotopes values obtained (bulk sample analyses and high-resolution incremental analyses) range between -5.45% and -0.57% for δ^{18} O and between -0.74% and 2.53% for δ^{13} C (Figure

5). For each sample, the average value derived from averaging all incremental analyses compares well with values from the bulk sample analyses (Figure 9a-d and Supplementary data). Specimens retrieved from the same stratigraphic bed usually exhibit similar values - standard deviation within 0.3% for δ^{13} C and 0.4% for δ^{18} O- indicating homogeneous isotope ratios in oyster shells living at the same time and in a similar environment.

The well-resolved cyclicity observed in the oyster's isotopes confirms that sampling resolution (every 100-200 μm; depending on growth rates) was sufficiently high to clearly reveal intra-annual environmental variations with regular cyclicity. However, on two samples with particularly low growth rates in the oldest part of the shell (AT11-O04 and KY10-O01, Figure 5) the expression of seasonality is poor. This most likely results from a sampling bias, mixing material between closely spaced adjacent growth bands rather than an ontogenetic effect or a change in environmental variations or as previously shown in (Bougeois et al., 2014). We therefore discarded these parts of the shell record from further consideration.

5. Discussion

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In light of the obtained analytical results, we discuss below (1) the reliability of Mg/Ca records in the various shell species analysed (2) species and environmental effects and (3) the validity of proposed calibrations of the Mg/Ca as a proxy for temperature for various species and depositional environments.

246 5.1. Reliability of Mg/Ca profiles for different species

It appears that reliability of Mg/Ca records varies between species and the species-specific patterns in Mg/Ca are therefore discussed separately.

- O. (T.) strictiplicata and S. buhsii mostly yield suitable and some partly suitable Mg/Ca data (Figures 6 and 7, Table 2). Variations are cyclic and correlate well with the other geochemical proxy δ¹⁸O. No clear ontogenetic effect can be recognized for the Mg incorporation in oyster shells, and CL indicates that diagenesis did not affect the shells analysed here. Furthermore, for specimens with both bulk and incremental stable isotope data, the bulk values compare well with the average of all the incremental δ¹⁸O values within 0.4±0.2‰ (see Figure 9ad). This underlines the homogeneity of the isotopic composition within the ligamental area

and further excludes that ontogeny has a large effect on oyster stable isotope ratios. The stable isotope values display a well-defined cyclicity with low values coinciding with light bands and high δ^{18} O corresponding to darker bands (Figures 4 and 5) revealed by CL. This is in agreement with previous observations showing a seasonal isotopic signal corresponding to variation of the CL intensity (Lartaud et al., 2010b; Bougeois et al., 2014). Similarly, stable isotope values are correlated to the suitable and partly suitable Mg/Ca results with low δ^{18} O values coinciding with highest Mg/Ca values as shown in Figure 4. Considering that in modern oyster shells Mg/Ca covaries with temperature (Surge and Lohmann, 2008; Mouchi et al., 2013), CL intensity increases during summer months (Langlet et al., 2006; Lartaud et al., 2010b) and δ^{18} O is negatively correlated to temperature (Anderson and Arthur, 1983), we conclude that all the geochemical tracer analysed in the fossil shells selected here reflect a primary environmental signal. Moreover, the results from these species are all cyclic suggesting seasonal variations in conditions which makes Mg/Ca (and δ^{18} O) a promising tool for intraannual palaeoclimate reconstructions.

- F. bashibulakeensis specimens yielded no suitable profiles, a few partly suitable and mainly unsuitable results displaying a non-cyclic pattern with highly variable Mg/Ca ratios throughout the shell. This may be partly explained by the curvature of the ligamental area affecting the reliability of the positioning of the sampling transects. This is supported by the fact that parts of the transects that may be suitable are perpendicular to the growth direction while unsuitable part of the transects cross the growth bands at an oblique angle. We therefore conclude that species with a curved ligamental area are less suitable for trace element analyses as performed here. Developing a new sampling method following the curvature more closely may yield more reliable results in oysters with strongly curved ligamental areas.
- F. hemiglobosa and O. bellovacina yielded no suitable profiles, a few partly suitable and many unsuitable Mg/Ca patterns characterized by variable intra-shell composition and absence of a well-defined cyclicity (Figure 8). This was observed even from samples from the same stratigraphic level in the Kuhdara section suggesting this is not an actual environmental effect (Table 2). In addition, this unreliability is unlikely caused by diagenesis, since the incremental δ¹⁸O data of these shells display regular cycles (Figure Figure 5) and that CL values show no alteration through the ligamental area indicating good preservation. For some

specimens of F. hemiglobosa, the apparent mismatch in cyclicity between the stable isotopes and Mg/Ca, may be caused by the position of the cutting plane, which crossed the side of the ligamental area (within a side bead along the resilifer) instead of cutting through the middle. Considering the very low CL intensity (partially correlated to Mn concentration) and the unsuitable Mg/Ca signal, trace element incorporation appear to be altered in the side parts of the ligamental area, making them unsuitable for application of trace elements to environmental reconstructions. However, this can not explain all unsuitable data. A few other F. hemiglobosa and O. bellovacina present substantial intra-shell variability in Mg/Ca (e.g. MS13-O23, KZ12-O12.2 or Kuhdara samples, Table 2) although they have been cut properly and CL as well as $\delta^{18}O$ show cyclic variations with no diagenetic overprint. The cause of the mismatch between $\delta^{18}O$ and Mg/Ca remains to be investigated for these Palaeocene species. The non-cyclic pattern of trace element incorporation may relate to species-specific or environmental effects and are discussed below.

5.2. Environmental and species effects

Oyster shells from the same sedimentary level (that have calcified at the same time and location) yielded similar isotopic and Mg/Ca records (Figure 9j). However, these values differ significantly and sometimes systematically when comparing different species, time periods and/or living environments suggesting that the records depend both on species and on environmental conditions. Comparison of the δ^{13} C and δ^{18} O from bulk and incremental average data (Figure 9i) shows that the samples can be divided into two groups: one with both depleted $\delta^{18}O$ and $\delta^{13}C$ values and one with more enriched δ^{18} O and δ^{13} C values. This may reflect the influence of freshwater (by e.g. continental runoff), resulting in depleted carbon and oxygen isotope rations of shell for species living in the shallowest sites (Lartaud et al., 2010a). This is particularly clear when comparing values from the species F. hemiglobosa in offshore environments at the Aertashi section to values from the same species in a relatively nearshore bay environment at the Mine and Kuhdara sections (Figure 9i). Furthermore, O. (T.) strictiplicata, S. orientalis and S. buhsii are all collected from fully subtidal marine environments and have similar $\delta^{18}O$ and $\delta^{13}C$ to other fully marine specimens and differ strongly from, for example, P. asiatica that lives in a nearshore mangrove environment (Figure 2). Despite this apparent systematic environmental effect on isotopic fractionation, it remains to be investigated whether this is also reflected in the Mg/Ca since reliable profiles (i.e. suitable and

reliable parts of partly suitable profiles) were not available from specimens of the same species that
calcified in different depositional environments. Only *F. hemiglobosa* provides this opportunity, and
shows no significant differences between specimens collected from various environments as most
analyses are unsuitable.

To estimate potential species-specific effects, multiple reliable Mg/Ca profiles are compared from 320 different species from the same living environments (Figure 9). Mg/Ca results within a species col-321 lected from one area and the same geological period are relatively similar (based on the standard de-322 viation of their average Mg/Ca). This indicates consistency of the Mg/Ca signal between specimens 323 of one single oyster species. However, absolute values and the range of Mg/Ca ratio vary between 324 species. For instance, O. (T.) strictiplicata and S. buhsii (Middle Eocene) have Mg/Ca averages of 325 approximately 6 mmol/mol with a seasonal amplitude of \sim 5 mmol/mol. For F. hemiglobosa (Early 326 Eocene) the average and seasonal amplitude in Mg/Ca is slightly lower around 4 mmol/mol. This 327 difference may be attributed to a palaeoenvironmental effect as these species lived during different 328 geological time intervals (early Eocene for F. hemiglobosa and the middle Eocene for O. (T.) stric-329 tiplicata). However, it is more likely due to a species-specific effect on the magnesium incorporation 330 into the calcitic shell as clearly indicated below in our analyses comparing temperatures derived 331 from these results. 332

333 5.3. Validity of Mg/Ca -temperature calibrations depending on species

Mg/Ca-temperature calibrations have been successfully developed for different bivalve species 334 (e.g. fan mussel, mussel, scallop, and oyster; Table 1, Klein et al., 1996; Vander Putten et al., 335 2000; Freitas et al., 2005, 2006, 2009, 2012; Wanamaker et al., 2008; Surge and Lohmann, 2008; 336 Mouchi et al., 2013). Given that these calibrations vary strongly between bivalves according on 337 the shell species (Figure 10), we will therefore only focus on calibrations established on modern 338 oyster species. For modern oysters, calibrations have been obtained by Surge and Lohmann (2008) 339 for the estuaries species Crassostrea virginica and by Mouchi et al. (2013) for the marine species 340 Crassostrea qiqas. To apply these existing calibrations to the extinct fossil species, including those 341 analysed in this study, it is necessary to account for potential inter-species differences. To do 342 so, temperatures obtained using various existing oyster calibrations are compared to temperatures 343 estimated from measured δ^{18} O. The carbonaceous shells of most bivalves are precipitated in isotopic equilibrium with the seawater (e.g. Kirby et al., 1998) so that the oxygen isotopic composition of 345

different species can be directly compared. Thereby, relationships between seawater temperature, calcite $\delta^{18}O_c$, and seawater $\delta^{18}O_{sw}$ are commonly applied to estimate temperatures using fossil bivalve shells (e.g. Ivany et al., 2000; Gillikin et al., 2005). Here we apply calibration established by Anderson and Arthur (1983) between seawater temperature T (°C), bivalve calcite $\delta^{18}O_c$ (%0, VPDB), and seawater $\delta^{18}O_{sw}$ (%0, SMOW):

$$T = 16 - 4.14 \times (\delta^{18}O_c - \delta^{18}O_{sw}) + 0.13 \times (\delta^{18}O_c - \delta^{18}O_{sw})^2$$
(1)

To estimate $\delta^{18}O_{sw}$ we used a range between the commonly accepted -1‰ value for global icefree Eocene seawater (e.g. Ivany et al., 2000) and the more realistic value of 0.65‰ provided recently by numerical modelling near our study area at an Eocene position of 37.5°N, 71.25°E corresponding to the eastern Proto-Paratethys (Tindall et al., 2010). Resulting calculated seawater temperatures vary from approximately 22 to 41°C for average temperatures, ~25 to 46°C for summer temperatures and ~20 to 37°C for winter temperatures (Figure° 11).

Applying the Mg/Ca-temperature calibration for the extant oyster C. virginica (Surge and 357 Lohmann, 2008) to our fossil oysters results in temperatures that are considerably lower than those 358 based on $\delta^{18}{\rm O}$ assuming a $\delta^{18}{\rm O}_{\rm sw}$ of 0.65%0 or even -1%0 (Figure 10). The application of this 359 Mg/Ca-temperature calibration is therefore not suitable for any of the oyster species considered here, as was shown before on S. buhsii (Bougeois et al., 2014). In contrast, δ^{18} O-derived temperatures 361 compare better with those based for the Mg/Ca calibration of the marine oyster species C. qiqas 362 (Mouchi et al., 2013, Figure 10). Previous studies on calcifiers noticed that because seawater Mg/Ca 363 was 50% of modern in the Palaeogene (3 mmol/mol compare to 5 mmol/mol in average), this could 364 be a significant source of uncertainty which may have led to Palaeogene species being characterised 365 by lower Mg/Ca ratios (Evans and Müller, 2012). However, the calibration done by (Mouchi et al., 366 2013) was established in Arcachon Basin were the seawater Mg/Ca is more or less 5 mmol/mol 367 which is close to the modern seawater Mg/Ca ratio. Furthermore even for modern time, seawater 368 Mg/Ca is highly variable (1) between open and coastal environment and (2) at an infra annual 369 scale especially in coastal or near shore environment. According Lorens and Bender (1980) the 370 relationship between Mg in the environment, in the biological fluids and in the calcitic part the 371 carbonaceous shell is not clear (compare to Sr). Even if Mouchi et al. (2013) thermodependent 372 equation do not include seawater Mg/Ca, Surge and Lohmann (2008) showed that a large variation 373 of seawater Mg/Ca from 1 to 5 mmol/mol (due to the proximity of Mississipi delta) do not have a 374

significant influence on the oyster shell Mg/Ca. This encourage the sue of such calibrations, even in Palaeocene and Eocene times.

However, the validity of the calibration of Mouchi et al. (2013) varies between analysed species.

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- Mg/Ca values from O. bellovacina, F. hemiglobosa, P. asiatica and F. bashibulakeensis appear to result in unrealistic temperatures using the calibration of Mouchi et al. (2013). Summer temperatures from specimens MS13-O23 (F. hemiglobosa) and KZ12-O12.2 (O. bellovacina) are unrealistically high (>50-60°C). Comparatively, the Mg/Ca- based temperatures for specimens KA12-O02.2 (O. bellovacina), MS12-O06.5 (F. bashibulakeensis), AB11-O01.1 (P. asiatica), AT12-O07B and TK11-O04 (both F. hemiglobosa) are unrealistically low (down to 10°C) compared to the δ^{18} O-based temperatures (32 to 40°C in average with a constant δ^{18} O_{sw} = $0.65\%_0$, Figure 11b). An overestimation of $\delta^{18}O_{sw}$ of around -4\%0 would be required to match the temperatures deduced from both proxies, which is very unlikely in an open marine environment. Furthermore, given the overall warm conditions for the Palaeocene-Eocene and particularly for the Central Asian sample location, temperatures as low as 10°C are not realistic (Lan, 1997; Sun and Wang, 2005; Guo et al., 2008; Tindall et al., 2010; Quan et al., 2012). We conclude that the Mg/Ca-temperature calibration of Mouchi et al. (2013) does not apply to shells of O. bellovacina, F. hemiqlobosa, P. asiatica and F. bashibulakeensis. This suggests that incorporation of Mg differs significantly between species, even within the Ostreidae superfamily. Furthermore, specimens of the species P. asiatica analysed here were living in a marine environment under the influence of continental runoff (mangrove area, restricted marine waters, Figure 2). This may suggest also a potential additional impact from the environment on the magnesium incorporation for this species.
- S. buhsii and O. (T.) strictiplicata, in contrast, appear to yield reliable temperatures using the calibration of Mouchi et al. (2013) that are within the same range as those estimated using δ^{18} O (around 25 and 30°C in average with a δ^{18} O_{sw} = 0.65%, Figure 11b). Considering that these species also provide the most reliable Mg/Ca records discussed above (Figure 6), we conclude that these species are particularly suitable for estimating palaeotemperatures.

Although average Mg/Ca-derived temperatures are comparable to those based on δ^{18} O, the seasonal amplitudes in temperature are significantly higher for Mg/Ca compared to δ^{18} O (Figure

11; also discussed in Bougeois et al., 2014) this likely relates to seasonal fluctuations in the δ^{18} O of 404 seawater as expected in coastal marine living environments of the studied oysters according to the 405 interpreted depositional palaeoenvironments. Indeed sedimentological (Manceau et al., 2014) and 406 palynological (Sun and Wang, 2005; Guo et al., 2008; Quan et al., 2012) data indicate coeval arid 407 to semi-arid conditions within evaporitic precipitation in playa and sabkha environments. These 408 conditions are prone to seasonal fluctuations of salinity in subtidal environment which in turns 409 regulate the $\delta^{18}O_{sw}$. Furthermore, the Mg-estimated temperatures are consistent with SST derived 410 from numerical modelling for this area in the Eocene (Tindall et al., 2010) as well as SST in 411 modern analogous environments previously studied with shell sclerochronology in the Upper Gulf 412 of California (Goodwin et al., 2001) or the San Francisco Bay (Goodwin et al., 2012). Together, 413 these observations suggest that the modern Mg/Ca-temperature calibration for C. qiqas (Mouchi 414 et al., 2013) can be applied to extinct fossil species of O. (T.) strictiplicata and S. buhsii. In general, 415 this indicates that, after careful inspection of the diagenesis effect and independent characterization 416 of the depositional environments, Mg-temperature calibrations based on modern specimens can be 417 used to reconstruct past temperatures and seasonal patterns therein from fossil oyster shells. 418

419 6. Further development and conclusions

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By comparing Mg/Ca to δ^{18} O high-resolution profiles from various fossil species in different depositional environments we devised a protocol to identify fossil species yielding reliable temperatures when using Mg/Ca-temperature calibrations derived from existing studies on modern oyster species.

The palaeobiogeographic distributional patterns of the reliable oyster species identified here (S.424 buhsii and O. (T.) strictiplicata) show an extraordinarily wide dispersal beyond the Tarim Basin 425 into the regional Turkestan stage of Central Asia (Vyalov, 1937), in Northwest Afghanistan (Berizzi, 426 1970) and in northern Iran (Grewingk, 1853). Occurrences of Palaeogene oysters have also been 427 reported in the Transylvanian Basin (Rusu et al., 2004) and as far west as the Paris Basin (for 428 O. bellovacina Lan and Wei, 1995). This broad distribution makes these species an ideal choice 429 to track palaeo-seasonality in a large area during the particular Palaeogene period, when global 430 climate changed from greenhouse to icehouse conditions. 431

Combining Mg-estimated temperature with δ^{18} O allows reconstructions of seasonal variabil-

ity in $\delta^{18}O_{sw}$ ultimately reflecting changes in salinity (see equation 1 and supplementary data for 433 these calculations, Bougeois et al., 2014). Seasonally-resolved salinity can be used for palaeoclimate 434 interpretation of the basin hydrological cycle (evaporation and precipitation) at intra-annual reso-435 lution thereby significantly improving the comprehension of the environment compared to previous 436 palaeontological studies as exemplified in our study area (Sun and Wang, 2005; Guo et al., 2008; 437 Quan et al., 2012). Furthermore, by comparing Mg/Ca and δ^{18} O from multiple species from the 438 same stratigraphic level, one could in principle statistically deconvolve impact of differences in sea 439 water Mg/Ca or establish freshwater to seawater mixing lines from δ^{18} O. 440

In principle, the protocol developed here is applicable to oyster (or even more largely bivalve) 441 fossil species if they are found to yield reliable temperatures using the few existing temperaturecalibrated modern species. The necessity to perform additional calibration studies on modern 443 oyster species to improve calibrations for Mg/Ca and temperature for various Ostreidae species 444 follows is underscored by our inconsistent results with F. hemiglobosa or F. bashibulakeensis species 445 from which no reliable Mg-temperature could be extracted. Further calibration work may also 446 help identify possible specific vital effect on Mg incorporation into the shell (already observed on 447 modern species within a same genus such as Mytilus sp. or Crassostrea sp., Table 1), as well as 448 the environmental impact (observed on same species Pecten maximus in different environment such 449 as estuarine (Vander Putten et al., 2000), culturing (Freitas et al., 2008) or brackish (Wanamaker 450 et al., 2008)). Furthermore, these studies will certainly benefit from the calibration of other trace 451 elements and isotopes such as Ba and δ^{13} C recently used as indicators of primary productivity 452 (Goodwin et al., 2012). 453

Improving the knowledge on biomineralisation of Ostreidae shells is a prime target to develop
a reliable and powerful environmental proxies. Indeed, the widespread occurrence of Ostreidae in
the fossil records in terms of age (from Triassic until Quaternary) and of geolocalisation (from low
to high latitudes, from brackish to fully marine environments) make Ostreidae the most promising
group to infer palaeenvironments at intra-annual resolution.

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- Wefer, G., Berger, W.H., 1991. Isotope paleontology: growth and composition of extant calcareous
- species. Marine geology 100, 207–248.

635 Tables and Figures

Table 1: Comparison of Mg/Ca—seawater temperature relationships for calcitic bivalves.

Table 2: Summary of experiments performed on the 41 oyster slabs. The three last columns indicate the reliability of Mg/Ca transects based on the two main characteristics: Intra-shell variability and Signal cyclicity ("+", "=" and "-" correspond respectively to suitable, partly suitable and unsuitable signal). The reliability of the Mg/Ca result for each oyster is expressed by: S = suitable, PS = partly suitable, US = unsuitable.

Figure 1: **a.** Location of sampling area in Kyrgyzstan (yellow), Tajikistan (blue) and China (orange). **b.** Middle Eocene Palaeographical map showing Central Asia invaded by the Proto-Paratethys sea (map from Licht et al., 2014). **c.** Position of analysed species on schematic illustration of the third to fifth marine incursions into the Tarim basin (Bosboom et al., 2014b). **d.** Examples of analysed fossil oyster left valves with well-developed ligamental area. (**i.**) *F. hemiglobosa*, (**ii.**) *O.* (*T.*) strictiplicata, (**iii.**) *S. buhsii*, (**iv.**) *P. asiatica*, (**v**). *F. bashibulakeensis*. The first part of the specimen's name indicates the provenance with the code of the stratigraphic sections (see section codes below), and the year of sampling. The complete list of oysters from Central Asia analysed in this study is available in supplementary data. Code for stratigraphic sections - MS: Mine-Bashibulake (39° 51'N, 74° 32'E), KA: Kansu (39° 45'N, 74° 58'E), KZ: Kezi (38° 26'N, 76° 24'E), AT: Aertashi (37° 58'N, 76° 33'E), YK: Yarkand (37.7° N, 76.6° E), KY: Keyliand (37° 27'N, 77° 86'E), AB: Ala Buka (41.4° N, 71.4° E), TK: Tash Kumyr (41.3° N, 72.2° E), AL: Alai Valley (39.6° N, 72.4° E), UT: Uch Tobo (39.9° N, 73.4° E), DS: Kuhdara-Dushanbe (38.7° N, 68.9° E).

Figure 2: a. Environment of the studied species: age, formation, depositional environment and associated fauna. b. Living position of species through the carbonate platform.

Figure 3: Radial sections (top) of oyster ligamental areas showing light and dark alternations, each couplet corresponding to one living growth years. Cathodoluminescence assemblages (bottom) for these specimens show alternation of areas with high and low luminescence corresponding to light and dark bands observed in natural light. Transects followed by laser ablation for trace element analyses are indicated in blue and green. Main paths followed by the Micromill for stable isotopes analyses are indicated in black. Black arrows indicate growth directions. (a.) O. bellovacina, (b.) F. hemiglobosa, (c.) S. buhsii and (d.) F. bashibulakeensis. Supplementary data provides the radial sections and CL images of complete collection. e. Geochemical data for the specimen KZ12-O03 (S. buhsii): Cathodoluminescence grey scale (light red: raw data, dark red: moving average) results, Mg/Ca ratio for two parallel transects yielding similar results (light blue or green: raw data, dark blue or green: moving average), Oxygen stable isotopes from the selected area. Grey areas correspond to light and dark layers as revealed by CL.

Figure 4: Geochemical data for the specimen KZ12-O03 (S. buhsii): Cathodoluminescence grey scale (light red: raw data, dark red: moving average) results, Mg/Ca ratio for two parallel transects yielding similar results (light blue or green: raw data, dark blue or green: moving average), Oxygen stable isotopes from the selected area. Grey areas correspond to light and dark layers as revealed by CL.

Figure 5: Incremental stable oxygen ratio records (δ^{18} O) in oyster shells. Horizontal axis indicates distance in millimetres starting from the first drilled micro-sample. Grey areas correspond to visible dark bands on the shells, white areas correspond to the light bands revealed by CL analyses.

Figure 6: Suitable Mg/Ca result across the oyster shell ligamental area showing both well-expressed cyclicity and low intra-shell variability between parallel transects (Figure 3). Analyses were performed following the growth direction with 0 mm corresponding to the most juvenile part analysed. See Figure 8 for explanatory legend of used symbols and patterns.

Figure 7: Partly suitable Mg/Ca results across the oyster shell ligamental area showing only parts with well-expressed cyclicity and low intra-shell variability between parallel transects (Figure 3). Analyses were performed following the growth direction with 0 mm corresponding to the most juvenile part analysed.

Figure 8: Unsuitable Mg/Ca results across the oyster shell ligamental area showing poorly-expressed cyclicity and high intra-shell variability between parallel transects (Figure 3) of oyster shells. Analyses were performed following the growth direction with 0 mm corresponding to the most juvenile part analysed. See Figure 8 for explanatory legend of used symbols and patterns.

Figure 9: **a.-d.** δ^{18} O results from bulk and incremental analyses per stratigraphic section. **e.-h.** Mg/Ca results from incremental analyses per stratigraphic section. When the seasonal pattern could not be defined, we represented only the average (full black square) and a black line between the highest and the lowest values. Red depicts suitable signal, blue partly suitable signal and grey unsuitable signal. When oysters are coming from the same bed, the estimated age is the same. However, to better see the difference between them, we juxtaposed results next to each other. Numbers correspond to the Reference N° in Table 2. **i.** δ^{13} C vs δ^{18} O for bulk and averaged incremental results. Two groups can be clearly distinguished between relatively high (full line) and low (dashed line) values of δ^{18} O and δ^{13} C. **j.** Compiled Mg/Ca results from the complete collection with suitable and partly suitable criteria.

Figure 10: **a.** Infra-annual temperatures (0.2 mm moving average) for the specimen AT13-O20 estimated using various Mg/Ca calibrations established on different species of calcitic bivalves (1 to 10, cf Table 1), or using δ^{18} O calibration of Anderson and Arthur (1983) with a constant δ^{18} O_{sw} of 0.65% according to modelled values for the study area in the Eocene (Tindall et al., 2010) (11), or of -1% according to the value commonly used for the Eocene ice free world (12). **b.** Associated temperature range for each calibration.

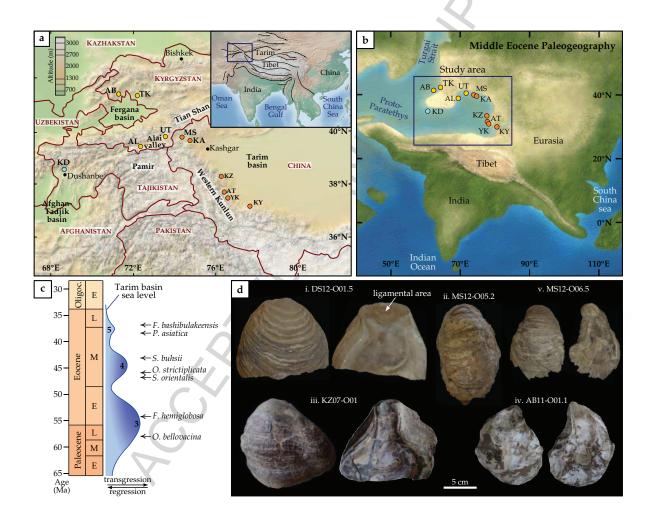
Figure 11: **a.** Estimated temperature using Mg/Ca ratio with the calibration of Mouchi et al. (2013) (red line; in order to not overcrowd the graphs, only the moving average of Mg estimated temperature are shown) compared to estimated temperature using δ^{18} O with the calibration of Anderson and Arthur (1983) and with a constant δ^{18} O_{sw} of 0.65‰ (green plain circles) or -1‰ (orange empty circles). Grey areas correspond to dark bands on the shells. **b.** Comparison between range of temperatures deduced from Mg/Ca (Mouchi et al., 2013) and δ^{18} O (Anderson and Arthur, 1983) with a constant δ^{18} O_{sw} of 0.65‰.

Species	Environment	Temperature range	Mg/Ca = f(T)	Reference	
Pinna nobilis	Pinna nobilis marine 10-2		$Mg/Ca = 17.16 (\pm 1.95) * exp(0.022 (\pm 0.004) * T)$	Freitas et al., 2005 (1)	
	culturing	10-20 °C	$Mg/Ca = 2.56 (\pm 0.42) + 0.17 (\pm 0.03) * T$	Freitas et al., 2012 (2)	
Pecten maximus	marine	5-19°C	$Mg/Ca = 4.92 (\pm 2.16) + 0.50 (\pm 0.16) * T$	Freitas et al., 2006 (3)	
	culturing	10-20 °C	$Mg/Ca = 9.89 (\pm 2.96) + 0.51 (\pm 0.19) * T$	Freitas et al., 2008 (4)	
Mytilus trossolus	marine	6-23°C	$Mg/Ca = 2.25 (\pm 0.63) + 0.30 (\pm 0.04) * T$	Klein et al., 1996 (5)	
	estuarine	5-20°C	$Mg/Ca = -0.63 (\pm 0.29) + 0.70 (\pm 0.02) * T$	Vander Putten., 2000 (6)	
Mytilus edulis	culturing	10-20 °C	$Mg/Ca = 1.50 (\pm 0.57) + 0.27 (\pm 0.04) * T$	Freitas et al., 2008 (7)	
	brakish water	7-19°C	$Mg/Ca = 5.44 (\pm 0.31) + 0.77 (\pm 0.22) * T$	Wanamaker et al., 2008 (8)	
Crassostrea gigas	marine	5-25°C	Mg/Ca = -0.50 + 0.27 * T	Mouchi et al., 2013 (9)	
Crassostrea virginica	estuarine	18-32°C	Mg/Ca = -0.23 + 0.72 * T	Surge et al., 2008 (10)	

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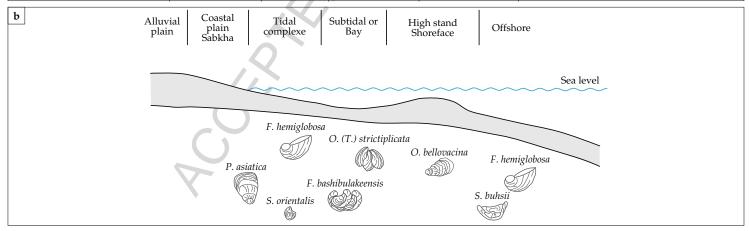
ACCEPTED MANUSCRIPT											
Section	Age	# Sample	Ref. N°	Species	CL	Bulk isotopes	Incremental isotopes	LA – ICP MS	Intra-shell variability	Cyclicity	Pattern
T/	Thanetian	KA12-O02.1	1	O. bellovacina	/	1		1	+	=	PS
Kanzu	Thanetian	KA12-O02.2	2	O. bellovacina	1	1	✓	1	+	=	PS
	Ypresian	MS12-O15	3	F. hemiglobosa	1	1		1	=	+	PS
	Ypresian	MS13-O23	4	F. hemiglobosa	1	1	✓	1	_	_	US
	Early Lutetian	MS12-O05.2	5	O. strictiplicata	1	1	✓	1	+	+	S
	Early Lutetian	MS10-O02.1	6	O. strictiplicata	1	1					
	Early Lutetian	MS13-O22	7	O. strictiplicata	1	✓			+	+	S
Mina	Late Lutetian	MS12-O10.2	8	S. buhsii	/	1		1	_	_	US
Mine	Late Lutetian	MS10-O03	9	S. buhsii	/	1					
	Late Lutetian	MS13-O20	10	S. buhsii	/	1		1	+	=	PS
	Early Priabonian	MS12-O06.2	11	F. bashibulakeensis	1	1		1	_	_	US
	Early Priabonian	MS12-O06.5	12	F. bashibulakeensis	/	1	/	1	=	+	PS
	Early Priabonian	MS10-O04	13	F. bashibulakeensis	/	1		1	_	_	US
	Early Priabonian	MS13-O21	14	F. bashibulakeensis	/	1		1	=	_	US
	Ypresian	AT12-O07.A	15	F. hemiglobosa	1			1	+	=	PS
	Ypresian	AT12-O07.B	16	F. hemiglobosa	>	1	✓	1	=	=	PS
	Ypresian	AT13-O21	17	F. hemiglobosa	>	1		1	+	_	US
	Middle Lutetian	AT12-O09.2	18	S. buhsii	1	1		1	+	+	S
	Middle Lutetian	AT12-O09.3	19	S. buhsii	1	1	✓	1	+	+	S
Aertashi	Middle Lutetian	AT13-O20	20	O. strictiplicata	/	1	1	1	+	+	S
	Late Lutetian	AT12-O15.1	21	S. buhsii	1	1		1	_	_	US
	Late Lutetian	AT12-O015.2	22	S. buhsii	/	1	1	1	+	+	S
	Late Lutetian	AT13-O19	23	S. buhsii	1	1	1	1	+	+	S
	Late Lutetian	AT12-O18	24	S. buhsii	1	1		1	=	+	PS
	Late Lutetian	AT11-O04	25	S. buhsii	1	1	✓	1	+	+	S
	Thanetian	KZ12-O12.2	26	O. bellovacina	/	1	1	1	_	+	US
	Middle Lutetian	KZ12-O05.1	27	S. buhsii	1	1	✓	1	_	+	US
Kezi	Late Lutetian	KZ12-O03	28	S. buhsii	1	✓	✓	1	+	+	S
	Late Lutetian	KZ07-O01	29	S. buhsii	1	1	✓	1	=	+	PS
Keyliand	Late Lutetian	KY10-O01	30	S. buhsii	/	1	✓	1	+	+	S
Yarkand	Ypresian	YK11-O01.1	31	F. hemiglobosa	/	1		1	=	+	PS
	Ypresian	DS12-O01.2A	32	F. hemiglobosa	/	1		1	_	-	US
	Ypresian	DS12-O01.2B	33	F. hemiglobosa	/	1		1	=	_	US
Kuhdara	Ypresian	DS12-O01.3	34	F. hemiglobosa	/	1		1	-	_	US
	Ypresian	DS12-O01.4	35	F. hemiglobosa	/	✓		✓	-	_	US
	Ypresian	DS12-O01.5	36	F. hemiglobosa	/	1	✓	1	_	=	US
	Ypresian	TK11-O04	37	F. hemiglobosa		1	✓	1	=	=	PS
	Early Lutetian	AL11-O02.1	38	O. strictiplicata	/	1	✓	1	+	+	S
Tash-Kumyr -	Early Lutetian	UT11-O01.1	39	S. orientalis	/	1		1	_	_	US
Uch Tobo -Alai Valley -	Early Lutetian	UT11-O02.1	40	S. orientlis	/	1		1	+	+	S
Ala Buka	Late Lutetian	TK11-O03.2	41	S. buhsii	1	1		1	=	+	PS
	Late Bartonian	AB11-O01.1	42	P. asiatica		1	✓	1	=	+	PS
	Late Bartonian	AB11-O01.2	43	P. asiatica		1		1	=	=	US

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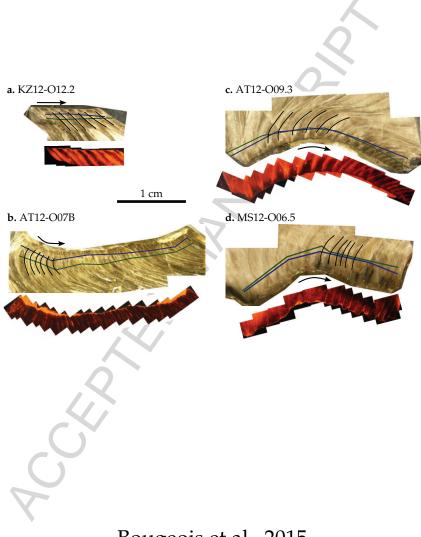


Bougeois et al., 2015 Figure 1

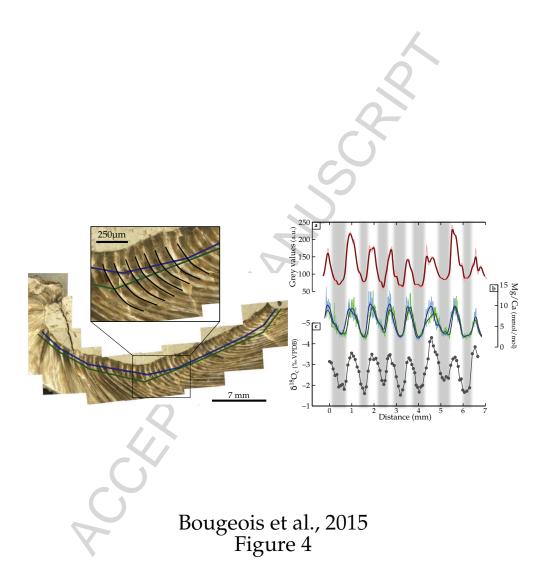
a	Species	Age	Formation	Environment	Sediment	Main associated fauna
	Ostrea bellovacina (Lamarck, 1896)	Thanetian	Lower Qimugen	shoreface	oolithic bioclastic wackestone	Pycnodonte nomada, Bryozoa, Ostracodes, Algae, Urchin spines
	Flemingostrea hemiglobosa (Romanovskiy, 1884)	Ypresian	Lower Qimugen	offshore or quiet bay	green marls	Foraminifera, Echinoids, Bryozoa, Fish scales
(0	Sokolowia orientalis Gekker, Osipova & Balskaya, 1962)	Early Lutetian	Kalatar	intertidal	bioclastic wackestone	Ostrea afghanica
	Ostrea (T.) strictiplicata (Roulin and Delbos, 1855)	Early to Middle Lutetian	Kalatar	subtidal reef	bioclastic wackestone	Ostrea (T.) cizancourti, Gastropods, Bryozoa, algae
	Sokolowia buhsii (Grewingk, 1853)	Early to Middle Lutetian	Wulagen	offshore	green mudstones	Foraminifera, Echinoids, Briozoa, Serpulids, Sponge, Chlamys sp., Kokanostrea kokanensis
	Platygena asiatica (Romanovskiy, 1879)	Late Bartonian	Bashibulake	mangrove	bioturbated sandy siltstones	Cubisostrea plicata, roots
	Ferganea bashibulakeensis (Wei, 1984)	Early Priabonian	Bashibulake	subtidal or restricted marine	bluish marly siltstones	Lithophaga, Foraminifera, Chlamys sp.

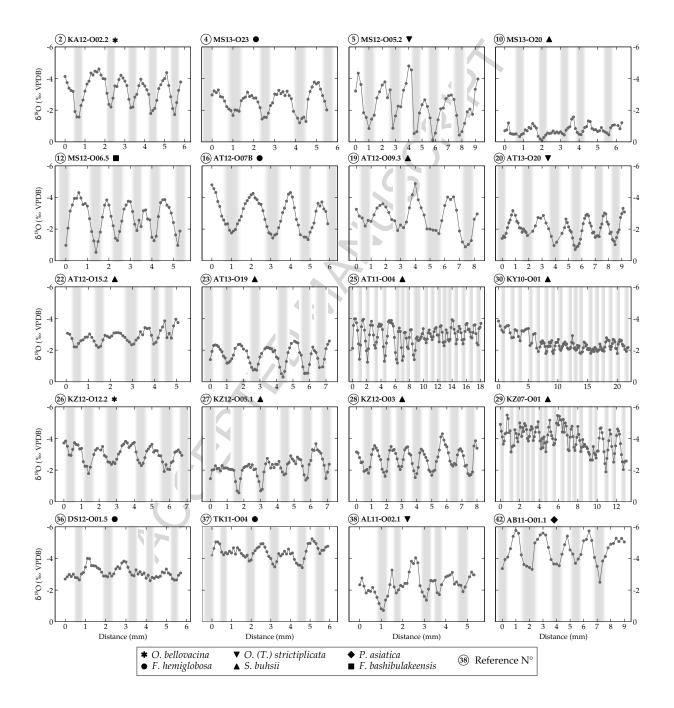


Bougeois et al., 2015 Figure 2

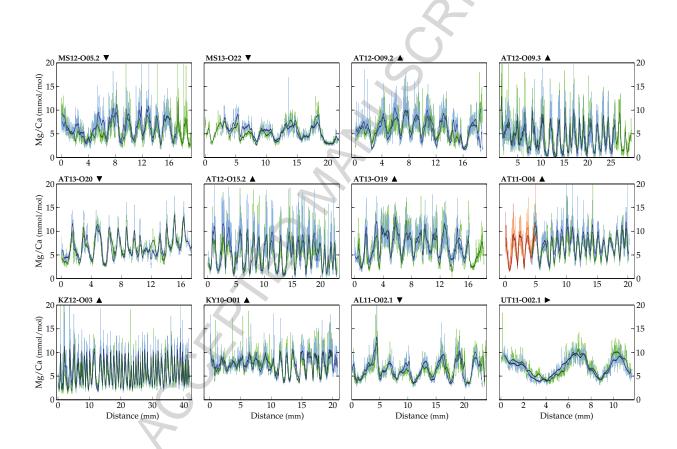


Bougeois et al., 2015 Figure 3

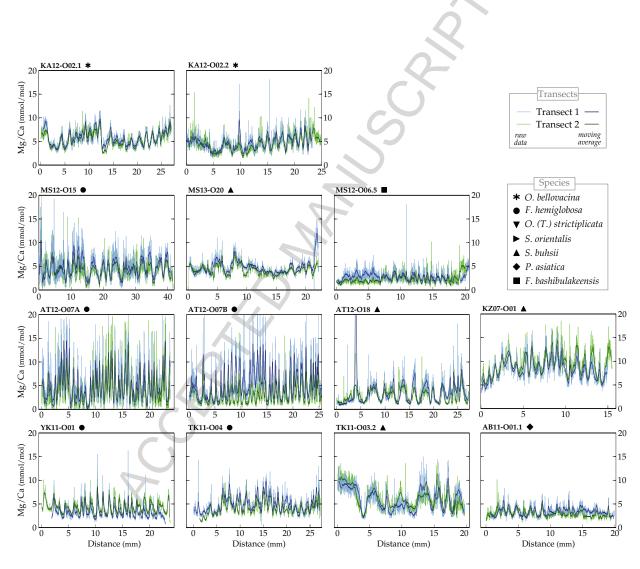




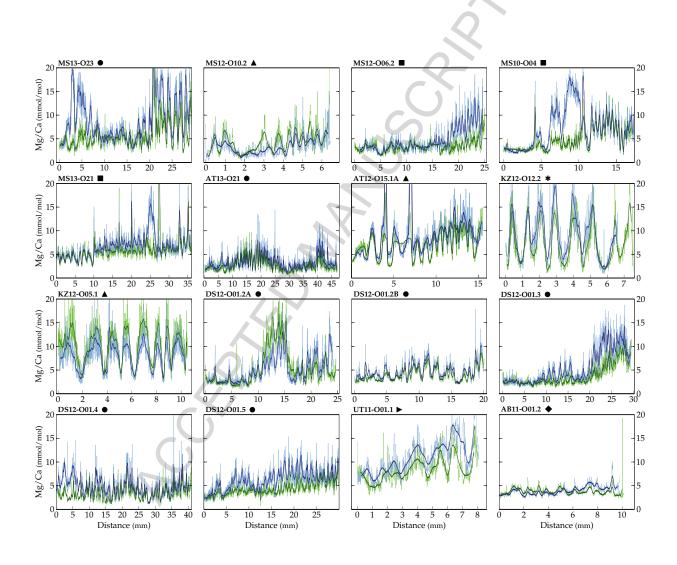
Bougeois et al., 2015 Figure 5



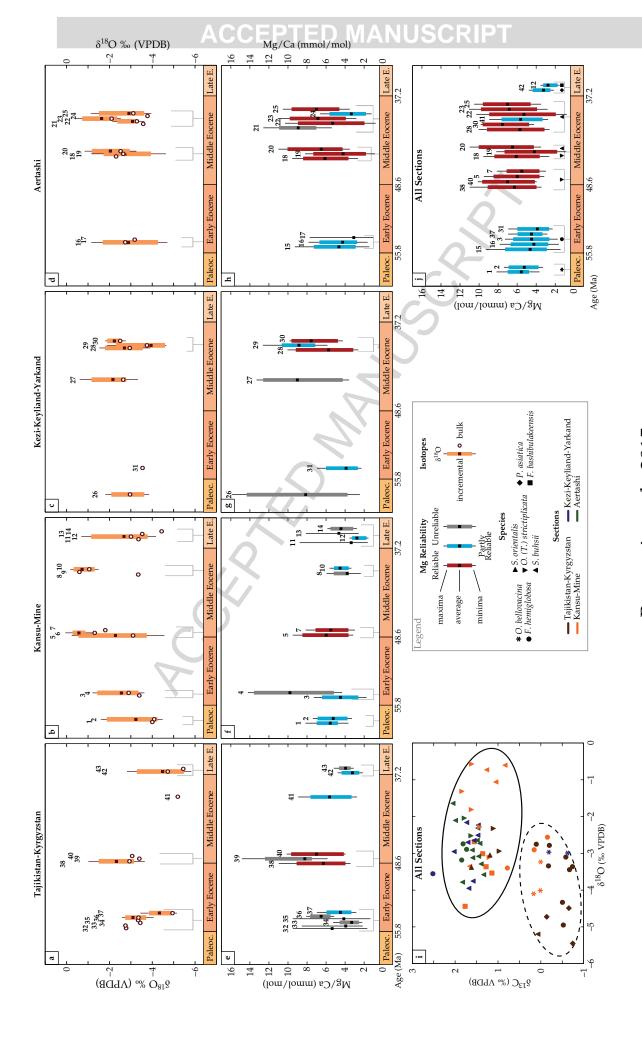
Bougeois et al., 2015 Figure 6



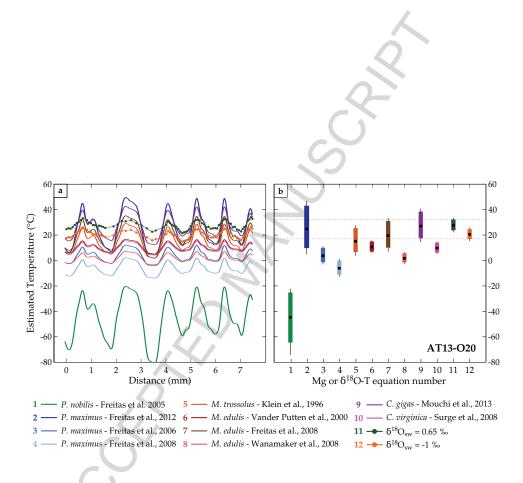
Bougeois et al., 2015 Figure 7



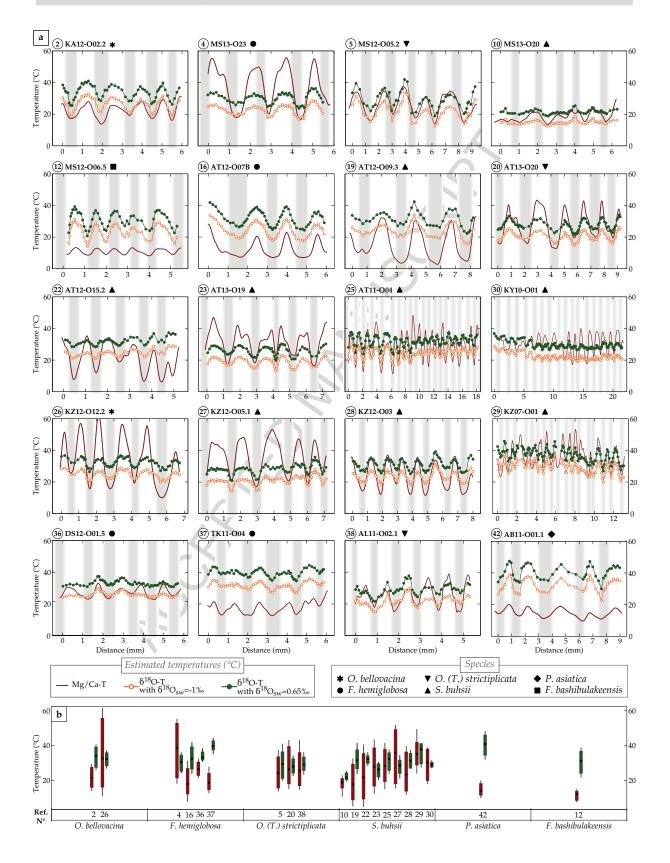
Bougeois et al., 2015 Figure 8



Bougeois et al., 2015 Figure 9



Bougeois et al., 2015 Figure 10



Bougeois et al., 2015 Figure 11

Highlights:

- New application to recover seasonal temperatures from Mg/Ca in fossil oyster shells
- 40 specimens analysed from 7 species living in the Palaeogene Paratethys Sea
- Combining Mg/Ca and δ 18O to identify reliable species and salinity variations
- Assessing seasonal temperatures in semi-arid area