



HAL
open science

Aided phytoextraction of Cu, Pb, Zn, and As in copper-contaminated soils with tobacco and sunflower in crop rotation: Mobility and phytoavailability assessment

Nour Hattab-Hambli, Mikael Motelica-Heino, Michel Mench

► To cite this version:

Nour Hattab-Hambli, Mikael Motelica-Heino, Michel Mench. Aided phytoextraction of Cu, Pb, Zn, and As in copper-contaminated soils with tobacco and sunflower in crop rotation: Mobility and phytoavailability assessment. *Chemosphere*, 2016, 145, pp.543-550. 10.1016/j.chemosphere.2015.11.051 . insu-01251108

HAL Id: insu-01251108

<https://insu.hal.science/insu-01251108>

Submitted on 18 Mar 2016

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Aided phytoextraction of Cu, Pb, Zn, and As in copper-contaminated soils with tobacco and sunflower in crop rotation: mobility and phytoavailability assessment.

Nour HATTAB- HAMBLI^{a,b*}, Mikael MOTELICA-HEINO^a, Michel MENCH^c

^a ISTO , UMR 7327 - CNRS/Université d'Orléans, Campus Géosciences, 1A, rue de la Férollerie, 45071 Orléans cedex 2, France
E-mail : nour.hattab@univ-orleans.fr

^b Université d'Orléans, UFR-Faculté des Sciences, Laboratoire de Biologie des Ligneux et des Grandes Cultures, UPRES EA 1207, Rue de Chartres, BP 6759, F-45067 Orléans Cedex 02, France

^c Université de Bordeaux, UMR 1202 BIOGECO, Bat B2, allée G. St Hilaire, CS50023, 33615 Pessac, France, and INRA, UMR 1202 BIOGECO, 69 Route d'Arcachon, 33613 Cestas cedex, France. E-mail : mench@bordeaux.inra

Keywords: Phytoextraction, Sunflower, Tobacco, Soil pore water, Potentially toxic trace elements, Phytoavailability

Abstract

Copper-contaminated soils were managed with aided phytoextraction in 31 field plots at a former wood preservation site, using a single incorporation of compost (OM) and dolomitic limestone (DL) followed by a crop rotation with tobacco and sunflower. Six amended plots, with increasing total soil Cu, and one unamended plot were selected together with a control uncontaminated plot. The mobility and phytoavailability of Cu, Zn, Cr and As were investigated after 2 and 3 years in soil samples collected in these eight plots. Total Cu, Zn, Cr and As concentrations were determined in the soil pore water (SPW) and available soil Cu and Zn fractions by DGT. The Cu, Zn, Cr and As phytoavailability was characterized by growing dwarf beans on potted soils and determining the biomass of their plant parts and their foliar ionome.

Total Cu concentrations in the SPW increased with total soil Cu. Total Cu, Zn, Cr and As concentrations in the SPW decreased in year 3 as compared to year 2, likely due to annual shoot removals by the plants and the lixiviation. Available soil Cu and Zn fractions also declined in year 3. The Cu, Zn, Cr and As phytoavailability, assessed by their concentration and mineral mass in the primary leaves of beans, was reduced in year 3.

1. Introduction

Phytoextraction is a less invasive, low-cost phytotechnology, which use the plants and their associated microorganisms to extract and translocate metal(loid)s from the soil to the harvestable plant parts. This technique aims at reducing either the total or extractable PTTE concentrations in contaminated soils to targeted levels, depending on the country legislation, within a reasonable time frame. In addition, plants used to phytoextract PTTE from contaminated soils must be tolerant to PTTE and adapted to the local soil and climate characteristics and biotic interactions (Keller et al., 2003). Phytoextraction is more economically feasible if, in addition to PTTE removal, crops produce biomass with an added value such as biofuel for the energy sector (e.g. oilseed, poplar and

38 willow short rotation coppices), fibers, essential oils and biosourced chemicals for ecocatalysis (Schwitzguébel et al., 2002; Vassilev
39 et al., 2004; Li et al., 2012). Phytoextraction can be applied on contaminated soils in combination with soil conditioners (so-called
40 aided phytoextraction) to promote the biomass production (Tangahu et al., 2011). Phytoextraction using sunflower (*Helianthus*
41 *annuus* L.) has several advantages, such as the plant's ability to accumulate moderate PTTE concentrations and extract PTTE such
42 as Zn, Pb, Cd and Cu from the soil due to its high biomass production (Nehnevajova et al., 2007a, 2009; Herzig et al 2014).

43 The PTTE accumulation in sunflower shoots depends on several soil and plant factors, notably PTTE exposure, root uptake,
44 root-to-shoot translocation, rooting depth and density, impacts of pests, pathogens, herbivores (Vassilev et al., 2002), soil pH, nature
45 of the sorbents, presence and concentration of organic and inorganic ligands, including humic and fulvic acids, root exudates,
46 microbial metabolites and nutrients (Efremova and Izosimova, 2012).

47 This work aimed at assessing the ability of a sunflower - tobacco crop rotation to remediate Cu-contaminated soils, with and
48 without an initial single addition of compost (5%) and dolomitic limestone (0.2%). For Cu, Zn, Cr and As, the hypotheses tested
49 were: (1) decreases in total dissolved concentrations in the soil pore water (SPW); (2) decreases in available concentrations in the
50 soil; and (3) decreases in the phytoavailability for bean plants. Six amended plots with increasing total soil Cu, i.e. 163, 268, 382,
51 518, 753, and 1170 mg Cu/kg, and one unamended plot (832 mg Cu/kg) were selected out of 28 field plots. Total dissolved
52 concentrations of Cu, Zn, Cr, and As were quantified in SPW sampled by Rhizon moisture probes to assess their mobility. The soil
53 exposure intensity of Cu and Zn was determined by DGT (diffusive gradients in thin films) probes. Phytoavailability of Cu, Zn, Cr
54 and As was characterized by cultivating dwarf beans on potted soils and analyzing the ionome of primary leaves.

55

56 **2. Material and Methods**

57

58 **2.1. Site and Soils**

59

60 The wood preservation site (6 ha) is located in the Gironde County (44°43'N; 0°30'O), southwest France. It has been used
61 for over a century to preserve and store timber, posts and utility poles and various Cu-based salts were successively utilized (Mench
62 and Bes, 2009). The soil is developed on an alluvial terrace (Fluvisol) containing alluvial materials from the Garonne River
63 combined with wind deposits (BRGM, 1978). Its texture is sandy, i.e. 85.8 % sand, 5.9 % clay, and 8.3 % silt. It contains 1.6 % of
64 organic matter (OM), and has a C/N ratio of 17.2, with generally a low cation exchange capacity (CEC, 3.5 cmol kg⁻¹) (Bes and
65 Mench, 2008). Copper is the major contaminant of topsoils, albeit with high spatial variation (65–2400 mg Cu kg⁻¹ soil DW). Total
66 soil As, Zn, and Cr, i.e. 10–53 mg As, 21–68 mg Zn and 20–87 mg Cr kg⁻¹, remain relatively low in the topsoils, close to
67 background values (Mench and Bes, 2009). The site was divided into fifteen sub-sites according to past and present activities, plant
68 communities, employees' evidence, aerial and archival photos and site history (Bes et al 2010; Kolbas et al., 2011).

69 The field trial, located at the P1-3 sub-site, consists in four blocks (2 m × 10 m), i.e. block #1: plots #1 to #10, block #2: plots
70 #11 to #20, block #3: plot #21 to #30 and block #4: plot 31 (Kolbas et al., 2011). Plots #1 to #30 were amended in March 2008 with
71 compost (5% w/w) and dolomitic limestone DL (0.2% w/w) (Kolbas et al., 2011) based on a previous pot experiment (Bes and
72 Mench, 1998). Block #4 remained unamended and was considered as a single plot (UNT #31). Soil amendments were carefully
73 mixed in the topsoil (0-0.25 m) with a stainless steel spade. Each amended block was divided into 10 plots (1 m × 2 m) (Fig. 1). An
74 uncontaminated plot (1 m × 2 m) from a kitchen garden (CTRL, Gradignan, France), located at 18 km from the site, from the same

75 alluvial terrace and with a similar soil type, was also studied. For detailed data on soil parameters in all plots see Kolbas et al.
 76 (2011).

77 The cropping history was sunflower in 2008 and 2009, tobacco in 2010 and sunflower in 2011 (Kolbas et al., 2011; Kolbas
 78 2012; Mench et al., 2012). In April 2010, three soil samples (1.5 kg soil FW, 0–25 cm soil layer) were collected in six amended
 79 plots with increasing total soil Cu, i.e. 163 (#20), 268 (#8), 382 (#14), 518 (#13), 753 (#26) and 1170 (#30) mg Cu/kg, in the
 80 unamended plot (832 mg Cu/kg) (Fig.1), and in the control plots (CTRL). The three samples from each plot were mixed with a
 81 stainless steel spade and combined to produce composite soil samples (1 kg FW) which were air-dried, sieved (<2 mm, nylon mesh)
 82 and manually homogenized.

83 The same plots were resampled in April 2011 to investigate the phytoextraction effects on the mobility, soil exposure
 84 intensity and phytoavailability of PTTE in the soils.

85 Figure 1: Diagram of total Cu concentration in the four blocks (mg kg⁻¹), adapted from Kolbas et al., 2011. Colors from red to purple
 86 reflect the increase in soil Cu contamination.

block #4	block #3	block #2	block #1
	2011		
	#30: 1170 mg Cu/kg	#20: 163 mg Cu/kg	#10: 306 mg Cu/kg
	#29: 1020 mg Cu/kg	#19: 258 mg Cu/kg	#9: 348 mg Cu/kg
	#28: 894 mg Cu/kg	#18: 357 mg Cu/kg	#8: 268 mg Cu/kg
	#27: 1070 mg Cu/kg	#17: 352 mg Cu/kg	#7: 359 mg Cu/kg
	#26 : 753 mg Cu/kg	#16: 317 mg Cu/kg	#6 : 239 mg Cu/kg
# 31 : 894 mg Cu/kg	#25 : 961 mg Cu/kg	#15: 379 mg Cu/kg	#5: 333 mg Cu/kg
#31 : 832 mg Cu/kg	#24 : 952 mg Cu/kg	#14: 382 mg Cu/kg	# 4 : 384 mg Cu/kg
#31 : 832 mg Cu/kg	#23 819 mg Cu/kg	#13: 518 mg Cu/kg	# 3: 334 mg Cu/kg
	#22 : 1140 mg Cu/kg	#12: 336 mg Cu/kg	#2: 273 mg Cu/kg
	#21: 944 mg Cu/kg	#11: 285 mg Cu/kg	# 1: 311 mg Cu/kg

87
 88
 89
 90
 91
 92

93 **2.2. Sampling and analysis of soil pore water**

94

95 One kg of air-dried soil from each plot (i.e. 163, 268, 382, 518, 753, 1170, 832 and CTRL) was potted (1.3L) after sieving
96 (2 mm). Soil pore waters (3X10 ml) were extracted from each pot by Rhizon soil moisture samplers (SMS, model MOM,
97 Rhizosphere Research Products, Wageningen, The Netherlands). A full description of the SPW extraction procedure is given in
98 Hattab et al. (2014).

99 The SPWs were stored at 4°C. A fraction of each soil pore water was acidified to 0.1 mol/L HNO₃ to measure the
100 concentrations of Cu, Zn, Cr, and As in the samples by HR-ICP-MS (Element 2, Thermofischer). The rest of the solutions were kept
101 for the following analyses: pH, electrical conductivity (EC), and dissolved organic carbon (DOC) which was determined by a carbon
102 analyzer (Shimadzu[®] TOC 5000A). The total organic carbon (TOC) was determined in the soil by Rock-Eval pyrolysis. This
103 instrument uses a ramped temperature pyrolysis technique whereby a small amount of soil sample (70 -100 mg) is heated in an inert
104 atmosphere (helium or nitrogen) and combusted with air to obtain several key geochemical parameters such as the total organic
105 carbon (TOC).

106

107 **2.3. Soil exposure intensity**

108

109 Bioavailable concentrations of Cu, Zn and Cr in the soils were measured by DGT devices with an active surface area of 3.14
110 cm². These probes consist of a plastic base containing three layers: the first one is a 0.45µm filter, the second is a polyacrylamide gel
111 diffusion layer and the third one a polyacrylamide gel incorporating a Chelex-100 resin that strongly binds the labile trace metal
112 species (Davison et al., 2000; Ernstberger et al., 2002).

113 The total mass of each metal (*M*), the flux, *F(t)*, of metal from the soil to the resin-gel, the available concentration, *C*_{DGT}, and
114 the ratio, *R*, which indicates the extent of the depletion of soil pore water concentrations at the DGT interface are detailed in Hattab
115 et al. (2014). Finally, the mass of each metal accumulated in the resin-gel layer was determined after extraction of the resin gel by 1
116 mL of HNO₃ (Suprapure, Marck Darmstadt, Germany) 5% for 24h. This solution was further diluted 10 times with HNO₃ 2% and
117 analyzed by HR-ICP-MS (Element 2, Thermo Fischer) to determine metal (Cu, Cr and Zn) concentrations.

118

119 **2.4. Plant testings**

120

121 Plant testing's were carried out on the soil samples collected in 2010 and 2011 (4 plants/pot), using dwarf beans and a 2-
122 week growing period, to assess the PTTE phytoavailability. The culture and harvest conditions and the digestion of primary bean
123 leaves (BL) were described in Hattab et al. (2014). The Cu, Cr, As and Zn concentrations in the leaf digests were determined by
124 ICP-MS (Varian 810-MS) using standard solutions of trace elements diluted from a stock solution 1,000 ppm (±1 %/certified).
125 Foliar element concentrations are expressed in mg kg⁻¹ DW. The mineral mass of each element in the bean primary leaves was
126 computed based on its foliar elemental concentration and the biomass (DW) of primary leaves.

127

128 **2.5. Statistical analysis**

129

130 Total element concentrations in the SPW, DGT concentrations, *R* ratios, foliar element concentrations, mineral masses of
131 elements in the primary leaves and leaf DW yields were statistically analyzed by ANOVA (Statistica) to evaluate the influence of

132 increasing total soil Cu and soil amendment on the mobility, availability and phytoavailability of Cu, Cr, Zn, and As. All analytical
 133 determinations were performed in two replicates. Differences were considered significant if the p-value was $p < 0.05$. R2 was the
 134 determination coefficient of the linear regression curve.

135

136 3. Results and discussion

137

138 3.1. Soil and soil pore water characteristics

139 Total soil As, Cr, and Zn were in the common ranges for French sandy soils but total soil Cu exceeded its background level
 140 and threshold value for soil contamination and risk assessment, *i.e.* 35 mg Cu kg⁻¹ (Baize, 1997; Baize et al., 2002), for such coarse
 141 sandy soils (Tab. 1). For soils sampled in the field plots, total soil content (in mg kg⁻¹) varied between 5.3–6.9 for As, 16.2–19.4 for
 142 Cr, 35–74 for Zn and 163–1170 for Cu (Tab. 1). Total soil As, Cr, and Zn did not differ very much between the plots. Globally, Cu
 143 was the main contaminant of the topsoils. The physico-chemical characteristics (pH, DOC, EC, cations and anions) of the SPW, and
 144 the TOC of the soil samples are presented in Tab. 2.

145

146 Table 1: Total As, Cu, Cr and Zn concentrations in the topsoils (2010) adapted from Kolbas et al. (2011)

	mg kg ⁻¹			
plot#	Cu	Zn	Cr	As
T163	163	73.8	17.7	6.54
T268	268	58.1	16.4	5.33
T382	382	50.1	17.4	5.61
T518	518	50.7	19.4	6.90
T753	753	39.0	16.2	5.35
T832 (unt)	832	35.2	18.8	5.73
T1170	1170	59.8	16.3	6.19
Control soil ^a	21	51	18	3.60
BL ^b	3.2–8.4	17–48	14.1–40.2	1–25

147 ^aControl soil (Mench and Bes 2009).

148 ^bBL: background levels (Baize 1997; Baize and Tercé 2002). Bold letters indicate concentrations exceeding (>10%) background levels in French sandy soils.

149

150

151 The SPW pH decreased slightly from 2010 to 2011 for all soils (Tab. 2). The pH of the untreated soil was lower than that of
 152 the amended soils in 2010 and 2011, still reflecting the dolomitic limestone and compost addition in these soils in 2008. The control
 153 soil was slightly more alkaline than both the amended and untreated soils. The DOC and TOC values were little changes between
 154 2010 and 2011. However, the DOC and TOC values peaked in 2011 for the CTRL soil compared with other soils and values for the
 155 CTRL soil in 2010. The Unt soil had the lowest TOC values. The EC values in 2011 were higher than in 2010 for all samples except
 156 the T163 and T268 soils. Our results agreed with previous findings for this field trial (Kolbas et al., 2011), Singh et al. (2007) also
 157 investigated the effect of organic amendment on the aided phytoextraction and reported highest pH and TOC values in the amended
 158 soils.

159

160

161

162

Table 2: Chemical characteristics of the SPW and soils (2010 and 2011).

2010				
Plot	TOC (%)	pH	EC ($\mu\text{S cm}^{-1}$)	DOC (mg L^{-1})
T163	0.89±0.10	7.57±0.18	1923±210	66.24±0.01
T268	1.04±0.05	7.67±0.20	1111±190	47.66±0.12
T382	0.86±0.07	7.65±0.22	1076±122	81.3±0.13
T518	1.01±0.08	7.79±0.41	1404±132	47.64±0.16
T753	1.2±0.06	7.66±0.44	1006±111	45.01±0.02
T832 (unt)	0.59±0.04	6.48±0.19	839±98	46.35±0.04
T1170	1.07±0.05	7.74±0.21	1021±107	104.1±0.05
CTRL	0.63±0.09	7.95±0.18	574±65	63.06±0.09
2011				
T163	1.18±0.01	7.17±0.13	1423±176	67.86±0.07
T268	1±0.01	7.27±0.17	1011±123	55.23±0.35
T382	1.01±0.00	7.25±0.26	1676±201	39.51±0.12
T518	1±0.01	7.39±0.22	1504±232	46.2±0.10
T753	0.96±0.01	7.26±0.19	1206±182	48.45±0.08
T832 (unt)	0.72±0.01	5.88±0.24	1339±188	35.37±0.10
T1170	1.2±0.00	7.34±0.32	1321±123	34.65±0.04
CTRL	3.94±0.18	7.77±0.15	1074±198	124.2±0.64

163

164 3.2 Mobility of Cu, Cr, Zn and As

165

166

167

168

169

170

171

172

173

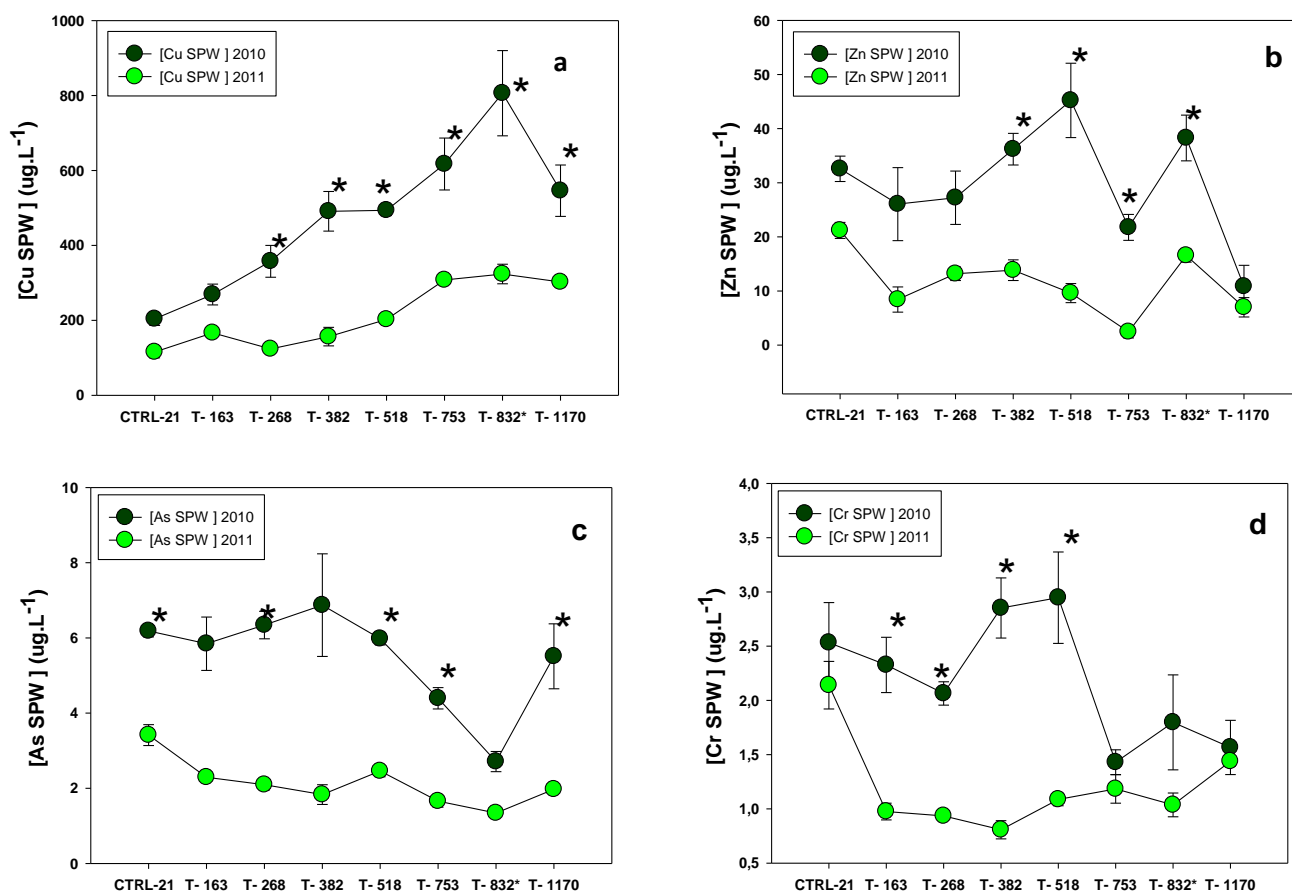
174

Concentrations of PTTE in SPW are relevant indicators of plant exposure to metals such as Cu for phytoextraction studies (Sauvé, 2003; Tandy et al., 2006; Forsberg et al., 2009). SPW samples from all tested soils showed a clear difference in the total dissolved concentration of PTTE between 2010 and 2011. The crop rotation on the tested plots, as well as annual leaching (Marchand et al. 2011), reduced the average concentrations of Cu, Zn, As, and Cr of all the treatments significantly from 2010 to 2011 by nearly 61%, 58%, 60% and 40% respectively (Fig. 2). For our field plots, Cu removal by shoots, capitula and seeds of sunflower in year 1 roughly reached 20-116 g Cu ha⁻¹ depending on soil Cu contamination and sunflower genotype (Kolbas et al., 2011). In year 2, Cu removal by the aboveground parts of sunflower was roughly 20 g Cu ha⁻¹ in the CTRL plot and increased from 16 (T268 soil) to 141 (T753 soil) g Cu ha⁻¹ in the contaminated plots (Mench, unpublished data). In year 3, shoot removals by aboveground parts of tobacco were 27 g (CTRL soil), 58 g (untreated soil) and 145-183 g (amended soils) Cu ha⁻¹ (Kolbas, 2012).

175 The total dissolved Cu concentrations in the SPW for the contaminated soils in 2010 and 2011 were higher than values for
 176 Zn, As, and Cr (Fig. 2) and the CTRL soil, i.e. 203.12 $\mu\text{g Cu L}^{-1}$ in 2010 and 115.13 $\mu\text{g Cu L}^{-1}$ in 2011 (Fig. 2.a). Total dissolved Cu
 177 concentrations in the SPW increased in 2010 and 2011 linearly with total soil Cu ($R^2 > 0.9$) confirming previous findings (Hattab et
 178 al., 2015). This agreed with Gonzalez et al. (2014), Kolbas et al. (2011) and Salati et al. (2010) who tested the efficiency of
 179 phytoextraction to reduce Cu excess in the soil pore water.

180 Total dissolved Zn concentrations in the SPW of the untreated and the control soil in 2011 were higher than those in the
 181 amended soils especially in 2011 (Fig. 2.b). Total dissolved Zn concentration in the SPW was correlated with the pH ($R^2 = 0.77$),
 182 DOC ($R^2 > 0.53$) and the TOC ($R^2 > 0.59$) of soil, 0.59 respectively) in 2011 but did not show any correlation with other soil and
 183 SPW parameters. Our results agreed with Hattab et al. (2014) who tested the effect of compost (OM) and dolomitic limestone (DL),
 184 singly and in combination, on the SPW Zn concentration of Zn. They found that Zn mobility was reduced in the amended plots as
 185 compared to the untreated and control soils.

186 Total dissolved Cr and As concentrations in the SPW of all tested soils significantly decreased between 2010 and 2011, except for
 187 Cr in the Unt soil and the T1170 soil (Fig. 2 c and d). Total dissolved Cr and As concentrations in the SPW in 2011 were lower
 188 ($p < 0.0001$) in contaminated soils compared with the control soil.



202 Figure 2: SPW concentrations of Cu, Zn, As and Cr measured in 2010 and 2011. Values are mean \pm standard error ($n=3$). * indicate
 203 a significant difference ($p < 0.05$).

204

205

206
207

3.3. Available soil Cu and Zn fractions

208

209

210

211

212

213

214

215

216

Data were obtained for available soil Cu and Zn fractions. Available Cu concentration in the contaminated soils varied from 116.123 to 11.25 $\mu\text{g L}^{-1}$ in 2010 and from 11.75 to 40.02 in 2011 (Fig. 3). This available fraction was generally low compared to total soil Cu, i.e. $<0.01\%$ in 2010 and $<0.007\%$ in 2011. This available soil Cu fraction increased with total soil Cu, except for the CTRL soil in 2010. For three out of eight soil samples, i.e. CTRL, T832 and T1170 soils, the available soil Cu fraction decreased in 2011 compared to 2010, but no clear influence of the crop rotation was marked on the whole soil series. The CTRL soil had lowest total soil Cu and both TOC and DOC increased in 2011 (Table 2) which may promote the Cu complexation. The T832 and T1170 soils had the highest total soil Cu but their soil pH decreased in 2011 which did not explain the decreased available soil Cu fraction. Available soil Cu, total soil Cu, total dissolved Cu concentration in the SPW and the SPW pH were correlated in 2011 ($R^2 > 0.6$). In contrast, no correlation was found between the soil and SPW parameters and the available soil Cu in 2010.

217

218

219

Available Zn concentrations ranged from 11.28 to 19.42 $\mu\text{g L}^{-1}$ in 2010 and from 1.29 to 12.26 in 2011 in the contaminated soils. They declined from 2010 to 2011, except in the T1170 soil. The available soil Zn represented a low fraction compared to total soil Zn, i.e. $<0.1\%$ in 2010 and $<0.04\%$ in 2011.

220

221

222

223

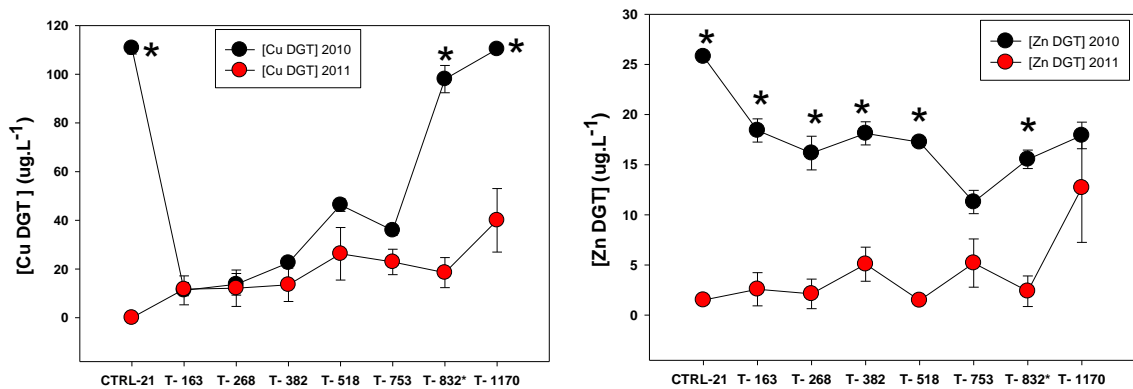
224

225

226

Available Zn and Cu concentrations in the soils were investigated in field plots managed by aided phytostabilisation at the same site 4 years after the incorporation of compost and dolomitic limestone, singly and in combination (OMDL) by Hattab et al. (2014) and similar results were found. The decrease in the available concentration of Cu can be explained by the formation of Cu-SOM complexes, particularly with non-soluble, high molecular mass organic acids which can decrease Cu phytoavailability (Balasoiu et al., 2001; Bolan and Duraisamy, 2003). The decrease in Zn availability in a contaminated soil can be explained by the presence of the organic and inorganic amendments, which can increase the precipitation and sorption of available Zn on the mineral phases (Lee et al., 2009).

227



228

229

Figure 3: Available Cu and Zn concentration in the soil samples measured by DGT in 2010 and 2011. Values are mean \pm standard error (n=3). * indicate a significant difference ($p < 0.05$).

230 **3.4. Phytoavailability of Cu, Cr, Zn and As**

231 **3.4.1. Concentration of PTTE in the bean primary leaves**

232 Generally the Cu, Cr, Zn and As concentrations in the primary leaves of beans (BL) were higher in 2010 than in 2011,
233 whereas the BL biomasses did not display general changes (Tab. 3). In average, these decreases were roughly 49% for Cu, 43% for
234 Cr, 70% for Zn and 22% for As. Our results agree with Zhou et al. (2015) reporting that the repeated phytoextraction by *Sedum*
235 *plumbizincicola* decreased the trace element phytoavailability in the soil especially for Zn.

236 Increase in total soil Cu enhanced the foliar Cu concentrations (Tab. 3). Data splitting according to two clusters for total soil
237 Cu values (i.e. 200-500 and 800–1200 mg Cu kg⁻¹) resulted in a linear relationship with the foliar Cu concentration in 2010 and
238 2011, such concentrations increased from 40.6±1.3 µg Cu L⁻¹ to 151.1±1.6 µg Cu L⁻¹ (UNT soil) in 2010 and from 22.8±0.3 µg Cu
239 L⁻¹ to 65.5 ±0.4 µg Cu L⁻¹ in 2011.

240 The highest foliar Cu concentrations in both 2010 and 2011 occurred in the untreated soil, showing likely the remaining
241 beneficial effects of the initial soil amendment and enhanced crop yields. Influence of soil amendments for reducing both soil Cu
242 exposure and plant Cu concentration was previously reported (Garrido et al., 2005). Hattab et al. (2014) investigated field plots
243 managed by aided phytostabilisation at this site. They also found that beans grown on the untreated soil had higher foliar Cu
244 concentrations than beans cultivated on the amended plots. Phytotoxic ranges of Cu for most plants are (in mg Cu kg⁻¹), e.g., 15–30
245 (MacNicol and Beckett, 1985), 25–40 (Chaney 1989), and 10–70 (Gupta and Gupta 1998). Accordingly the foliar Cu concentrations
246 of our plants exceed these upper critical threshold values, especially for beans from in the untreated soil which had the highest total
247 soil Cu. The foliar Cu concentrations were correlated with total soil Cu (R²=0.96 in 2010 and 0.99 in 2011), total Cu concentration
248 in the SPW (R² = 0.86 in 2010 and 0.94 in 2011) and the available soil Cu fraction (R²=0.53 in 2010 and 0.58 in 2011) confirming
249 previous findings with sunflower (Kolbas et al., 2011). Foliar Zn concentration in 2011 negatively correlated with total soil
250 Cu (R²= -0.88).Foliar Cr and Zn concentrations correlated with DOC and TOC in 2011 (R²> 0.65). Correlations were weak between
251 the foliar Cr, As and Zn concentrations measured in 2010 and 2011 and the other soil and plant parameters measured.

252

253

254

255

256

257

258

259

260

Table 3: Dry weight biomass (g DW plant⁻¹) and concentrations of Cu, Cr, Zn, and As (mg kg⁻¹) of the bean primary leaves.

	Dry weight (g) 2010	Dry weight g (2011)	[Cu] (mg Kg ⁻¹) 2010	[Cu] (mg Kg ⁻¹) 2011	[Zn] (mg Kg ⁻¹) 2010	[Zn] (mg Kg ⁻¹) 2011	[Cr] (mg.Kg ⁻¹) 2010	[Cr] (mg Kg ⁻¹) 2011	[As] (mg Kg ⁻¹) 2010	[As] (mg Kg ⁻¹) 2011
T163	0.12 ±0.00 ***	0.11±0.01 NS	40.6±1.3 ***	22.8±0.3 ***	72.0±0.5 ***	46.0±0.5 ***	2.83±0.13 ***	0.71±0.12 ***	0.42±0.01 ***	0.20±0.03 NS
T268	0.05±0.00 ***	0.10±0.00 NS	74.9±0.4 ***	29.0±0.1 ***	77.8±0.9 ***	49.3±0.3 ***	2.60±0.16 ***	1.00±0.01 ***	1.33±0.04 ***	0.16±0.01 NS
T382	0.21±0.01 ***	0.12±0.01 NS	60.1±0.01 ***	29.9±0.4 ***	56.2±0.8 ***	39.3±0.6 ***	2.17±0.09 ***	0.89±0.09 ***	0.70±0.00 ***	0.19±0.02 NS
T518	0.15±0.00 ***	0.11±0.01 NS	70.4±0.4 ***	40.3±0.2 ***	46.3±0.5 ***	38.8±0.3 ***	1.63±0.03 ***	1.20±0.08 ***	0.81±0.01 ***	0.25±0.01 NS
T753	0.09±0.00 ***	0.11±0.02 NS	142.5±2.1 ***	59.9±0.2 ***	68.9±1.2 ***	39.8±0.5 ***	1.29±0.03 ***	0.64±0.02 ***	1.17±0.06 ***	0.20±0.06 NS
T832 (unt)	0.12±0.01 ***	0.13±0.00 NS	173.1±1.6 ***	86.1±0.3 ***	40.4±0.4 ***	33.3±0.5 ***	1.47±0.05 ***	0.67±0.04 ***	1.34±0.06 ***	0.18±0.01 NS
T1170	0.09±0.00 ***	0.10±0.00 NS	151.1±1.6 ***	65.5±0.4 ***	67.7±1.5 ***	39.1±0.2 ***	3.75±0.09 ***	0.95±0.05 ***	1.13±0.05 ***	0.15±0.01 NS
CTRL	0.09±0.00 ***	0.10±0.00 NS	38.0±1.2 ***	19.5±0.3 ***	83.07±0.56 ***	53.65±0.07 ***	3.39±0.25 ***	1.46±0.09 ***	1.60±0.03 ***	0.23±0.02 NS

262 Effect of blocks: Significance level: NS : Not significant, * P<0.05, **P<0.01, *** P<0.001.

263

264 3.4.2 Mineral masses of Cu, Cr, Zn and As in the bean primary leaves

265 The mineral masses of Cu, Cr, As, and Zn in the primary leaves of beans cultivated in potted soils (mg/pot) was computed
 266 with foliar element concentrations (µg g⁻¹ DW) and the primary leaf biomass (g DW pot⁻¹) (Fig. 4). The mineral masses of Cu, Cr,
 267 As, and Zn decreased significantly from 2010 to 2011, especially for Cu. In average, the mineral masses decreased by 49% for Cu,
 268 55% for Cr, 78% for As, and only 34% for Zn.

269 Globally, the Cu mineral masses of bean primary leaves (Cu_{mm}) increased with total soil Cu, but peaked in the untreated soil
 270 due to its high Cu contamination and lower soil pH leading to a high foliar Cu concentration. Lower foliar Cu concentration and
 271 primary leaf DW yield explained the lower Cu_{mm} of beans on the T1170 soil compared to the untreated soil (T832). Total soil Cu
 272 and total Cu concentration in the SPW in 2010 and 2011 were correlated (R² >0.9) with Cu_{mm}.

273 The Zn mineral masses of bean primary leaves (Zn_{mm}) were mainly driven by the primary leaf DW yield (Fig. 4 and Tab. 3),
 274 explaining its peak for the T382 beans in 2010. Except for this last case, Zn_{mm} globally decreased as Cu_{mm} raised. The Cr mineral
 275 masses of bean primary leaves (Cr_{mm}) were lower than Cu_{mm} and Zn_{mm}. Two lower Cr_{mm} values in 2010 corresponded to low
 276 primary leaf DW yield. The As mineral masses of bean primary leaves (As_{mm}) were the lowest compared to Cu_{mm}, Zn_{mm}, and Cr_{mm}.
 277 The Cu_{mm} did not influence Cr_{mm} and As_{mm}. Lower Cr_{mm} and As_{mm} values in 2011 in overall reflected the lower SPW Cr and As
 278 concentrations in 2011 (Fig. 2). The Cr_{mm} showed a good correlation with total soil Cr and total Cr concentration in the SPW in
 279 2011 (R² >0.5). In contrast a negative correlation was found between As_{mm} in 2010 and the total soil As (R²= -57).

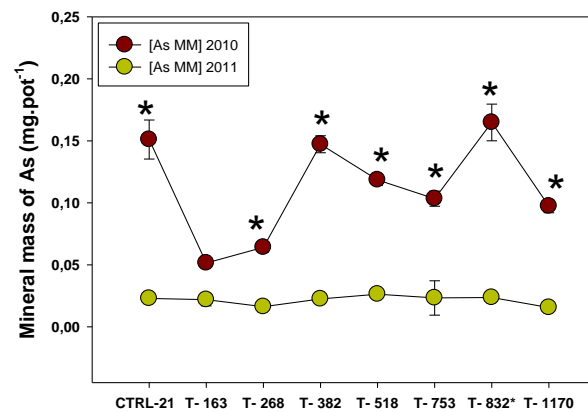
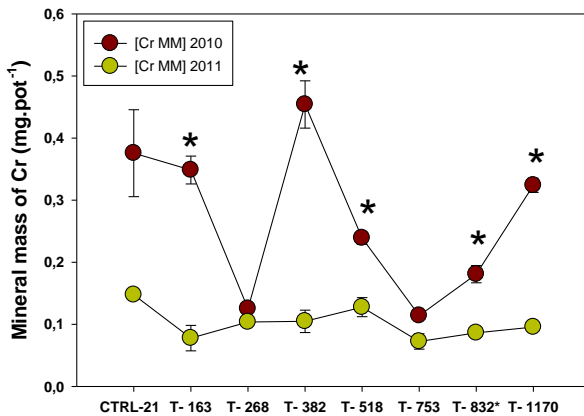
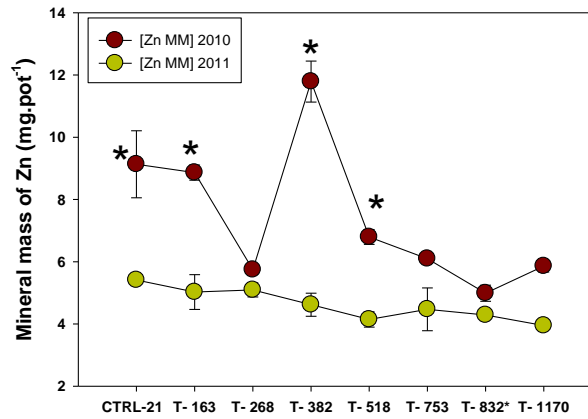
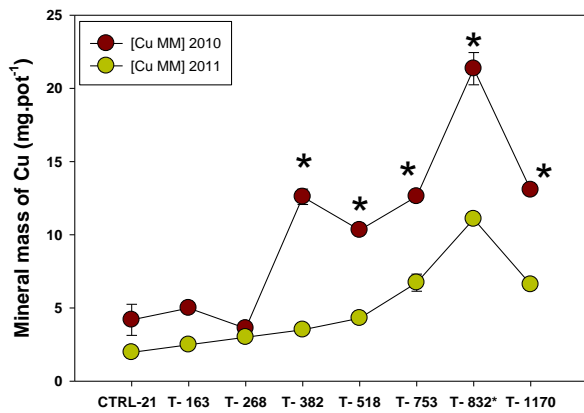


Figure 4: Mineral mass of Cu, Zn, Cr and As measured in the bean leaves. Values are mean \pm standard error (n=4). * indicate a significant difference ($p < 0.05$).

4. Conclusion

Changes in the mobility (using soil pore water), availability (using DGT) and phytoavailability of Cu, Zn, Cr and As (using plant testings with beans) were investigated at a wood preservation site, in topsoils of field plots, either amended or not with a single addition of dolomitic limestone (DL) and compost (OM), phytomanaged with a crop rotation of sunflower and tobacco, after two and three years. For purpose of comparison a similar uncontaminated control plot was also investigated. Total dissolved Cu concentrations in the SPW were higher than for Zn, As, and Cr. Total dissolved concentrations of Cu, Zn, Cr and As in the SPW decreased in year 3 as compared to year 2, likely due to shoot removals by the crop rotation and the lixiviation. Available soil Cu and Zn fractions also declined in year 3. For the contaminated field plots, available soil Cu fraction increased with the total soil Cu soil, but depended also on soil pH. The phytoavailability of Cu, assessed by the foliar Cu concentration and the Cu mineral mass of the primary leaves of beans, was reduced in year 3.

295 **References**

296

- 297 Baize, D., 1997. Un Point sur Les Teneurs Totales des Eléments Traces Métalliques dans les Sols, INRA Editions, Paris, France, pp
298 408.
- 299 Baize, D., Tercé, M., 2002. Les Éléments Traces Métalliques dans les Sols –Approches Fonctionnelles et Spatiales, INRA Éditions,
300 Paris, France. pp 570
- 301 Balasoiu, C.F., Zagury, G.J., Deschênes, L., 2001. Partitioning and speciation of chromium, copper, and arsenic in CCA-
302 contaminated soils: influence of soil composition. *Sci Total Environ* 280(1–3):239–255
- 303 Bes, C., Mench, M., 2008. Remediation of copper-contaminated topsoils from a wood treatment facility using in situ stabilisation.
304 *Environ Pollut.* 156:1128–1138.
- 305 Bes, C., Mench M., Aulen, M., Gasté H., Taberly, J. 2010. Spatial variation of plant communities and shoot Cu concentrations of
306 plant species at a timber treatment site. *Plant Soil* 330:267-280
- 307 Bolan, N.S., and Duraisamy, V.P. 2003. Role of inorganic and organic soil amendments on immobilisation and phytoavailability of
308 heavy metals: a review involving specific case studies. *Aust J Soil Res*, 41, 3, 533-555.
- 309 Bureau de Recherches Géologiques et Minières (BRGM). 1978. Graves Entre-deux-Mers, Geological Map of France 1/50000,
310 Pessac XV-37 (in French). BRGM, Orléans, France.
- 311 Chaney RL, 1989. Toxic element accumulation in soils and crops: protecting soil fertility and agricultural food-chains. In: Bar-Yosef
312 B, Barrow NJ, Goldshmid J, eds. *Inorganic contaminants in the vadose zone*. Berlin: Springer-Verlag. p. 140–158.
- 313 Davison, W., Fones, G., Harper, M., Teasdale, P., Zhang, H., 2000. Dialysis, DET and DGT: in Situ Diffusional Techniques for
314 Studying Water, Sediments and Soils. In: Buffle, J., Horvai, G. (Eds.). *In Situ Chemical Analysis in Aquatic Systems*, Wiley.
- 315 Efremova, M, Izosimova, A., 2012. Contamination of agricultural soils with heavy metals. In: Jakobsson C (ed) *Ecosystem health*
316 *and agriculture. Sustainable agriculture. The Baltic University Program*. Uppsala University, Uppsala, Sweden, pp 250–252
- 317 Ernstberger, H., Zhang, H. and Davison, W., 2002, Determination of chromium speciation in natural systems using DGT, *Anal.*
318 *Bioanal. Chem.* 373, 873–879.
- 319 Forsberg, LS; Kleja, DB; Greger, M; Ledin, S., 2009. Effects of sewage sludge on solution chemistry and plant uptake of Cu in
320 sulphide mine tailings at different weathering stages. *Appl Geochem.* 24:475–482.
- 321 Garrido, F., Illera, V., Garcia-Gonzalez, MT., 2005. Effect of the addition of gypsum- and lime-rich industrial by-products on Cd,
322 Cu and Pb availability and leachability in metal-spiked acid soils. *Appl Geochem* 20:397–408
- 323 Gonzalez, I., Neaman, A., Cortes, A., Rubio, P., 2014. Effect of compost and biodegradable chelate addition on phytoextraction of
324 copper by *Oenothera picensis* grown in Cu-contaminated acid soils. *Chemosphere* 95:111-115
- 325 Gupta UC, Gupta SC. 1998. Trace element toxicity relationships to crop production and livestock and human health: implications
326 for management. *Commun Soil Sci Plant Anal.* 29:1491–1522.
- 327 Hattab, N., Motelica-Heino, M., Bourrat, X., Mench, M., 2014. Mobility and phytoavailability of Cu, Cr, Zn, and As in a
328 contaminated soil at a wood preservation site after 4 years of aided phytostabilization. *Environ Sci Pollut Res.* 2014
329 21(17). p.10307
- 330 Hattab, N., Motelica-Heino, M., Faure, O., & Bouchardon, J. L., 2015. Effect of fresh and mature organic amendments on the
331 phytoremediation of technosols contaminated with high concentrations of trace elements. *Journal of Environmental*
332 *Management*, 159, 37-47.

333 Herzig R, Nehnevajova E, Pfister C, Schwitzguébel JP, Ricci A, Keller C (2014) Feasibility of labile Zn phytoextraction using
334 enhanced tobacco and sunflower: results of five- and one-year field-scale experiments in Switzerland. *Int J Phytoremediation*
335 16(7–8):735–754.

336 Keller, C., Hammer, D., Kayser, A., Richner, W., Brodbeck, M., and Sennhauser, M., 2003. Root development and heavy metal
337 phytoextraction efficiency: comparison of different plant species in the field. *Plant and Soil* 249, 67-81.

338 Kolbas, A., Mench, M., Herzig, R., Nehnevajova, E., Bes, C. M., 2011. Copper phytoextraction tandem with oilseed production
339 using commercial cultivars and mutant lines of sunflower. *Int J Phytoremediation*, 13: sup.1, 55-76.

340 Kolbas, A. 2012. Phenotypic traits and development of plants exposed to trace elements; use for phytoremediation and
341 biomonitoring. PhD thesis, Doctorate School Sciences & Environment, specialty Evolutionary, Functional and Community
342 Ecology, University of Bordeaux 1, Talence, France and University of Minsk, Belarus. 301 p.

343 Lee, SH., Lee, JS., Choi, YJ., Kim, JG., 2009. In situ stabilization of cadmium- lead- and zinc-contaminated soil using various
344 amendments. *Chemosphere* 77:1069–1075

345 Li, J.T., Baker, A.J.M., Ye, Z.H., Wang, H.B., Shu, W.S., 2012. Phytoextraction of Cd-contaminated soils: Current status and future
346 challenges. *Crit. Rev. Environ.Sci. Technol.* 42, 2113-2152.

347 MacNicol RD, Beckett PHT., 1985. Critical tissue concentrations of potentially toxic elements. *Plant Soil.* 85:107–129.

348 Marchand, L., Mench, M., Marchand, C., Lecoustumer, P., Kolbas, A., Maalouf, J.P., 2011. Phytotoxicity testing of lysimeter
349 leachates from aided phytostabilized Cu-contaminated soils using duckweed (*Lemna minor* L.). *Sci Total Environ*, 410, 146-153.

350 Mench, M, Bes, C., 2009. Assessment of ecotoxicity of topsoils from a wood treatment site. *Pedosphere.* 19:143–55

351 Mench, M., Lepp, N., Bert, V., Schwitzguébel, J.P., Gawronski, S.W., Schöder, P., Vangronsveld, J., 2010. Success and limitations
352 of phytotechnologies at field scale: outcomes, assessment and outlook from COST action 859. *J Soils & Sediments*, 10, 1039-
353 1070.

354 Mench M., Kolbas A., Atziria A., Herzig R., Marchand L., Maalouf J.-P., Ricci A. 2012. Field evaluation of one Cu-resistant
355 tobacco variant and its parental lines for copper phytoextraction at a wood preservation site. **Proc. 4th Int. Congress Eurosoil**
356 **2012**, Soil Science for the Benefice of Mankind and Environment, 12. Soil Pollution and Remediation, S12.08-Potentially
357 harmful elements in soils, Bari, July 2-6, Italy. p. 2542.

358 Nehnevajova, E., Herzig, R., Bourigault, C., Bangerter, S., Schwitzguébel, JP., 2009. Stability of enhanced yield and metal uptake
359 by sunflower mutants for improved phytoremediation. *Int J Phytorem.* 4:329–346.

360 Nehnevajova, E., Herzig, R., Erismann, KH., Schwitzguébel, JP., 2007a. In vitro breeding of *Brassica juncea* L. to enhance metal
361 accumulation and extraction properties. *Plant Cell Rep.* 26:429– 437.

362 Nehnevajova, E., Herzig, R., Federer, G., Erismann, KH., Schwitzguébel, JP., 2007b. Chemical mutagenesis – a promising
363 technique to increase metal concentration and extraction in sunflowers. *Int J Phytorem.* 9:149–165.

364 Salati, S., Quadri, G., Tambone, F., Adani, F., 2010. Fresh organic matter of municipal solid waste enhances phytoextraction of
365 heavy metals from contaminated soil. *Environ Pollut*, 158:1899–1906

366 Sauve, S 2003 Modelling trace element exposure and effects on plants. In Risk assessment and sustainable land management using
367 plants in trace element-contaminated soils. Ed. M Mench, Mocquot, B.69-70, Centre INRA Bordeaux-Aquitaine, Villenave
368 d’Ornon, France.

369 Schwitzguébel, J.P., Van der Lelie, D., Baker, A., Glass, D.J., Vangronsveld, J., 2002. Phytoremediation: European and American
370 trends, success, obstacles and needs. *J. Soils Sediments*, 2, 91-99.

371 Singh, SK., Juwarkar, A.A., Kumar, S., Meshram, J., Fan, M., 2007. Effet des modifications sur la phytoextraction de l'arsenic par
372 *Vetiveria zizanioides* du sol. *Int. J. Environ. Sci. Tech.*, 4 (3), 339-344.

373 Tandy, S., Schulin, R., Nowack, B., 2006. The influence of EDDS on the uptake of heavy metals in hydroponically grown sunflowers.
374 *Chemosphere* 62, 1454-1463.

375 Tangahu, B. V., Sheikh Abdullah, S. R., Basri, H., Idris, M., Anuar, N., & Mukhlisin, M. (2011). A review on heavy metals (As, Pb,
376 and Hg) uptake by plants through phytoremediation. *International Journal of Chemical Engineering*, 2011.

377 Vassilev, A., Schwitzguébel, J.P., Thewys, T., van der Lelie, D., and Vangronsveld, J., 2004. The use of plants for remediation of
378 metal-contaminated soils. *Sci World J Journal*, 4, 9-34.

379 Vassilev, A., Vangronsveld, J., and Yordanov, I., 2002. Cadmium phytoextraction: present state, biological backgrounds and
380 research needs. *Bulg. J. Plant Physiol.* 28(3-4), 68- 95.

381 Zhou, L., Wu, L., Li, Z., Yang, B., Yin, B., Luo, Y., & Christie, P., 2015. Influence of rapeseed cake on heavy metal uptake by a
382 subsequent rice crop after phytoextraction using *Sedum plumbizincicola*. *Int J Phytoremediation*, 17(1), 76-84.

383

384