

Vascular plant-mediated controls on atmospheric carbon assimilation and peat carbon decomposition under climate change

Konstantin Gavazov, Remy Albrecht, Alexandre Buttler, Ellen Dorrepaal, Mark Garnett, Sébastien Gogo, Frank Hagedorn, Robert T.E. Mills, Bjorn J.M. Robroek, Luca Bragazza

▶ To cite this version:

Konstantin Gavazov, Remy Albrecht, Alexandre Buttler, Ellen Dorrepaal, Mark Garnett, et al.. Vascular plant-mediated controls on atmospheric carbon assimilation and peat carbon decomposition under climate change. Global Change Biology, 2018, 24 (9), pp.3911-3921. 10.1111/gcb.14140. insu-01744835

HAL Id: insu-01744835 https://insu.hal.science/insu-01744835v1

Submitted on 28 Nov 2019

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Vascular plant-mediated controls on atmospheric carbon assimilation and

2	peat carbon decomposition under climate change
3	
4	Running head: Vascular plants and peat carbon cycling
5	
6	Konstantin Gavazov ^{1,2,3} , Remy Albrecht ^{1,2} , Alexandre Buttler ^{1,2,4} , Ellen Dorrepaal ³ , Mark H.
7	Garnett ⁵ , Sebastien Gogo ^{6,7,8} , Frank Hagedorn ⁹ , Robert T.E. Mills ^{1,2,10} , Bjorn J.M. Robroek ^{1,2,11} ,
8	Luca Bragazza ^{1,2,12}
9	
10	¹ Swiss Federal Institute for Forest, Snow and Landscape Research, WSL Site Lausanne, Station
11	2, 1015 Lausanne, Switzerland
12	² Ecole Polytechnique Fédérale de Lausanne EPFL, School of Architecture, Civil and
13	Environmental Engineering ENAC, Laboratory of ecological systems ECOS, Station 2, 1015
14	Lausanne, Switzerland
15	³ Climate Impacts Research Centre, Department of Ecology and Environmental Science, Umeå
16	University, 98107 Abisko, Sweden
17	⁴ Laboratoire de Chrono Environnement, UMR CNRS 6249, UFR des Sciences et Techniques,
18	Université de Franche-Comté, Besançon 25030, France
19	⁵ NERC Radiocarbon Facility (East Kilbride), Scottish Enterprise Technology Park, Rankine
20	Avenue, East Kilbride, Scotland, G75 OQF, United Kingdom
21	⁶ Université d'Orléans, ISTO, UMR 7327, 45071 Orléans, France
22	⁷ CNRS, ISTO, UMR 7327, 45071 Orléans, France
23	⁸ BRGM, ISTO, UMR 7327, BP 36009, 45060 Orléans, France

- ⁹Swiss Federal Institute for Forest, Snow and Landscape, Research, WSL Site Birmensdorf,
- 25 8903, Birmensdorf, Switzerland
- ¹⁰Lancaster Environment Centre, Lancaster University, Lancaster LA1 4YQ, United Kingdom
- ¹¹Biological Sciences, University of Southampton, Southampton SO17 1BJ, United Kingdom
- ¹²Department of Life Science and Biotechnologies, University of Ferrara, Corso Ercole I d'Este
- 29 32, I-44121 Ferrara, Italy

31 Correspondence

Luca Bragazza, Tel: +39 0532 293773, Email: luca.bragazza@unife.it

33

34 **KEYWORDS**

- 35 climate warming, decomposition, ecosystem respiration, net ecosystem CO₂ exchange, peatlands,
- 36 rhizosphere priming, vascular plant biomass, elevation gradient

37

38 Paper type: PRIMARY RESEARCH ARTICLE

40

41

42

43

44

45

46

47

48

49

50

51

52

53

54

55

Abstract

Climate change can alter peatland plant community composition by promoting the growth of vascular plants. How such vegetation change affects peatland carbon dynamics remains, however, unclear. In order to assess the effect of vegetation change on carbon uptake and release. we performed a vascular-plant removal experiment in two Sphagnum-dominated peatlands that represent contrasting stages of natural vegetation succession along a climatic gradient. Periodic measurements of net ecosystem CO₂ exchange revealed that vascular plants play a crucial role in assuring the potential for net carbon uptake, particularly with a warmer climate. The presence of vascular plants, however, also increased ecosystem respiration, and by using the seasonal variation of respired CO₂ radiocarbon (bomb-¹⁴C) signature we demonstrate an enhanced heterotrophic decomposition of peat carbon due to rhizosphere priming. The observed rhizosphere priming of peat carbon decomposition was matched by more advanced humification of dissolved organic matter, which remained apparent beyond the plant growing season. Our results underline the relevance of rhizosphere priming in peatlands, especially when assessing the future carbon sink function of peatlands undergoing a shift in vegetation community composition in association with climate change.

INTRODUCTION

56

57

58

59

60

61

62

63

64

65

66

67

68

69

70

71

72

73

74

75

76

77

78

Peatlands are formed under climatic, hydrological and biological conditions that suppress the decomposition of plant litter, resulting in the accumulation of organic matter as peat. The imbalance between litter production and decomposition makes peatlands effective long-term sinks of atmospheric carbon (Yu, 2012). Indeed, despite a distribution limited to c. 3% of the Earth's land surface, peatlands contain c. 30% of its soil carbon (Yu, 2012). In northern peatlands, Sphagnum mosses play a crucial role in peat accumulation due to the production of litter particularly resistant to microbial decomposition (Hájek et al., 2011). However, alteration of the hydrological balance (Talbot et al., 2014; Munir et al., 2015), increase in nitrogen availability (Bubier et al., 2007; Juutinen et al., 2010) and, more specifically, climate warming (Weltzin et al., 2000; Dieleman et al., 2015) can promote the growth and cover of vascular plants in peatlands. Such increase in the abundance of vascular plants can potentially reduce the productivity of Sphagnum mosses due to, for example, lower light availability (Chong et al., 2012) and increasing evapotranspiration (Hirano et al., 2016). In addition, considering that the decomposition rate of vascular plant litter is much faster than that of Sphagnum mosses (Dorrepaal, 2007), it is crucial to clarify the role of a higher vascular plant biomass, particularly with a warmer climate, with reference to the capability of the system to act as carbon sink (Breeuwer et al., 2010; Ward et al., 2013; Wang et al., 2015). From a belowground biogeochemical perspective, the presence of vascular plants can supply soil microbes with labile carbon substrates via rhizosphere exudates, providing additional energy to invest in decomposition of old organic matter (Fontaine et al., 2007; Schmidt et al., 2011): a process known as the priming effect (Kuzyakov, 2010). Nevertheless, it is still unknown whether the priming effect in peatlands is directly linked to the abundance of vascular plants, and whether

the priming effect shows any seasonal trend, particularly in relation to plant phenology. Answering these questions can help to better predict how climate warming will alter the carbon cycle of northern latitude peatlands when vascular plants become more abundant (Dieleman *et al.*, 2015) and the length of the growing season increases (Piao *et al.*, 2008).

To investigate the role of vascular plants in the seasonal net assimilation of atmospheric carbon and in the decomposition of peat carbon under contrasting climatic conditions, we performed a vascular plant-removal experiment in two *Sphagnum*-dominated peatlands that were located at different elevations and thereby naturally differed in mean annual temperature, growing season length and vascular plant abundance (Table 1). As such, the comparison of our study sites should be interpreted as a space-for-time substitution (Blois *et al.*, 2013), whereby the low elevation peatland represents the ecosystem state resulting from a natural response of vegetation to warmer climate compared to the upslope site (Klaus, 2007). During a full growing season, we periodically measured the net ecosystem CO₂ exchange (NEE), the ecosystem respiration (R_{eco}) and its radiocarbon signature, and the humification degree of dissolved organic matter in order to: 1) assess the role of vascular plants in assimilation of atmospheric CO₂ compared to *Sphagnum* mosses under contrasting climatic conditions; and 2) detect any temporal link between the presence of vascular plants and enhanced decomposition of peat through rhizosphere priming and the associated carbon release into both the atmosphere and pore water.

MATERIAL AND METHODS

Study sites and experimental design

We selected two *Sphagnum*-dominated peatlands located at 1035 m and 1885 m a.s.l. in Switzerland (Table 1, Table S1). At both sites, dominant vascular plant species were *Eriophorum*

vaginatum, Calluna vulgaris and Vaccinium spp., whereas Sphagnum magellanicum, S. capillifolium and S. fallax were the dominant peat moss species (Gavazov et al., 2016). Peat depth was estimated to be > 1.5 m.

102

103

104

105

106

107

108

109

110

111

112

113

114

115

116

117

118

119

120

121

122

123

124

At each site, six paired 40x40 cm plots were selected in midsummer 2013. To study the effect of vascular plants on peat decomposition, we clipped the aboveground biomass of vascular plants in half of the plots (-V treatment) without disturbing the moss layer, and left the belowground roots to decompose until measurements commenced the following year (cf. Ward et al., 2009; Kuiper et al., 2014). We started the field measurements after one year in order to allow the decomposition of most living roots, in particular the most labile fine roots. Due to the low rate of root decomposition in peatlands (Laiho et al., 2004; Moore et al., 2007), we are aware that root biomass did not completely decompose after one year, but the delay of one year in field measurements after biomass removal is, however, expected to have significantly reduced the belowground carbon allocation and root exudation in clipped plots. Vascular plants were left intact in paired control plots (+V treatment) so that our experimental set-up represents a pairedplot design with three replicates. Individual plots were delimited by 10 cm high quadratic stainless steel frames pushed to c. 5 cm depth in the living moss layer, which served as collars for gas flux measurements (see below). At regular intervals during 2013, all plots were trenched along the collars with a 30-cm long knife to avoid root colonisation. In the -V treatments vascular plant regrowth was regularly clipped in 2013, whereas no or very limited regrowth was observed in 2014. Because our primary aim was to study the biogeochemical role of vascular plants on carbon cycling, we decided to limit any secondary effects associated with vascular plant removal by shading the clipped plots (-V) with a UV-resistant mesh simulating the original vascular plant cover as preliminarily measured by photosynthetically active radiation (PAR) extinction

underneath the canopy (60% and 30% at low and high elevation, respectively). By doing so, the presence of a mesh avoided the desiccation of peat mosses, maintained the natural PAR intensity, and avoided changes in soil microclimate. During gas exchange measurements this mesh was instead used over the chamber to maintain the natural light intensity. Ancillary measurements of soil temperature and soil water content in the clipped and control plots at both elevations confirmed the suitability of this approach (Table S2).

Plant biomass and cover estimations

In each peatland, total aboveground vascular plant biomass was estimated by destructively harvesting five random 25x25 cm quadrats at peak standing biomass in 2013. Vascular plant cover within the experimental plots was estimated non-destructively with the point-intercept method. In brief, a pin was lowered over the vegetation through a 40x40 cm grid with 361 holes and the identity of the first intercepted plant was recorded. Species were attributed to one of the existing five categories: graminoids, shrubs, forbs, *Sphagnum spp.* mosses and standing litter, where the total surface cover equals 100%. Control plots (+V) as well as vascular plant removal plots (-V) had 100% *Sphagnum* cover.

CO₂ flux measurements

In 2014, one year after the start of the vascular plant removal treatment, a series of monthly CO₂ flux measurements were carried out during the snow-free period at each peatland starting in mid-April at 1035 m a.s.l. and in mid-June at 1885 m a.s.l. and lasting through to the end of October. CO₂ fluxes at ecosystem level were measured using a custom made transparent (95% light transmission) Plexiglas chamber (40x40 cm wide, 50 cm high) equipped with a diffusion infrared

gas analyser (GMP343, Vaisala, Finland) and a humidity-temperature probe (HMP57, Vaisala, Finland). Air inside the chamber was mixed with a system of inbuilt computer fans. The chamber had a rubber seal on the bottom and a small capped hole on the top which remained open when the chamber was placed on the permanent stainless-steel collar to avoid pressurisation. Net ecosystem CO₂ exchange (NEE) measurements were done in full light or and were immediately followed by ecosystem respiration (R_{eco}) measurements for which the chamber was darkened with a non-transparent cover. All flux measurements were carried out between 10 am and 12 am local time, ensuring optimal photosynthetic conditions for both Sphagnum and vascular plants (Bragazza et al., 2016) with vapour-pressure deficit (VPD) below 1.5 kPa and PAR intensity above the vegetation canopy between 1400 and 2100 µmol m⁻² s⁻¹. Each measurement lasted 1 minute in order to avoid excessive heat and humidity causing condensation inside the chamber. The CO₂ flux was calculated as a linear change in concentration (ppm), taking into account ambient atmospheric pressure and gas temperature. In this study, negative NEE represents a net CO₂ assimilation by the plant community and the calculated gross ecosystem productivity (GEP) was derived from the difference between R_{eco} and NEE.

163

164

165

166

167

168

169

170

148

149

150

151

152

153

154

155

156

157

158

159

160

161

162

Radiocarbon analysis and age interpretation

To determine the seasonal contribution of vascular plant-associated R_{eco}, we sampled respired CO₂ for radiocarbon analysis in both the clipped and unclipped plots in July and at the end of September 2014 (Table S3). We used a passive CO₂ sampling method to determine the radiocarbon signature of ecosystem respiration (Garnett *et al.*, 2009). In brief, specific PVC chambers (10 cm diameter, 10 cm height) connected to a zeolite molecular sieve cartridge (MSC) were gas-tight mounted on collars of similar size pre-installed in 2013 in each plot at 7 cm depth.

172

173

174

175

176

177

178

179

180

181

182

183

184

185

186

187

188

189

190

191

192

193

Prior to sample collection, the atmospheric CO₂ in the chamber headspace was scrubbed by pumping it at a constant rate (0.5 1 min⁻¹) through a soda lime column for 10 min while monitoring the concentration. The chambers were then left in place for 10 days until, based on CO₂ production rates, a sufficient amount of CO₂ (i.e. > 3 ml) was passively trapped on the MSCs by diffusion without the use of an external pump. The trapped CO₂ was thermally recovered (425 °C) while purging with high-purity nitrogen gas, converted to graphite by Fe/Zn reduction and analysed by acceleration mass spectrometry (AMS) at the NERC Radiocarbon Facility in Scotland, UK (Garnett & Murray, 2013). Subsamples of CO₂ were measured on a dual inlet stable isotope mass spectrometer (Thermo Fisher Delta V) to determine their δ^{13} C signature. In addition, we evaluated the apparent carbon accumulation rate at the two peatlands by splitting one 20-cm long peat core from each site into 5 cm segments. From these segments, vascular plant remnants were removed, after which the carbon content, peat density and peat age were determined (Table S4). After the combustion of these samples in a high-pressure bomb in the presence of oxygen, CO₂ was converted to graphite as above. The peat age was estimated by matching the ¹⁴C content with a known record (Levin et al., 2013) of atmospheric ¹⁴CO₂ in order to estimate the date of carbon fixation (Goodsite et al., 2001) according to Cali-Bomb program for post-bomb ¹⁴C samples (see Reimer *et al.*, 2004). Following convention (Stuiver & Polach, 1977), all radiocarbon data were corrected for mass dependent fractionation by normalising to δ^{13} CVPDB values of -25% using the measured δ^{13} C values, and reported as %Modern with reference to the Oxalic Acid international radiocarbon standard. These data (Table S3) were not subjected to further corrections for atmospheric air contamination during chamber enclosure (Czimczik et al., 2006; Natali et al., 2011; Hartley et al., 2012; Hicks Pries et al., 2013) as the fraction of atmospheric CO₂ in the recovered samples proved low and subsequent 14 C corrections remained within the AMS measurement uncertainty (± 0.5 %Modern).

194

195

196

197

198

199

200

201

202

203

204

205

206

207

208

209

210

211

212

213

214

To distinguish between vascular plant-derived respiration and respiration from peat induced by the presence of vascular plants (i.e. the rhizosphere priming effect), we made a commonpractice assumption that vascular plants respire contemporary carbon with a ¹⁴C signature identical to the current atmosphere (Subke et al., 2011; Hartley et al., 2012), whereas subsurface peat respires older carbon (Hardie et al., 2007), which has become very likely incorporated during the post-1963 bomb-14C spike at our studied sites (Table S3). Experimental evidence from peatlands dominated by herbaceous vegetation supports this assumption for vascular plant shoots (Czimczik et al., 2006; Hicks Pries et al., 2013), and further indicates only a small contribution of carbon fixed in the preceding year (i.e. up to 0.7 %Modern enrichment) for roots (Hicks Pries et al., 2013, 2015). A sensitivity analysis confirmed that our conclusions were robust regardless of whether we used current year atmospheric ¹⁴C values or those from the preceding year. We used data from the nearby Jungfraujoch (53°20′ N, 9°54′ W, 3580 m a.s.l.) long-term atmospheric ¹⁴C monitoring station (Levin et al., 2013) to calculate a mean seasonal value (June, July, August and September) for the expected vascular plant-derived ¹⁴C signature for the 2014 growing season, which was 102.91 %Modern ¹⁴C. Elevation differences between the study sites and the ¹⁴C monitoring station at the Jungfraujoch are expected not to affect the ¹⁴C values due to atmospheric mixing (Levin et al., 2008). To this value, we compared the ¹⁴C signature of the vascular plant-associated respiration (D_{V plant associated}) in our experiment. Following Hardie et al. (2009), the latter was calculated using a two pool mixing model:

215
$$D_{V \text{ plant associated}} = (D_{+V} \cdot R_{+V} - D_{-V} \cdot R_{-V})/(R_{+V} - R_{-V})$$

where R represents the rate of ecosystem respiration measured on the day of chamber enclosure and D its ¹⁴C signature in plots with (+V) and without (-V) vascular plants. The vascular plant-associated respiration in our experiment consisted of vascular plant-derived respiration and, potentially, of vascular plant-induced peat respiration. A significant deviation in the calculated ¹⁴C signature of vascular plant associated respiration from that of the expected vascular plant-derived ¹⁴C signature (ambient atmosphere) was interpreted as an extra source of heterotrophic peat respiration in the presence of vascular plants, i.e. the rhizosphere priming effect.

Pore-water sampling and chemical analyses

Representative pore-water samples were collected monthly during May-October 2014 from four permanent microporous (0.20 μm) rhizons (Rhizosphere Research Products, The Netherlands) installed in each plot at the rooting depth, i.e. from 5 to 15 cm below the surface. Concentration of ammonium (NH₄⁺) and nitrate (NO₃⁻) were obtained colorimetrically on a continuous-flow autoanalyser (SEAL Analytical, Germany), whereas total nitrogen (TN) and dissolved organic carbon (DOC) were determined by combustion on a Shimadzu analyser (TOC-V CPHCTNM-1). In order to characterise the quality of dissolved organic matter (DOM) and its level of humification, lyophilised subsamples were analysed on a Frontier FT–NIR/MIR Spectrometer (PerkinElmer, USA). The recorded spectra (4000 to 400 cm⁻¹) of diamond attenuated total reflectance provide information on the DOM functional groups aliphatics (2943 cm⁻¹ region), carboxylic acids and aromatic esters (1720 cm⁻¹ region), aromatics (1630 cm⁻¹ region), lignins (1510 cm⁻¹ region), phenols (1420 cm⁻¹ region), and polysaccharides (1090 cm⁻¹ region) (Niemeyer *et al.*, 1992; Biester *et al.*, 2014). During peat decomposition, substances enriched in carboxylic, aromatic and phenolic groups accumulate with respect to labile polysaccharides.

thereby increasing the specific ratios between these moieties (Biester *et al.*, 2014). Commonly referred to as humification indices, the ratios 2943/1090, 1720/1090, 1630/1090, 1510/1090, 1420/1090 can be used as an indication of the level of organic matter decomposition (Broder *et al.*, 2012; Hodgkins *et al.*, 2014).

Statistical analysis

Data on seasonal CO₂ fluxes and pore-water chemistry were analysed as a paired-plot design for the effect of vascular plant removal in each peatland. The hierarchical structure was coded in R3.3 (R Core Team, 2016) as a linear mixed-effects model using the *nlme* package (Pinheiro *et al.*, 2016), accounting for the spatial proximity of +V (control) and -V (clipped) pairs and the repeated measurements in individual plots. Assumptions of normality and homoscedasticity of the residuals in all final models were verified visually using diagnostic plots, and the simplest models were retained by pooling non-significant interactions with the error term. Comparisons between the radiocarbon signature of the atmosphere and the estimated vascular plant-associated CO₂ were done in R using One Sample t-tests for each site and sampling date.

RESULTS

The main hydrological (water-table depth) and hydro-chemical (water pH and electrical conductivity) parameters were comparable between the two peatlands and overall typical of ombrotrophic peatlands, i.e. rain-fed bogs (Table 1). The main differences between the two sites were related to climatic conditions and abundance of vascular plants, with characteristically higher air temperature, lower precipitation and higher shrub biomass at low elevation (Table 1, Table S1). Because the cover of peat mosses at plot level did not differ between the two sites

(100% at both sites), the study sites can be regarded as a natural climate change experiment where the natural climatic gradient resulted in similar plant community composition, but with different abundance of vascular plants in response to soil microclimate (Table 1, Table S2).

Overall, the net ecosystem exchange of atmospheric carbon (i.e., NEE) was weaker (i.e. smaller sink capacity) at low elevation with warmer climatic conditions ($F_{1,4} = 15.5$, P = 0.017) despite the larger vascular plant biomass (Fig. 1a). Nevertheless, in both peatlands the presence of vascular plants (+V treatment) increased ($F_{1,35} = 19.4$, P < 0.001) the net CO₂ assimilation by c. 50% compared to the treatment with *Sphagnum* mosses alone (-V treatment) (Fig. 1a). Furthermore, at low elevation the absence of vascular plants prevented a net assimilation of atmospheric carbon during most of the growing season (Fig. 1a).

During the peak of the growing season, vascular plants contributed to about 50% to gross photosynthesis at both elevations (Fig. 1b) as indicated by the two-fold reduction of potential gross ecosystem productivity (GEP) upon vascular plant removal ($F_{1,35} = 50.2$, P < 0.001). Although vascular plant biomass was significantly larger at low elevation (Table 1), the GEP was similar at both sites in the presence of vascular plants (+V). In the absence of vascular plants (-V), GEP of *Sphagnum* mosses was also comparable between the two sites (Fig. 1b).

Overall, ecosystem respiration (R_{eco}) was slightly larger at low elevation as compared to high elevation site ($F_{1,4} = 6.4$, P = 0.064) and at both sites the presence of vascular plants increased R_{eco} ($F_{1,35} = 46.9$, P < 0.001) (Fig. 1c). By taking into account the CO_2 flux and the corresponding ¹⁴C signature in plots with and without vascular plants, the two-pool mixing model revealed that late in the growing season the vascular plant-associated respiration was more enriched in bomb-¹⁴C than would be expected from autotrophic contributions alone at both low ($t_{1,2} = 8.8$, P = 0.006) and high ($t_{1,2} = 2.7$, P = 0.053) elevation (Fig. 2). An enrichment in bomb-

¹⁴C likely occurred also during the peak of the growing season in July (Fig. 2); however, it was masked by larger variability at both low ($t_{1,2} = 1.4$, P = 0.153) and high elevation ($t_{1,2} = 1.9$, P = 0.096).

On the basis of pore-water chemistry, the peatland at low elevation was characterized by a c. 30% greater concentration of dissolved organic carbon (DOC) (Table 1). Furthermore, the FT-IR spectroscopy demonstrated that the presence of vascular plants (+V) was associated with a consistently higher humification of dissolved organic matter in both peatlands. This was indicated by higher concentrations of aliphatic, carboxylic, aromatic, lignin, and phenolic compounds in proportion to more labile polysaccharides (Fig. 3).

DISCUSSION

Our net ecosystem exchange (NEE) measurements under optimal conditions and during the peak of diurnal photosynthetic activity provide an objective assessment of the potential contribution of vascular plants and *Sphagnum* mosses to the exchange of atmospheric CO₂ in the two contrasting peatlands (Bahn *et al.*, 2009; Bragazza *et al.*, 2016). The NEE results demonstrate that the presence of vascular plants is crucial to maintain high levels of net CO₂ assimilation in the two peatlands during the growing season (Laine *et al.*, 2012; Ward *et al.*, 2013; Kuiper *et al.*, 2014). *Sphagnum* mosses alone were much less efficient in offsetting ecosystem respiratory CO₂ losses under warmer conditions. Concurrently, gross ecosystem productivity (GEP) in both peatlands was much higher in the presence of vascular plants, in accordance with the higher photosynthetic efficiency of vascular plants compared to mosses (Ward *et al.*, 2009; Armstrong *et al.*, 2015). Despite the larger aboveground plant biomass at lower elevation, GEP was not significantly different between the two peatlands, a result that can be at least partly explained by a larger

309

310

311

312

313

314

315

316

317

318

319

320

321

322

323

324

325

326

327

328

329

abundance of shrubs with a greater portion of ligneous, non-photosynthetic tissues at low elevation (Table 1: Fig. S1) (Bubier et al., 2003: Leppälä et al., 2008: Juutinen et al., 2010), On the basis of our field measurements, Sphagnum mosses in the two peatlands contributed equally to GEP per surface area, indicating that under suitable conditions of temperature and moisture the photosynthetic potential was the same at both elevations even if the annual productivity can be different (Bragazza et al., 2013). Indeed, albeit a longer growing season (Fig. 1), both vascular plants and *Sphagnum* mosses are likely to experience more limiting photosynthetic conditions at lower elevation due to higher VPD values (Table 1). The overall greater R_{eco} in the presence of vascular plants, particularly at low elevation, is likely due to a higher contribution of autotrophic respiration, in line with: the larger vascular plant biomass (Ward et al., 2013; Kuiper et al., 2014; Hicks Pries et al., 2015), the corresponding larger biomass of decomposing roots (Murphy et al., 2009), as well as to an enhancement of heterotrophic respiration, i.e. a boosting of microbial metabolism. The latter can be directly stimulated by the improved soil physical conditions due to warmer climate (Updegraff et al., 2001; Dorrepaal et al., 2009; Conant et al., 2011; Philben et al., 2014) or indirectly promoted by vascular plants through the rhizosphere priming. The two-pool mixing model, taking into account the CO₂ efflux and the corresponding ¹⁴C signature in plots with and without vascular plants at each peatland, shows a consistent enrichment of vascular plant-associated respiration with older carbon, a result that clearly indicates a stimulation of heterotrophic peat decomposition through the rhizosphere priming effect (Hardie et al., 2009; Basiliko et al., 2012; Walker et al., 2016).

Although we cannot exclude a contribution of residual decomposing roots to the priming (even

after one year of decomposition), if this should take place in the clipped plots it means that the

differences in the ¹⁴C enrichment with the control plots would be still larger in total absence of roots in the peat.

A sustained expression of the rhizosphere priming towards the end of the growing season is intriguing because it indicates that the priming can still be active beyond the plant growing season, i.e. when soil and air temperatures as well as GEP already decline (Fig. 1, Table S2). We interpret this apparent temporal asynchrony between aboveground plant productivity and belowground microbial decomposition as result of: i) the low thermal diffusivity of peat that maintained favourable soil temperatures later into the growing season (Table S2), ii) the preferential allocation of carbon resources by vascular plants to their roots well beyond their aboveground senescence (Abramoff & Finzi, 2015; Blume-Werry *et al.*, 2016), and iii) the leaching from fresh leaf litter (Singh *et al.*, 2014) providing soil microbes with labile carbon substrates to access older organic matter (Qiao *et al.*, 2014).

On the basis of FT-IR spectroscopy, at both studied sites there was a higher humification degree of dissolved organic matter in plots with intact vascular plants (Fig. 3), a result that seems to reflect an enhanced microbial metabolism (Glatzel *et al.*, 2003; Höll *et al.*, 2009; Ward *et al.*, 2013; Lou *et al.*, 2014; Dieleman *et al.*, 2016). The removal of vascular plants is indeed expected to reduce the availability of more labile, root-mediated carbon (Gogo *et al.*, 2011) that is used by microbes for synthesising extracellular enzymes (Robroek *et al.*, 2016). Additionally, we observe that warmer climatic conditions and larger vascular plant biomass are ultimately associated with a larger release of dissolved organic carbon (DOC) in the peat soil at low elevation (Table 1). This latter result points to the synergetic role of climate and vegetation in increasing the production and release of DOC from peatlands (Neff & Hooper, 2002; Frey & Smith, 2005; Ward *et al.*, 2013; Kane *et al.*, 2014).

354

355

356

357

358

359

360

361

362

363

364

365

366

367

368

369

370

371

372

373

374

375

On the basis of the ¹⁴C signature in the subsurface peat (Table 1), the estimated age of top 20 cm peat, particularly for the low elevation site, was in line with other studies (see e.g. Shotyk et al., 1997) suggesting a relatively recent peat accumulation, i.e. primarily after the bomb- ¹⁴C peak of 1963. We conclude that the carbon released by the priming effect was only a few decades old (< 20 years) and stored above the mean water-table depth (Table 1). This observation highlights the importance of water table in protecting peat from microbial decomposition (Ise et al., 2008), but also indicates the potential vulnerability of peat carbon if hydrological feedbacks to climate change increase soil oxygenation (Waddington et al., 2015) and, consequently, promote the spreading of vascular plants (Dieleman et al., 2015; Potvin et al., 2015). The apparent rate of carbon accumulation in the peat layers above the mean-water table depth approximated c. 90 g C m⁻² y⁻¹ for the low elevation site and c. 154 g C m⁻² y⁻¹ for the high elevation site over a reference period of 26-30 years for both sites, a result suggesting a lower carbon sink capacity of peatlands at the lower end of their altitudinal distribution (Millar et al., 2017). The effective annual contribution of the rhizosphere priming to the observed difference in carbon accumulation rate in relation to the dominant vascular plant functional types and their symbiotic microorganisms remains an open question that deserves further attention (Gavazov et al., 2016; Walker et al., 2016; Sulman et al., 2017). Our CO₂ exchange measurements, whilst covering only a single growing season, show that the presence of vascular plants is crucial to sustain high potential levels of net atmospheric carbon assimilation in peatlands, particularly with warmer climate. Differently, Sphagnum mosses alone could not keep pace with the observed higher ecosystem respiratory C-losses with warmer climate. A sustained temporal expression of the rhizosphere priming effect, although deserving

further investigation (Luo et al., 2016), suggests that the decomposition of older peat can be

promoted even beyond the plant growing season, so affecting the humification of peat and loss of C in both gaseous and dissolved form. We argue that to accurately predict the future capacity of peatlands to act as atmospheric carbon sinks the synergetic interactions among vegetation dynamics, plant phenology and soil microbes must be taken into account, particularly if the rhizosphere priming effect can be further stimulated in warmer soils (Zhu & Cheng, 2011), thereby exerting a positive feedback to carbon emissions from organic rich soils.

ACKNOWLEDGEMENTS

We thank Lucas Wettstein and Adrian Pulgarin for field assistance and to Vincent Jassey and Laurent Jeanneau for laboratory assistance. Thanks are also due to Ingeborg Levin and Samuel Hammer for providing data on ¹⁴C signature of atmospheric CO₂. The Service des Forêts, de la Faune et de la Nature (Canton de Vaud) and the Service de la promotion de la nature – Office de l'agriculture et de la nature (Canton de Berne) are acknowledged for giving the permission to access the studied peatlands. This work was supported by the Swiss National Science Foundation grant number: 205320 144024 to L.B.

REFERENCES

- Abramoff RZ, Finzi AC (2015) Are above- and below-ground phenology in sync? New
- *Phytologist*, **205**, 1054–1061.
- Armstrong A, Waldron S, Ostle NJ, Richardson H, Whitaker J (2015) Biotic and abiotic factors
- interact to regulate northern peatland carbon cycling. *Ecosystems*, **18**, 1395–1409.
- Bahn M, Schmitt M, Siegwolf R, Richter A, Brüggemann N (2009) Does photosynthesis affect
- grassland soil-respired CO₂ and its carbon isotope composition on a diurnal timescale? *New*

399	Phytologist, 182 , 451–460.
400	Basiliko N, Stewart H, Roulet NT, Moore TR (2012) Do root exudates enhance eeat
401	decomposition? Geomicrobiology Journal, 29, 374–378.
402	Biester H, Knorr K-H, Schellekens J, Basler A, Hermanns Y-M (2014) Comparison of different
403	methods to determine the degree of peat decomposition in peat bogs. Biogeosciences, 11,
404	2691–2707.
405	Blois JL, Williams JW, Fitzpatrick MC, Jackson ST, Ferrier S (2013) Space can substitute for
406	time in predicting climate-change effects on biodiversity. Proceedings of the National
407	Academy of Sciences, 110 , 9374–9379.
408	Blume-Werry G, Wilson SD, Kreyling J, Milbau A (2016) The hidden season: growing season is
409	50% longer below than above ground along an arctic elevation gradient. New Phytologist,
410	209 , 978–986.
411	Bragazza L, Parisod J, Buttler A, Bardgett RD (2013) Biogeochemical plant-soil microbe
412	feedback in response to climate warming in peatlands. Nature Climate Change, 3, 273–277.
413	Bragazza L, Buttler A, Robroek BJM, Albrecht R, Zaccone C, Jassey VEJ, Signarbieux C (2016)
414	Persistent high temperature and low precipitation reduce peat carbon accumulation. Global
415	Change Biology, 22 , 4114–4123.
416	Breeuwer A, Heijmans M, Robroek BJM, Berendse F (2010) Field Simulation of Global Change
417	Transplanting Northern Bog Mesocosms Southward. <i>Ecosystems</i> , 13 , 712–726.
418	Broder T, Blodau C, Biester H, Knorr KH (2012) Peat decomposition records in three pristine
419	ombrotrophic bogs in southern Patagonia. <i>Biogeosciences</i> , 9 , 1479–1491.
420	Bubier J, Crill P, Mosedale A, Frolking S, Linder E (2003) Peatland responses to varying
421	interannual moisture conditions as measured by automatic CO ₂ chambers. Global

Biogeochemical Cycles, 17, no. 2, 1066. 422 423 Bubier JL, Moore TR, Bledzki LA (2007) Effects of nutrient addition on vegetation and carbon cycling in an ombrotrophic bog. Global Change Biology, 13, 1168–1186. 424 Chong M, Humphreys E, Moore TR (2012) Microclimatic response to increasing shrub cover and 425 426 its effect on Sphagnum CO₂ exchange in a bog. Écoscience, 19, 89–97. Conant RT, Ryan MG, Ågren GI et al. (2011) Temperature and soil organic matter 427 decomposition rates - synthesis of current knowledge and a way forward. Global Change 428 Biology, 17, 3392–3404. 429 Czimczik CI, Trumbore SE, Carbone MS, Winston GC (2006) Changing sources of soil 430 respiration with time since fire in a boreal forest. Global Change Biology, 12, 957–971. 431 Dieleman CM, Branfireun BA, Mclaughlin JW, Lindo Z (2015) Climate change drives a shift in 432 peatland ecosystem plant community: Implications for ecosystem function and stability. 433 Global Change Biology, 21, 388–395. 434 Dieleman CM, Lindo Z, McLaughlin JW, Craig AE, Branfireun BA (2016) Climate change 435 effects on peatland decomposition and porewater dissolved organic carbon biogeochemistry. 436 *Biogeochemistry*, **128**, 385–396. 437 Dorrepaal E (2007) Are plant growth-form-based classifications useful in predicting northern 438 ecosystem carbon cycling feedbacks to climate change? *Journal of Ecology*, **95**, 1167–1180. 439 Dorrepaal E, Toet S, van Logtestijn RSP, Swart E, van de Weg MJ, Callaghan T V., Aerts R 440 (2009) Carbon respiration from subsurface peat accelerated by climate warming in the 441 subarctic. *Nature*, **460**, 616-U79. 442 Fontaine S, Barré P, Bdioui N, Mary B, Rumpel C (2007) Stability of organic carbon in 443 deep soil layers controlled by fresh carbon supply. *Nature*, **450**, 277–80. 444

Frey KE, Smith LC (2005) Amplified carbon release from vast West Siberian peatlands by 2100. 445 Geophysical Research Letters. 32. L09401. 446 Garnett MH, Murray C (2013) Processing of CO₂ samples collected using zeolite molecular sieve 447 for ¹⁴C analysis at the NERC Radiocarbon Facility (East Kilbride, UK). *Radiocarbon*, 55, 448 449 410-415. Garnett MH, Hartley IP, Hopkins DW, Sommerkorn M, Wookey PA (2009) A passive sampling 450 method for radiocarbon analysis of soil respiration using molecular sieve. Soil Biology and 451 Biochemistry, 41, 1450–1456. 452 Gavazov K, Hagedorn F, Buttler A, Siegwolf R, Bragazza L (2016) Environmental drivers of 453 carbon and nitrogen isotopic signatures in peatland vascular plants along an altitude 454 gradient. *Oecologia*, **180**, 257–264. 455 Glatzel S, Kalbitz K, Dalva M, Moore T (2003) Dissolved organic matter properties and their 456 relationship to carbon dioxide efflux from restored peat bogs. Geoderma, 113, 397–411. 457 Gogo S, Laggoun-Défarge F, Delarue F, Lottier N (2011) Invasion of a Sphagnum-peatland by 458 Betula spp and Molinia caerulea impacts organic matter biochemistry. Implications for 459 carbon and nutrient cycling. *Biogeochemistry*, **106**, 53–69. 460 Goodsite ME. Rom W. Heinemeier J et al. (2001) High-resolution AMS ¹⁴C dating of post-bomb 461 peat archives of atmospheric pollutants. Radiocarbon, 43, 495–515. 462 Hájek T, Ballance S, Limpens J, Zijlstra M, Verhoeven JTA (2011) Cell-wall polysaccharides 463 play an important role in decay resistance of *Sphagnum* and actively depressed 464 decomposition in vitro. *Biogeochemistry*, **103**, 45–57. 465 Hardie SML, Garnett MMH, Fallik AE et al. (2007) Spatial variability of bomb ¹⁴C in an upland 466 peat bog. *Radiocarbon*, **49**, 1055–1063. 467

Hardie SML, Garnett MH, Fallick AE, Ostle NJ, Rowland AP (2009) Bomb-14C analysis of 468 469 ecosystem respiration reveals that peatland vegetation facilitates release of old carbon. *Geoderma*, **153**, 393–401. 470 Hartley IP, Garnett MH, Sommerkorn M et al. (2012) A potential loss of carbon associated with 471 472 greater plant growth in the European Arctic. *Nature Climate Change*, **2**, 875–879. Hicks Pries CE, Schuur EAG, Crummer KG (2013) Thawing permafrost increases old soil and 473 autotrophic respiration in tundra: Partitioning ecosystem respiration using δ^{13} C and Δ^{14} C. 474 Global Change Biology, 19, 649–661. 475 Hicks Pries CE, van Logtestijn RSP, Schuur EAG, Natali SM, Cornelissen JHC, Aerts R, 476 Dorrepaal E (2015) Decadal warming causes a consistent and persistent shift from 477 heterotrophic to autotrophic respiration in contrasting permafrost ecosystems. Global 478 *Change Biology*, **21**, 4508–4519. 479 Hirano T, Yamada H, Takada M, Fujimura Y, Fujita H, Takahashi H (2016) Effects of the 480 expansion of vascular plants in *Sphagnum*-dominated bog on evapotranspiration. 481 Agricultural and Forest Meteorology, **220**, 90–100. 482 Hodgkins SB, Tfaily MM, McCalley CK et al. (2014) Changes in peat chemistry associated with 483 permafrost thaw increase greenhouse gas production. *Proceedings of the National Academy* 484 of Sciences, 111, 5819-5824. 485 486 Höll BS, Fiedler S, Jungkunst HF, Kalbitz K, Freibauer A, Drösler M, Stahr K (2009) Characteristics of dissolved organic matter following 20 years of peatland restoration. 487 Science of The Total Environment, 408, 78–83. 488 Ise T, Dunn AL, Wofsy SC, Moorcroft PR (2008) High sensitivity of peat decomposition to 489 climate change through water-table feedback. *Nature Geoscience*, 1, 763–766. 490

491	Juutinen S, Bubier JL, Moore TR (2010) Responses of vegetation and ecosystem CO ₂ exchange
492	to 9 years of nutrient addition at Mer Bleue bog. Ecosystems, 13, 874–887.
493	Kane ES, Mazzoleni LR, Kratz CJ, Hribljan JA, Johnson CP, Pypker TG, Chimner R (2014) Peat
494	porewater dissolved organic carbon concentration and lability increase with warming: A
495	field temperature manipulation experiment in a poor-fen. <i>Biogeochemistry</i> , 119 , 161–178.
496	Klaus G (2007) État et évolution des marais en Suisse. Résultats du suivi de la protection des
497	marais. Etat de l'environnment no 0730. Office fédéral de l'environnement, Berne,
498	Switzerland, 97 pp.
499	Kuiper JJ, Mooij WM, Bragazza L, Robroek BJM (2014) Plant functional types define magnitude
500	of drought response in peatland CO ₂ exchange. <i>Ecology</i> , 95 , 123–131.
501	Kuzyakov Y (2010) Priming effects: Interactions between living and dead organic matter. Soil
502	Biology and Biochemistry, 42, 1363–1371.
503	Laiho R, Laine J, Trettin CC, Finér L (2004) Scots pine litter decomposition along drainage
504	succession and soil nutrient gradients in peatland forests, and the effects of inter-annual
505	weather variation. Soil Biology and Biochemistry, 36, 1095–1109.
506	Laine AM, Bubier JL, Riutta T, Nilsson MB, Moore TR, Vasander H, Tuittila E-S (2012)
507	Abundance and composition of plant biomass as potential controls for mire net ecosytem
508	CO ₂ exchange. <i>Botany</i> , 90 , 63–74.
509	Leppälä M, Kukko-Oja K, Laine J, Tuittila E-S (2008) Seasonal dynamics of CO ₂ exchange
510	during primary succession of boreal mires as controlled by phenology of plants. Écoscience,
511	15 , 460–471.
512	Levin I, Hammer S, Kromer B, Meinhardt F (2008) Radiocarbon observations in atmospheric
513	CO ₂ : Determining fossil fuel CO ₂ over Europe using Jungfraujoch observations as

514	background. Science of the Total Environment, 391, 211–216.		
515	Levin I, Kromer B, Hammer S (2013) Atmospheric $\Delta^{14}CO_2$ trend in Western European		
516	background air from 2000 to 2012. Tellus B: Chemical and Physical Meteorology, 65.		
517	Lou XD, Zhai SQ, Kang B, Hu YL, Hu LL (2014) Rapid response of hydrological loss of DOC		
518	to water table drawdown and warming in Zoige peatland: Results from a mesocosm		
519	experiment. PLoS ONE, 9, 19–21.		
520	Luo Z, Wang E, Sun OJ (2016) A meta-analysis of the temporal dynamics of priming soil carbon		
521	decomposition by fresh carbon inputs across ecosystems. Soil Biology Biochemistry, 10, 96-		
522	103.		
523	Millar DJ, Cooper DJ, Dwire KA, Hubbard RM, von Fischer J (2017) Mountain peatlands range		
524	from CO ₂ sinks at high elevations to sources at low elevations: Implications for a changing		
525	climate. <i>Ecosystems</i> , 20 , 416–432.		
526	Moore TR, Bubier JL, Bledzki L (2007) Litter decomposition in temperate peatland ecosystems:		
527	The effect of substrate and site. <i>Ecosystems</i> , 10 , 949–963.		
528	Munir TM, Perkins M, Kaing E, Strack M (2015) Carbon dioxide flux and net primary		
529	production of a boreal treed bog: Responses to warming and water-table-lowering		
530	simulations of climate change. <i>Biogeosciences</i> , 12 , 1091–1111.		
531	Murphy MT, McKinley A, Moore TR (2009) Variations in above- and below-ground vascular		
532	plant biomass and water table on a temperate ombrotrophic peatland. <i>Botany</i> , 87 , 845–853.		
533	Natali SM, Schuur EAG, Trucco C, Hicks Pries CE, Crummer KG, Baron Lopez AF (2011)		
534	Effects of experimental warming of air, soil and permafrost on carbon balance in Alaskan		
535	tundra. Global Change Biology, 17, 1394–1407.		
536	Neff JC, Hooper DU (2002) Vegetation and climate controls on potential CO ₂ , DOC and DON		

537	production in northern latitude soils. <i>Global Change Biology</i> , 8 , 872–884.
538	Niemeyer J, Chen Y, Bollag J (1992) Characterization of humic acids, composts, and peat by
539	diffuse reflectance fourier-transform infrared-spectroscopy. Soil Science Society of America
540	Journal, 56 , 135–140.
541	Philben M, Kaiser K, Benner R (2014) Does oxygen exposure time control the extent of organic
542	matter decomposition in peatlands? Journal of Geophysical Research: Biogeosciences, 119,
543	897–909.
544	Piao S, Ciais P, Friedlingstein P et al. (2008) Net carbon dioxide losses of northern ecosystems in
545	response to autumn warming. <i>Nature</i> , 451 , 49–52.
546	Pinheiro J, Bates D, DebRoy S, Sarkar D, R Core Team (2016) nlme: Linear and Nonlinear
547	Mixed Effects Models. R package version 3.1-128.
548	Potvin LR, Kane ES, Chimner RA, Kolka RK, Lilleskov EA (2015) Effects of water table
549	position and plant functional group on plant community, aboveground production, and peat
550	properties in a peatland mesocosm experiment (PEATcosm). Plant and Soil, 387, 277-294.
551	Qiao N, Schaefer D, Blagodatskaya E, Zou X, Xu X, Kuzyakov Y (2014) Labile carbon retention
552	compensates for CO ₂ released by priming in forest soils. <i>Global Change Biology</i> , 20 , 1943–
553	1954.
554	R Core Team (2016) R: A language and environment for statistical computing.
555	Reimer PJ, Brown, TA, and Reimer, RW (2004) Discussion: Reporting and calibration of post-
556	bomb 14C data. Radiocarbon, 46, 1299-1304.
557	Robroek BJM, Albrecht RJH, Hamard S, Pulgarin A, Bragazza L, Buttler A, Jassey VE (2016)
558	Peatland vascular plant functional types affect dissolved organic matter chemistry. Plant and
559	Soil, 407 , 135–143.

560	Schmidt MWI, Torn MS, Abiven S et al. (2011) Persistence of soil organic matter as an
561	ecosystem property. <i>Nature</i> , 478 , 49–56.
562	Shotyk W, Cheburkin AK, Appleby PG, Fankhauser A, Kramers JD (1997) Lead in three peat
563	bog profiles, Jura Mountains, Switzerland: enrichment factors, isotopic composition, and
564	chronology of atmospheric deposition. Water Air and Soil Pollution, 100, 297–310.
565	Singh S, Inamdar S, Mitchell M, McHale P (2014) Seasonal pattern of dissolved organic matter
566	(DOM) in watershed sources: Influence of hydrologic flow paths and autumn leaf fall.
567	Biogeochemistry, 118, 321–337.
568	Stuiver M, Polach HA (1977) Reporting of ¹⁴ C data. Discussion. <i>Radiocarbon</i> , 19 , 355–363.
569	Subke JA, Voke NR, Leronni V, Garnett MH, Ineson P (2011) Dynamics and pathways of
570	autotrophic and heterotrophic soil CO ₂ efflux revealed by forest girdling. Journal of
571	Ecology, 99, 186–193.
572	Sulman BN, Brzostek ER, Medici C, Shevliakova E, Menge DNL, Phillips RP (2017) Feedbacks
573	between plant N demand and rhizosphere priming depend on type of mycorrhizal
574	association. Ecology Letters, 20, 1043–1053.
575	Talbot J, Roulet NT, Sonnentag O, Moore TR (2014) Increases in aboveground biomass and leaf
576	area 85 years after drainage in a bog. Botany, 92, 713-721.
577	Updegraff K, Bridgham SD, Pastor J, Weishampel P, Harth C (2001) Response of CO ₂ and CH ₄
578	emissions from peatlands to warming and water table manipulation. Ecological
579	Applications, 11, 311–326.
580	Waddington JM, Morris PJ, Kettridge N, Granath G, Thompson DK, Moore PA (2015)
581	Hydrological feedbacks in northern peatlands. <i>Ecohydrology</i> , 8 , 113–127.
582	Walker TN, Garnett MH, Ward SE, Oakley S, Bardgett RD, Ostle NJ (2016) Vascular plants

583	promote ancient peatland carbon loss with climate warming. Global Change Biology, 22,
584	1880–1889.
585	Wang H, Richardson CJ, Ho M (2015) Dual controls on carbon loss during drought in peatlands
586	Nature Climate Change, 5 , 584–588.
587	Ward SE, Bardgett RD, McNamara NP, Ostle NJ (2009) Plant functional group identity
588	influences short-term peatland ecosystem carbon flux: Evidence from a plant removal
589	experiment. Functional Ecology, 23, 454–462.
590	Ward SE, Ostle NJ, Oakley S, Quirk H, Henrys PA, Bardgett RD (2013) Warming effects on
591	greenhouse gas fluxes in peatlands are modulated by vegetation composition. <i>Ecology</i>
592	Letters, 16, 1285–1293.
593	Weltzin JF, Pastor J, Harth C, Bridgham SD, Updegraff K, Chapin CT (2000) Response of bod
594	and fen plant communities to warming and ware table manipulations. <i>Ecology</i> , 81 , 3464–
595	3478.
596	Yu ZC (2012) Northern peatland carbon stocks and dynamics: A review. Biogeosciences, 9,
597	4071–4085.
598	Zhu B, Cheng W (2011) Rhizosphere priming effect increases the temperature sensitivity of soil
599	organic matter decomposition. Global Change Biology, 17, 2172–2183.
600	

TABLE 1 Geographical location, climatic conditions for the year 2014 and the average for the period 2005-2013 (in brackets), cumulative hours of vapour-pressure deficit (VPD) and water-table depth during plant growing season 2014, vascular plant biomass and surface cover (%), pore-water chemistry, and estimated age of surface peat (with corresponding 14 C signature) at the two study sites. Aboveground plant biomass is the mean of 5 replicates for each site. Water table depth is the seasonal mean of continuous measurements collected in one representative piezometer during the plant growing season at each site. Plant cover estimates are mean (\pm 1 SE) of 3 replicates for each site. Pore-water chemistry is the seasonal mean (\pm 1 SE) of 3 replicates for each site as measured in the control plots where vascular plants were not clipped (+V treatment). Different superscripts indicate significant differences between the peatlands (P < 0.05). The age of the peat was calculated from a reference dataset of atmospheric 14 C measurements carried out at the Jungfraujoch station and refers to years before 2014.

	Praz Rodet	Hochrajen
Elevation (m a.s.l.)	1035	1885
Latitude and longitude	46°33'N, 6°10'E	46°36'N, 7°58'E
Mean annual air temperature (°C)	7.0 (6.2)	3.7 (3.3)
Mean annual precipitation (mm)	1219 (1264)	1427 (1372)
VPD > 1.5 kPa (hours)	150	53
Mean water-table depth (cm)	-17.8	-15.6
Total aboveground vascular plant biomass (g m ⁻²)	$160^{a} \pm 21$	$74^{\mathrm{b}}\pm7$
Total vascular plant cover (%)	$63.0^{a} \pm 2.0$	$35.4^{b} \pm 5.3$
Graminoids (%)	$26.5^{a} \pm 3.3$	$19.0^{a} \pm 3.4$
Shrubs (%)	$20.0^{a} \pm 2.6$	$8.3^{\rm b} \pm 1.8$
Forbs (%)	$1.2^{a} \pm 0.5$	$1.8^{a} \pm 1.8$
Standing litter (%)	$15. 2^a \pm 1.4$	$6.3^{\rm b} \pm 1.9$
Total moss cover (%)	$100^{a} \pm 0.0$	$100^{a} \pm 0.0$
pH	$4.25^a \pm 0.02$	$4.37^a \pm 0.15$
Conductivity (µS cm ⁻¹)	$31.03^a \pm 0.64$	$27.81^a \pm 7.34$
Total N (mg l ⁻¹)	$0.39^{a} \pm 0.03$	$0.30^{b} \pm 0.02$
$NH_4^+ \text{ (mg l}^{-1}\text{)}$	$0.018^a \pm 0.003$	$0.007^{b} \pm 0.002$
NO_3^- (mg l ⁻¹)	$0.034^a \pm 0.002$	$0.032^a \pm 0.001$
DOC (mg l ⁻¹)	$22.14^{a} \pm 1.12$	$16.57^{\rm b} \pm 1.53$
Age at 5-10 cm (years / $\%$ Mod ¹⁴ C \pm 1 σ)	$16 / 110.23 \pm 0.48$	$1 / 103.41 \pm 0.47$
Age at 10-15 cm (years / $\%$ Mod ¹⁴ C ± 1 σ)	$30 / 120.75 \pm 0.53$	$16 / 110.01 \pm 0.48$
Age at 15-20 cm (years / $\%$ Mod ¹⁴ C \pm 1 σ)	$40 / 139.28 \pm 0.61$	$26 / 117.12 \pm 0.51$

FIGURE CAPTIONS

Fig. 1 Seasonal dynamics of ecosystem CO_2 fluxes in relation to the presence or absence of vascular plants in two peatlands at different elevations. (a) Net ecosystem exchange (NEE), (b) Gross ecosystem productivity (GEP), (c) Ecosystem respiration (R_{eco}). The two peatlands were located at low (1035 m a.s.l.) and high elevation (1885 m a.s.l.) and two treatments were applied, i.e. mosses plus vascular plants present (+V, controls) or removal of vascular plants by clipping (-V, *Sphagnum* mosses alone). Panels on the left show the seasonal dynamics of mean values (\pm 1 SE, n = 3), whereas those on the right show the overall seasonal means (\pm 1 SE, n = 3). Asterisks indicate significant differences of mean CO_2 fluxes between treatments at each site and between elevations ('*' = P < 0.1; * = P < 0.05; *** = P < 0.001). Negative values of NEE indicate a net assimilation of atmospheric CO_2 .

Fig. 2 14 C content of ecosystem respired CO₂ associated with the presence of vascular plants in two peatlands at low and high elevations (1035 and 1885 m a.s.l., respectively) at peak biomass (July) and late in the growing season (end of September). Median and range (n = 3) of respired 14 C-CO₂ associated with the presence of vascular plants (DV_plant_associated). Dotted line indicates the mean atmospheric 14 C-CO₂ signature (102.91 %Modern) as measured in June, July, August and September 2014 at the Jungfraujoch atmospheric research station. Numbers beneath the dotted line are P-values from t-tests for differences of each date and elevation to this reference line. Significant differences (P < 0.05) are interpreted as a signal of plant-induced heterotrophic respiration of peat carbon through the rhizosphere priming effect.

Fig. 3 Mean seasonal values (\pm 1 SE, n = 3) of humification indices of dissolved organic matter based on FT-IR spectra. Pore-water samples were collected monthly from May through October 2014 in the two peatlands at low and high elevations (1035 and 1885 m a.s.l., respectively) in relation to the presence (+V) or absence (-V) of vascular plants. The selected functional groups are: aliphatics (2943 cm⁻¹ region), carboxylic acids and aromatic esters (1720 cm⁻¹ region), aromatics (1630 cm⁻¹ region), lignins (1510 cm⁻¹ region), phenols (1420 cm⁻¹ region) and they are expressed as a ratio to polysaccharides (1090 cm⁻¹ region). Significant differences between treatments are indicated with asterisks ('*' = P < 0.1, * = P < 0.05, ** = P < 0.01; *** = P < 0.001), whereas no significant differences were observed between the two sites.





