

Co-binding of pharmaceutical compounds at mineral surfaces: mechanistic modeling of binding and co-binding of nalidixic acid and niflumic acid at goethite surfaces

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1	Co-binding of pharmaceutical compounds at mineral
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28 Abstract

Although emerging contaminants rarely exist individually in environmental 29 30 contaminated systems, only limited information on their adsorption mechanisms in multi-components solutions is currently available. To address this shortcoming, this 31 work examines for the first time the accuracy of a surface complexation model in 32 33 predicting the cooperative adsorption of nalidixic acid (NA) and niflumic acid (NFA) at 34 goethite (a-FeOOH) surfaces. Our model adequately predicts co-binding of an 35 outer-sphere (OS) complex of NFA onto NA bound to goethite through metal bonded (MB), hydrogen bonded (HB) or OS complexes. More positive charge is introduced in 36 the system via sodium interactions in order to describe the NFA adsorption at high NaCl 37 38 concentrations in both single and binary systems. Our model confidently predicts multilayers of NA on goethite as well as NFA binding on goethite-bound NA over a 39 40 large range of pH, salinity as well as NA and NFA loadings. These findings have strong implications in the assessment and prediction of contaminant fate in multi-component 41 42 contaminated systems by invoking a non-traditional form of ligand-ligand interaction 43 in this field of study.

44 Introduction

Although emerging contaminants (e.g. pharmaceutical and personal care products) 45 often exist in mixture in environmental systems^{1,2}, they have been often studied 46 individually with respect to the sorption and/or complexation with naturally occurring 47 subsurface solids^{3,4}. At environmental surfaces, the adsorption of multiple 48 contaminants (or pollutant mixtures) can give way to competitive or cooperative 49 binding. While competitive adsorption has been widely investigated⁵⁻⁷, very few 50 51 studies have resolved the impact of emerging contaminants co-binding at mineral surfaces. Very recently, we have demonstrated using vibrational spectroscopy and 52 quantum chemical calculations⁸, that nalidixic acid (NA, a quinolone antibiotic that is 53 widely used in humans and animals medicine⁹) and niflumic acid (NFA, a non-steroidal 54 anti-inflammatory that is often used for rheumatoid arthritis²) cooperatively adsorb to 55 56 naturally-occurring mineral particles by intermolecular interactions (van der Waals). This novel finding raised the importance of recognizing drug co-binding as a 57 58 mechanism contributing to the fate and transport of complex mixtures of antibiotics and anti-inflammatory agents in nature. Because most traditional environmental models are 59 based on a single/individual contaminant basis, little is known on their fate in mixed 60 contaminant systems. Omission of co-binding effects occurring in multi-component 61 solutions might have unknown consequences on the prediction of fate and transport of 62 these contaminants in the environment. 63

64 Some studies have used predictive models involving empirical adsorption isotherm 65 equations with limited (site specific) applicability to describe the observed data in

multi-component solutions^{10,11}. However, these macroscopic models cannot account 66 for co-binding mechanisms for accurately predicting the behavior of antibiotics at 67 mineral/water interfaces under a large range of physico-chemical conditions. In 68 contrast, mechanistic models include details of the surface (in terms of structure), take 69 into account the structure of the electrical interfacial layer 1^{12-14} , and constrain structure 70 and bonding of surface complexes by information from spectroscopic studies and/or 71 theoretical calculations^{15–17}. Although mechanistic models have extensively been 72 applied to predict pharmaceutical compounds adsorption to metal-(hydr)oxides^{18,19}, no 73 74 attempt has been made to account for co-binding mechanisms between two or more components. This becomes even more important considering the necessity to 75 adequately account for binding in reactive transport models to be used to predict the 76 77 fate of antibiotics and anti-inflammatory agents in nature.

In this study, we used molecular-level information (Attenuated Total 78 Reflectance-Fourier Transform InfraRed (ATR-FTIR) spectroscopy and quantum 79 chemical calculations) obtained from our previous work⁸, to develop a surface 80 complexation model to account for the pH and salinity dependence of NA and NFA 81 loadings achieved at goethite surfaces. We developed, for the first time, a model that 82 accounts for co-binding of ligands resulting from van der Waals-type interactions. This 83 opens new possibilities to account these generally overlooked forms of interactions in 84 these types of studies in the future. 85

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2. Materials and Methods

88 2.1. Chemicals

All chemicals used were of pro-analytical quality or better and were purchased from Sigma-Aldrich. Ultrapure water was used in all experiments. Goethite was prepared following Mazeina and Navrotsky's work²⁰. Goethite (α -FeOOH) was selected due to its great thermodynamic stability, high surface reactivity with organic and inorganic ligands, and abundance in soils and sediments²¹. It has been characterized in detail in our previous work⁸. The specific surface area was of $81 \pm 4 \text{ m}^2/\text{g}$ and the isoelectric point (IEP) is 9.1.

96

97 **2.3. Batch experiments**

The pH (3-10) and NaCl concentrations (3 - 300 mM) dependences of NA and 98 NFA binding to goethite was first investigated in single ([NA]_{tot} = 10 or 20 μ M or 99 $[NFA]_{tot} = 20 \ \mu M$) and binary $([NA]_{tot} = [NFA]_{tot} = 20 \ \mu M)$ systems. pH adsorption 100 101 edges in 10 mM NaCl and adsorption isotherms at pH 6 were taken from previous work⁸, while pH-dependence of NA and NFA adsorption in single and binary systems at 102 103 other NaCl concentrations (3, 100 and 300 mM) were conducted in the present work. 104 Because high background electrolyte concentrations can affect pH measurements, the pH electrode was calibrated using solutions of known $[H^+]$ (10⁻⁵-10⁻³ M) at different 105 106 [NaCl]. Therefore, pH refers to the molarity of the proton $(-\log [H^+])$. Adsorption 107 isotherms were obtained by experiments with (i) equimolar concentrations of NA and NFA (0.1 - 100 μ M), (ii) varying concentrations of NFA (0.1 - 100 μ M) with 20 μ M 108

109 NA, and (iii) varying concentrations of NA (0.1 - 100 μ M) with 20 μ M NFA. 110 Additionally, desorption tests conducted at pH = 11 were used to verify the mass 111 balance of NA and NFA in the suspensions. They showed that solutes were removed 112 only by adsorption, and that transformations by, for example, oxidation, did not occur 113 under our experimental conditions (a result that is consistent with previous 114 findings²²).

Batch experiments were conducted in 10 mL suspensions of 0.5 g/L goethite in polypropylene tubes under an atmosphere of $N_2(g)$ in order to avoid the effect of dissolved CO₂. Suspensions were mixed for a period of 8 h by end-over-end shaking, which is sufficient to reach a steady state⁸. Aliquots of the resulting suspensions were thereafter sampled and filtered (0.2 µm) for analysis. All adsorption experiments were performed at least twice, with a reproducibility of ~5% for NA- and ~10% for NFA-bearing systems.

Aqueous concentrations of NA and NFA were determined using high performance liquid chromatography (Waters 600 Controller) equipped with a photodiode array detector (Waters 996) and a reversed-phase C18 column (250 mm×4.6 mm i.d., 5 µm). The mobile phase (1 mL/min) was a mixture of acetonitrile/water (60/40 v/v) contained 0.1% formic acid. The detector was set to 258 nm for NA, 283 nm for NFA and both molecules could be analyzed with a single injection (NA: 4.5 min; NFA: 10.1 min).

128

129 **2.4. Surface complexation modeling**

130 The geochemical speciation code PHREEQC (version 2)²³ was used for surface

131	complexation calculations using thermodynamic database of Minteq. The pK_{as} of NA
132	(pK _a =6.19) and NFA (pK _{a,1} =2.28 and pK _{a,2} =5.10) at infinite dilution were obtained
133	from conditional pK_a values ^{24,25} and the Davies equation. NA exists as neutral (NAH,
134	$pH < pK_a$) or anionic form (NA ⁻ , $pH > pK_a$), and NFA as cationic (NFAH ₂ ⁺ , $pH < pK_{a,1}$),
135	anionic (NFA ⁻ , pH > pK _{a,2}), neutral (NFAH ⁰) or zwiterionic form (NFAH ^{+/-}) at pK _{a,1} <
136	$pH < pK_{a,2}$. Molecular structures of the different dissolved NA and NFA species and
137	calculated NA and NFA speciation versus pH in 10 mM NaCl solution are shown in
138	Figure S1. Protonation reactions of NA and NFA are given in Table 1.
139	The charge of the goethite/water interface was treated with using the three plane
140	model (TPM) ¹³ , where charges of the adsorbates can be distributed between the 0-
141	(metal-bonded complexes), 1- (hydrogen-bonded complexes), and 2-planes (Na ⁺ , Cl ⁻ ,
142	outer-sphere complexes). The TPM requires two capacitances for the 0- (C_1) and the
143	1-plane (C ₂) which were taken from a previous study in our group ²⁶ . Predictions of NA
144	and NFA adsorption to goethite were made using the multi-site complexation (MUSIC)
145	model ¹² . For goethite, different types of surface sites can exist depending of the
146	proportion of crystal planes, which should have different binding affinities. The
147	goethite surface is considered to exhibit (101), (001) and (210) planes (group Pnma),
148	which represent 63%, 27% and 10% of the total surface area, respectively. Singly
149	(=FeOH ^{-0.5}), doubly (=Fe ₂ OH) and triply (=Fe ₃ O ^{-0.5} and =Fe ₃ OH ^{+0.5}) coordinated
150	oxygens can be found at the goethite surface, depending on the crystal face. To simplify
151	the model, we adopted the 1-pK approximation of the MUSIC model ¹² , neglecting the
152	contributions of doubly- and part of the triply-coordinated oxygens. Crystallographic

site densities in this model are: $[=FeOH^{-0.5}]=3.03$ sites/nm² at the (101), 3.34 sites/nm² at the (001) and 7.4 sites/nm² at the (210) planes, and $[=Fe_3O^{-0.5}]=3.03$ sites/nm² at the (101) and 3.34 site/nm² at the (001) planes. The protonation constants of the groups are set to that of the IEP (1-pK approach¹⁴). Formation constants of all surface species are reported in Table 1.

NA and NFA are considered to bind only to \equiv FeOH^{-0.5} groups. From a molecular 158 159 viewpoint, ligands might also form H-bonds with doubly (=Fe₂OH) and triply $(\equiv Fe_3O^{-0.5} \text{ and } \equiv Fe_3OH^{+0.5})$ coordinated sites. Recently, Boily²⁷ showed that 160 161 protonation of doubly-coordinated hydroxo groups and one type of triply-coordinated oxo group, which are considered to be proton-silent within the 1-pK approach, may be 162 163 favored in the presence of strongly binding negatively charged ligand. However, a full version of the MUSIC model for goethite would be required to account for all these 164 165 sites. This would lead to a drastic increase in the number of parameters and then complexity in the model, and thus not considered in this modeling approach, as 166 generally carried out in literature^{17–19,26,28–30}. 167

The modeling strategy proceeded as follows. First, NA and NFA adsorption was modeled in single systems. The corresponding surface complexation constants were kept constant to model the NA-NFA binary system. All surface complexes reactions involved in binding and co-binding mechanisms were proposed based on our own molecular-level information and theoretical calculations and literature data, as detailed below in section 3.

174 PhreePlot³¹ was used to determine surface complexation constants and related

175 uncertainties for NA and NFA adsorption to goethite (single and binary systems). Charge variations at the 0-, 1- and 2-planes ($\Delta z_0, \Delta z_1, \Delta z_2$, respectively) are constrained 176 by spatial considerations and are not adjustable parameters. PhreePlot contains an 177 178 embedded version of the geochemical speciation program PHREEQC and it includes a 179 parameter optimization procedure, which automatically fits a model to experimental 180 data by minimizing the weighted sum of squares of the residuals. A modified Marquardt–Levenberg procedure was applied³². With this method, PhreePlot provides 181 182 also a statistical uncertainty of the estimated parameters, which is given for each of 183 optimized parameters in Table 1. Note that PHREEQC version 2 uses surface coverage fractions as standard state for surface species^{33,34}. 184

185

3. Results and discussion

3.1. Nalidixic acid (NA) adsorption on goethite (single system). As typically observed for many organic ligands, NA adsorption versus pH showed a bell-shape, and the maximum adsorption amount was observed at a pH near the pK_a (Fig. 1a). Increasing NaCl concentration decreased NA adsorption on the whole investigated pH range ([NA]_{tot} = 20 μ M; Fig. 1), an effect that is generally attributed to non-specifically bound ligands at mineral surfaces^{29,35}.

According to the spectroscopic investigations of NA binding to goethite⁸ and in agreement with previous studies on quinolone binding to minerals^{29,36}, MB surface complexes at the goethite (101)/(001) and (210) planes (Pnma group) were expected to 196 form *via* a NA keto group and one oxygen of the carboxylate group. This can be197 expressed through the surface complexation reaction:

198

199
$$2 \text{ H}^+ + 2 \equiv \text{FeOH}^{-0.5} + \text{NA}^- \rightleftharpoons (\equiv \text{Fe})_2(\text{NA})^0 + 2 \text{ H}_2\text{O}$$
; $\log^{\text{NA}}K_{\text{MB}}$ (1)

200

Only singly coordinated (\equiv FeOH^{-0.5}) surface sites were considered to be involved in the 201 202 latter reaction given the propensity for ligand exchange of these sites. Eq. 1 does not denote the number of Fe(III) octahedra in a complex, but only the number of \equiv FeOH^{-0.5} 203 sites that may or may not be of the same Fe(III) octahedron³⁰. Ideally, the steric 204 constraints at the dominant (101)/(001) planes (Pnma group) should promote bridging 205 between two Fe atoms separated by 3 Å from one another³⁷, while at the (210) plane, 206 two \equiv FeOH^{-0.5} should be located on the same Fe(III) octahedron, hence our preference 207 for modeling NA binding as a 1:2 NA:=FeOH^{-0.5} species (Table 1). 208 Hydrogen-bonded (HB) complexation (surface hydration-shared ion pair) was 209 also modeled through singly \equiv FeOH^{-0.5} sites with:³⁰ 210 211 $2 \text{ H}^+ + 2 \equiv \text{FeOH}^{-0.5} + \text{NA}^- \rightleftharpoons (\equiv \text{FeOH}_2)_2^{+\dots}(\text{NA})^- ; \log^{\text{NA}} K_{\text{HB}}$ (2)212 213 Formation of an outer-sphere (OS) complex (solvent-surface hydration-separated ion 214 pair), where NA keeps its first hydration sphere and its charge is located in the 2-plane, 215 was also considered to form with protonated singly-coordinated sites (\equiv FeOH₂^{+0.5}) 216

217 (Table 1).

218	As shown in Figure S2, our model satisfactorily predicts the pH dependence of NA
219	adsorption to goethite at low NA loading ([NA] _{tot} \leq 20 μ M; Fig. S2a,b,c), but
220	underestimated the NA adsorption isotherm at pH 6 at high NA loadings (Fig. S2d).
221	Furthermore, the effect of NaCl concentration on NA adsorption is not very well
222	captured in this model, since this effect is small for $[NA]_{tot} = 10 \ \mu M$ (Fig. S2a) but large
223	for $[NA]_{tot} = 20 \ \mu M$ (Fig. S2a,b). The corresponding highest adsorbed amount lies at
224	0.85 NA/nm ² (i.e. 1.4 μ mol/m ²), and did not exceed the estimated NA loadings of ~1.5
225	NA/nm ² that can be expected if all the ~ 3 –OH/nm ² were to be consumed. Attempts to
226	reproduce our data with the model of Boily ²⁷ (i.e. using a full version of the MUSIC
227	model for goethite including more surface sites for HB and OS complexes) led to
228	similar conclusions (not shown). We thus conclude that calculated NA adsorption
229	amount at high loading is not limited by the number of available surface sites, but by
230	less favorable electrostatic effects due to surface charge neutralization by NA.

In fact, higher loadings favor additional binding mechanisms (e.g. multilayer, dimerization, hydrophobic interactions and surface precipitation) involving direct NA-NA interactions. Drawing from our previous work on intermolecular interactions between naphthoic acids¹⁶, we can write:

235

236
$$2 \text{ H}^+ + 2 \equiv \text{FeOH}^{-0.5} + 2 \text{ NA}^- \rightleftharpoons (\equiv \text{Fe})_2 \text{NA}^{0...}(\text{NA})^- + 2 \text{ H}_2\text{O}; \log^{\text{NA}} K_{\text{dimer}}$$
 (3)

237

In this equation, a NA-NA dimer is formed by co-bonding to individual NA molecules,resulting in an interfacial distribution of charges between the 0- and 1-planes.

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Combining the equations with monomer and dimer of NA, the model better predictsNA adsorption at high loadings (Fig. 1d).

In summary, surface complexation modeling results (Table 1, Fig. 1 and Fig. S3), predict NA binding as a function of pH (Fig. 1a and Fig. S3 for $[NA]_{tot} = 20$ and 10 μ M, respectively), NaCl concentration (Fig. 1 a,c) and NA loadings (Fig. 1d). In accordance with previous reports on ligand binding (e.g. for benzenecarboxylates³⁰), NA predominantly formed HB and OS complexes at high pH while MB complexes only formed at pH < 7 (Fig. 1a). The model also predicts NA dimer formation with NA between pH 3 and 7 (Fig. 1 a,d).

249

250 3.2. Niflumic acid (NFA) adsorption on goethite (single system). NFA binding 251 followed the same pH dependence as NA (Fig. 1b) but is favored at NaCl concentration 252 greater than 50 mM (Fig. 1c). This alongside the weaker dependence of binding on 253 NFA loadings (Fig. 1d) suggests distinct binding mechanisms to those of NA. In fact, 254 based on the disposition of its functional groups, HB and OS complexation should 255 predominate over MB species under all conditions considered in this work, as for 256 example for other monocarboxylate ligands such as acetate, benzoate, cyclohexanecarboxylate or naphthoate^{17,35}. Both HB and OS complexes were 257 258 expressed by the following reaction:

259

260 $2 \text{ H}^+ + 2 \equiv \text{FeOH}^{-0.5} + \text{NFA}^- \rightleftharpoons (\equiv \text{FeOH}_2)_2^{+...} \text{NFA}^- ; \log^{\text{NFA}} K_{\text{HB/OS}}$ (4)

262	here omitting the intervening roles of water. The -1 charge of the HB complex of NFA
263	is located at the 1-plane ($\Delta z_1 = -1$; $\Delta z_2 = 0$) whereas that of the OS complex is in the
264	2-plane ($\Delta z_1 = 0$; $\Delta z_2 = -1$). This model adequately accounts for all goethite-NFA data,
265	except the effect of NaCl concentration. Alternatively, replacing the HB complex with
266	its zwitterion form provides both an adequate prediction of binding with pH and NaCl
267	concentration below pH 6, and is consistent with previous accounts ^{18,19,38} supporting
268	this type of species.
269	
270	$3 \text{ H}^{+} + 2 \equiv \text{FeOH}^{-0.5} + \text{NFA}^{-} \rightleftharpoons (\equiv \text{FeOH}_2)_2^{+} \text{NFAH}^0 ; \log^{\text{NFA}} K_{\text{HBH}}$ (5)
271	
272	Preliminary calculations showed that HB complexes with anionic or zwitterionic form
273	of NFA occurred in a comparable pH-range (not shown). To simplify the model, the
274	HB complex with the anionic form was removed from the speciation scheme. The OS
275	species is however needed to account for the high pH data at NaCl concentrations
276	below 10 mM. This model accurately predicts NFA adsorption at 10 mM NaCl but
277	provides a poor description of NaCl effect (Fig. S4). Because of the increasing
278	adsorption of NFA at high NaCl concentration, a sodium-OS ion pair is however
279	needed to account for the data above 10 mM NaCl through:
280	
281	$2 H^{+} + 2 \equiv FeOH^{-0.5} + NFA^{-} + Na^{+} \rightleftharpoons (\equiv FeOH_2)_2^{+\dots}NFA^{-\dots}Na^{+} ; \log^{NFA}K_{HBNa} $ (6)

where Na⁺ charge is located in the 2-plane. Sodium co-binding has been previously proposed to describe the adsorption of phosphate to goethite³⁹. This model allows a relatively good prediction of NFA adsorption to goethite at various pH (Fig. 1b), [NaCl] (Fig. 1b,c) and [NFA] (Fig. 1d).

287

288 **3.3. Binary NA-NFA system**

289 NA-NFA co-binding occurred over a large range of pH (Fig. 2a,b), NaCl 290 concentrations (Fig. 2a,b,e) and NA and NFA loadings (Fig. 2c,d). Co-binding effect is 291 more pronounced for NFA, because of its weaker adsorption in single system. 292 However, NA adsorption to goethite is rather strong in single system, and so co-binding 293 with NFA only further stabilizes NA at the surface. As a result, to accurately predict 294 binding in mixed NA-NFA system, the model requires reactions accounting of 295 cooperative effects between NA and NFA besides the competitive binding mechanisms 296 of eqs. 1-6. This is further illustrated in Figure S5, where NA and NFA adsorption in 297 binary system is strongly underestimated when using only eqs. 1-6.

For NA, all MB, HB and OS complexes of NA are considered to be formed because of the similar shape of the adsorption edges in mixtures with those of the single system (Figs. 1a and 2a), while only OS complexes are considered for NFA because the maximal loadings of NFA (Fig. 2b) coincided with that achieved in the single system (Fig. 1b). To keep the model at a reasonable level of complexity we however only considered the binary NA-NFA surface complex at the (101)/(001) planes and model co-binding with:

305
306
$$2 H^{+} + 2 \equiv FeOH^{-0.5} + NA^{-} + NFA^{-} \Rightarrow$$

307 $(\equiv Fe)_{2}(NA)^{0...}(NFA)^{-} + 2 H_{2}O$; log K_{MB-OS} (7)
308
309 $2 H^{+} + 2 \equiv FeOH^{-0.5} + NA^{-} + NFA^{-} \Rightarrow$
310 $(\equiv FeOH_{2})_{2}^{+...}(NA)^{...}(NFA)^{-}$; log K_{HB-OS} (8)
311
312 $2 H^{+} + 2 \equiv FeOH^{-0.5} + NA^{-} + NFA^{-} \Rightarrow$
313 $(\equiv FeOH_{2})_{2}^{+...}(NA)^{...}(NFA)^{-}$; log K_{OS-OS} (9)

In eq. 7-9, NFA binds to one NA at the goethite surface predominantly via van der 315 Waals interactions⁸, and its -1 charge is located at the 2-plane. In contrast, NA is 316 317 assumed to form MB, HB and OS complexes with goethite, respectively, and its -1 charge should therefore be located at the 0-plane ($\Delta z_0 = +1$; $\Delta z_1 = 0$; $\Delta z_2 = -1$, eq. 7), the 318 1-plane ($\Delta z_0 = +2$; $\Delta z_1 = -1$; $\Delta z_2 = -1$, eq. 8) or the 2-plane ($\Delta z_0 = +2$; $\Delta z_1 = 0$; $\Delta z_2 = -2$, eq. 9). 319 320 Note that the reverse case (NA binding to adsorbed NFA) involves the same 321 stoichiometry and charge distribution over the 0-, 1- and 2-planes. Therefore, they are 322 equivalent in PHREEQC. However, previous quantum chemical calculations⁸ 323 suggested that the reverse case was less plausible, that is, NA was expected to be located closer than NFA to the surface. 324

Our first modeling attempts using eqns. 1-9 predicted a decrease of NA/NFA adsorption with increasing [NaCl], which is inconsistent with experimental

327	observations (see Figure S6). Most notably, experimental NA binding became almost
328	independent of NaCl concentration (Fig. 2a,e) while NFA binding followed three
329	stages: (i) slight decrease in loadings below 10 mM, (ii) increased loadings at 10-50
330	mM, and (iii) constant loadings above 50 mM NaCl (Fig. 2b,e). Including of an ion pair
331	with sodium

333
$$2 \text{ H}^+ + 2 \equiv \text{FeOH}^{-0.5} + \text{NA}^- + \text{NFA}^- + \text{Na}^+ \rightleftharpoons$$

334
$$(\equiv FeOH_2)_2^{+\dots}(NA)^{-\dots}(NFA)^{-\dots}Na^+$$
; $\log K_{HB-HB-Na}$ (10)

335

considerably improved model predictions of the effect of NaCl (Fig. 2a,b,e). The best fit was obtained by allocating charges of both NA and NFA in the 1-plane and Na⁺ in the 2-plane ($\Delta z_0 = +2$; $\Delta z_1 = -2$; $\Delta z_2 = +1$) (Table 1).

The full model (i.e. eqs.1-10) provides an accurate description of co-binding data vs pH and NaCl concentration (Fig. 2a, b) and at a wide range of aqueous concentration (0.1-100 μ M) (Fig. 2c). This also permits to fit very well the variation of [NFA]_{ads} as a function of [NA]_{ads} regardless of the adopted approach : (i) varying [NA]_{tot} at constant [NFA]_{tot}, (ii) varying [NFA]_{tot} at constant [NA]_{tot} or (iii) varying equimolar concentration of both (Fig. 2d).

Model predictions of NA and NFA surface speciation for 10 mM NaCl (Figs. 2 a,b) suggest that NA-NFA dimers remarkably prevail at pH > 4. Although NA adsorption slightly increased from the single to the binary system, its surface speciation drastically

changes, probably due to competition between the three forms of NA (NA monomer,

NA dimer and NA-NFA dimer). Indeed, proportions of monomeric and dimeric forms of NA strongly diminished in favor of NA-NFA surface complexes (Fig. 2a, c). NFA adsorption was, however, mainly dominated by the formation of NA-NFA surface complexes (Fig. 2b, c).

353 Finally, our proposed model not only explains batch adsorption data generated for 354 this work but also concentration profiles of dominant species extracted by a chemometric analysis⁴⁰ of our previously published⁸ Fourier Transform Infrared 355 356 spectra on these systems (Fig. 2f). Our work had shown that vibration spectra of mixed 357 NA+NFA+goethite could not be explained as a linear combination of isolated 358 NA+goethite and NFA+goethite systems. The concentration profiles in Fig. 2f show that increasing fractions of NA-NFA complexed with increasing [NA]tot (constant 359 360 [NFA]tot) or with increasing [NFA]tot ([NA]tot) are well-predicted by the model. This 361 adds further confidence to the ability of our proposed model to account for NA and 362 NFA binding in single and mixed systems.

363

Implications for reactive transport modeling. Because of the increase of multiple contaminations in aquatic ecosystems worldwide, environmental subsurface and groundwaters often contain multiple contaminants including nonprescription drugs, antibiotics, hormones and prescription drugs⁴¹. Protection of groundwater resources requires an accurate assessment of processes controlling the contaminant fate and transport in complex environmental mixtures.

The development of new surface complexation models to account for co-binding at

371	mineral surfaces is central to the prediction of the fate of contaminants in
372	multi-component systems. We demonstrate, for the first time, that co-binding of
373	emerging organic contaminants (e.g. NA and NFA) can be predicted over a large range
374	of environmentally relevant conditions using a mechanistic model based on insights
375	from vibration spectroscopy and quantum chemical calculations ⁸ . The modeling
376	approach accounting for new forms of drugs co-binding at mineral surfaces is
377	becoming a powerful tool for implementation of co-binding phenomena in reactive
378	transport models, and for the accurate prediction of the transport of antibiotics and
379	anti-inflammatory agents in soils and sediments.
380	
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386	
386 387	Supporting information available

information is available free of charge via the Internet at <u>http://pubs.acs.org/</u>.

390

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512 **Table 1. Model parameters.**

	Complex	log K	Δz_0	Δz_1	Δz_2
$NA^{-} + H^{+} = NAH$		6.19			
$NFA^{-} + H^{+} = NFAH$		5.10			
$NFAH + H^+ = NFAH_2^+$		2.28			
$\equiv Fe_{3}O^{-0.5} + H^{+} \rightleftharpoons \equiv Fe_{3}OH^{+0.5}$		9.1	+1	0	0
$\equiv \mathrm{Fe_3O^{-0.5} + H^+ + Cl^-} \rightleftharpoons \equiv \mathrm{Fe_3OH_2^{+0.5}Cl^-}$		8.1	+1	0	-1
$\equiv Fe_{3}O^{-0.5} + Na^{+} \rightleftharpoons \equiv Fe_{3}OH^{-0.5}Na^{+}$		-1	0	0	+1
$\equiv \text{FeOH}^{-0.5} + \text{H}^+ \rightleftharpoons \equiv \text{FeOH}_2^{+0.5}$		9.1	+1	0	0
$\equiv \text{FeOH}^{-0.5} + \text{H}^+ + \text{Cl}^- \rightleftharpoons \equiv \text{FeOH}_2^{+0.5}\text{Cl}^-$		8.1	+1	0	-1
$\equiv FeOH^{-0.5} + Na^{+} \rightleftharpoons \equiv FeOH^{-0.5}Na^{+}$		-1	0	0	+1
$2 \text{ H}^+ + 2 \equiv \text{FeOH}^{-0.5} + \text{NA}^- \rightleftharpoons (\equiv \text{Fe})_2(\text{NA})^0 + 2 \text{ H}_2\text{O}$	MB	19.7±0.1	+1	0	0
$2 \text{ H}^+ + 2 \equiv \text{FeOH}^{-0.5} + \text{NA}^- \rightleftharpoons (\equiv \text{FeOH}_2)_2^{+}\text{NA}^-$	HB	20.0±0.1	+2	-1	0
$2 \text{ H}^+ + 2 \equiv \text{FeOH}^{-0.5} + \text{NA}^- \rightleftharpoons (\equiv \text{FeOH}_2)_2^{+}\text{NA}^-$	OS	20.8±0.1	+2	0	-1
$2 \text{ H}^+ + 2 \equiv \text{FeOH}^{-0.5} + 2 \text{ NA}^- \rightleftharpoons (\equiv \text{Fe})_2(\text{NA})^0 \dots \text{NA}^-$	dimer	22.2±0.1	+1	-1	0
$3 \text{ H}^+ + 2 \equiv \text{FeOH}^{-0.5} + \text{NFA}^- \rightleftharpoons (\equiv$	HBH	27.0±0.1	+2	0	0
FeOH ₂) ₂ ⁺ NFAH					
$2 \text{ H}^+ + 2 \equiv \text{FeOH}^{-0.5} + \text{NFA}^- \rightleftharpoons (\equiv$	OS	19.9±0.1	+2	0	-1
FeOH ₂) ₂ ⁺ NFA ⁻					
$2 H^+ + 2 \equiv FeOH^{-0.5} + NFA^- + Na^+$	HB-Na	20.8±0.1	+2	-1	+1
$\rightleftharpoons (\equiv \text{FeOH}_2)_2^{+\dots} \text{NFA}^{-\dots} \text{Na}^+$					
$2 H^+ + 2 \equiv FeOH^{-0.5} + NA^- + NFA^-$	MB-OS	23.2±0.2	+1	0	-1
\Rightarrow (=Fe) ₂ (NA) ⁰ ···NFA ⁻					
$2 H^+ + 2 \equiv FeOH^{-0.5} + NA^- + NFA^-$	HB-OS	23.2±0.2	+2	-1	-1
$\rightleftharpoons (\equiv \text{FeOH}_2)_2^+ \text{ NA}^- \text{NFA}^-$					
$2 H^+ + 2 \equiv FeOH^{-0.5} + NA^- + NFA^-$	OS-OS	25.4±0.2	+2	0	-2
$\rightleftharpoons (\equiv \text{FeOH}_2)_2^+ \text{ NA}^- \text{NFA}^-$					
$2 H^+ + 2 \equiv FeOH^{-0.5} + NA^- + NFA^- + Na^+$	HB-HB-Na	26.3±0.2	+2	-2	+1
$\rightleftharpoons (\equiv \text{FeOH}_2)_2^{+\dots}\text{NA-NFA}^{2\dots}\text{Na}^+$					

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TPM with $C_1=2.3 \text{ F/m}^2$, $C_2=1.07 \text{ F/m}^2$; 63% of (101), 27% of (001) and 10% of (210). Site densities: $[=\text{FeOH}^{-0.5}]=3.03$, 3.34 and 7.4 site/nm² at the (101), (001) and (210) planes, respectively; $[=\text{Fe}_3O^{-0.5}]=3.03$ and 3.34 site/nm² at the (101) and (001) planes, respectively. HB, OS and binary NA-NFA complexes are considered to form at all planes ((101)/(001)/(210)). NA and NFA surface complexation constants and uncertainty obtained with Phreeplot. All other parameters were fixed.

521 Figure captions

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523 **Figure 1.** Single systems. Adsorption of 20 μ M (a) NA and (b) NFA to goethite (0.5 g/L) versus pH and [NaCl] (3-100 mM). (c) Adsorption of 20 μ M NA or NFA (20 μ M) 524 525 on goethite (0.5 g/L) versus [NaCl] at pH 5. (d) NA-goethite and NFA-goethite adsorption isotherms at pH 6 in 10 mM NaCl, where NA surface speciation is shown 526 ("total" refers to overall modeled adsorption results). In all graphs, full lines correspond 527 to overall adsorption predicted by surface complexation modeling. In (a) and (b) NA 528 529 and NFA surface speciation is shown as discontinuous lines (for 10 mM NaCl). MB, HB and OS denote metal-bonded, hydrogen-bonded and outer-sphere complexes, 530 531 respectively.

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533 Figure 2. Binary NA-NFA system. Adsorption of (a) NA and (b) NFA to goethite (0.5 g/L) versus pH and [NaCl] (3-300 mM) for $[NA]_{tot} = [NFA]_{tot} = 20 \ \mu M$. (c) NA and 534 NFA adsorption isotherms at pH 6 in 10 mM NaCl for $[NA]_{tot} = [NFA]_{tot}$. (d) $[NFA]_{ads}$ 535 vs $[NA]_{ads}$ for three experimental conditions: (i) ($[NFA]_{tot} = 20 \ \mu M$, varying $[NA]_{tot}$, 536 537 black), (ii) ($[NA]_{tot} = 20 \mu M$, varying $[NFA]_{tot}$, red), and (iii) varying both compounds with $[NA]_{tot} = [NFA]_{tot}$ (blue). (e) Adsorption of NA and NFA ($[NA]_{tot} = [NFA]_{tot} = 20$ 538 μ M) on goethite (0.5 g/L) versus [NaCl] at pH 5. In all graphs, full lines correspond to 539 overall adsorption predicted by surface complexation modeling (using the same color 540 as the corresponding symbol). In (a), (b) and (c) NA and NFA surface speciation is 541 542 shown as discontinuous lines (for 10 mM NaCl). "total" refers to overall modeled adsorption results in (c). (f) Concentration profiles extracted by chemometric modeling 543 of FTIR data⁸ for the series corresponding to constant [NA]_{tot} and varying [NFA]_{tot} 544 (black) or constant [NFA]tot and varying [NA]tot (red). 545

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