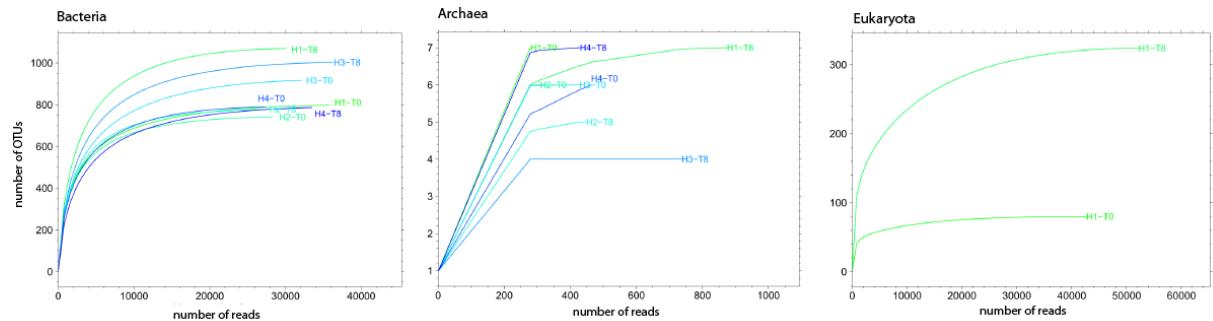
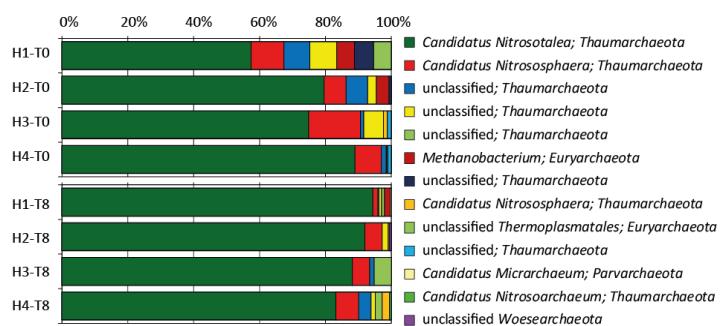


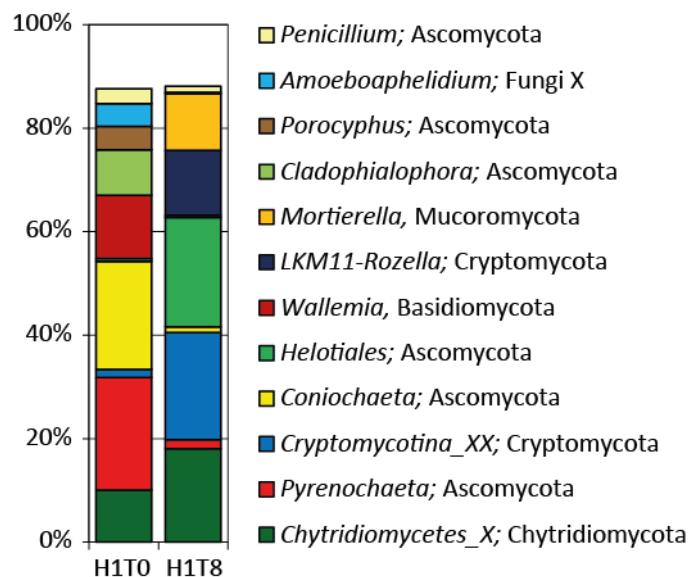
Supplementary material



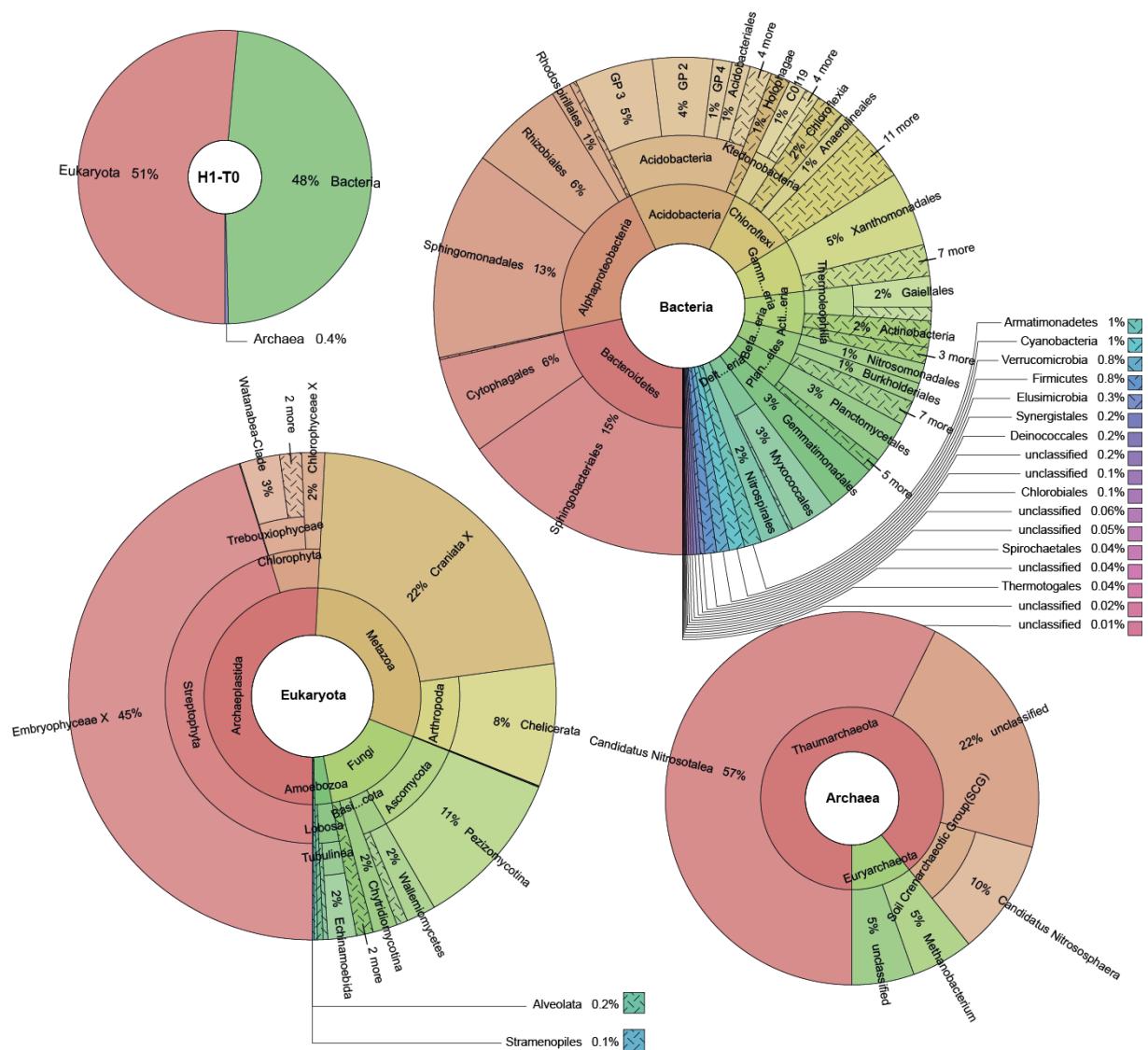
SM1: Rarefaction curves



SM2: Relative abundance of the archaea OTUs in the eight sample.



SM3: Relative abundance of the 12 dominant fungal OTUs in the H1 T0 and H1 T8 samples. Non saturated H1 0-12.5cm; T0 beginning of experiment; T8 after 8 months.



SM4: Community structure content in H1-T0 soil sample generated from Krona.

SM.5: Physiological characteristics of the identified genera among the 100 most abundant OTUs

	Metabolism	respiration	Reference	Evolution in the mesocosm
Leptospirillum	Lithoautotrophic, oxidize FeII Acidophile (optimum pH 1.5) Species can fix nitrogen	aerobic	Garcia-Moyano, 2008 ; Li et al., 2010 ;	Not in the surface layer, maintained, slight decrease in the saturated zone
Acidiferrobacter	Acidophile (optimum pH 2). Lithoautotrophic, oxidize FeII, sulfur Strictly autotrophic and facultative diazotroph.	Facultative anaerobe that can use either molecular oxygen or ferric iron as terminal electron acceptor isolated from coal spoil refuse in Missouri, US.	Hallberg et al., 2011	Not in the surface layer, decrease in the three bottom layers
Sphingomonas	Chemo-organotrophic, metabolically versatile, which means it can utilize a wide range of naturally occurring compounds as well as some types of environmental contaminants	strictly aerobic	Fredrickson et al., 1995 ; Zylstra and Kim, 1997; Macur et al., 2001 ; Kinegam et al., 2008 ; Escalante et al., 2009	Still the most abundant in all layers, increase in second non saturated level H2
Pseudolabrys	Chemo-organotrophic, degradation of organic acids, acid-tolerant (pH 4)	aerobic	Kämpfer et al., 2006	Increase in the saturated (permanently and alternatively) levels
Alkanibacter	Chemo-organotrophic, grows on pentane, hexane and decane	aerobic	Friedrich and Lipski, 2008	Strong decrease in all levels
Bryobacter	Chemo-organotrophic, acid-tolerant sugars, some heteropolysaccharides and galacturonic and glucuronic acids, Acidobacteria	strictly aerobic	Kulichevskaya et al., 2010	Increase or maintain of abundance in all levels
Bradyrhizobium	Chemo-organotrophic, acid tolerant	aerobic	Jordan 1982	Maintained or slight decrease
Gaiella	Nitrate	Strictly aerobic	Albuquerque et al., 2011	Decrease in all

	is reduced to nitrite. Chemo-organotrophic			levels
Geothrix	Respire FeIII with acetate or other organic acids as electron donors, chemo-organotrophic	Strict anaerobe	Coates et al., 1999	Increase in the saturated levels
Lacibacter	Chemo-organotrophic	aerobic	Qu et al., 2009	Increase in the surface (slight) and in the saturated levels (strong)
Sediminibacterium	Chemo-organotrophic	strictly aerobic	Qu and Yuan, 2008	Increase in the saturated levels
Opitutus	Nitrate is reduced to nitrite, fermentation of mono and polysaccharides	obligate anaerobe	Chin et al., 2001	Increase in the saturated levels
Polaromonas	Chemoorganotrophic, some strains degrade PAHs	aerobic	Jeon et al., 2004	Increase in the saturated levels
Gemmamimonas	Chemo-organotrophic	aerobic	Zhang et al., 2003	Decrease in the three lower levels
Varriibacter	Chemo-organotrophic	strictly aerobic	Kim et al., 2014	Decrease in all levels
Holophaga	Degradation aromatic compounds into acetate, homoacetogenesis	Strictly anaerobic	Liesack et al., 1994	Increase in the surface (strong) and lower levels
Rhizomicrobium	Ferric iron, nitrate, oxygen and fumarate served as electron acceptors	Facultative anaerobic	Kodama and Watanabe, 2011	Increase upper levels and in the transiently saturated level
Blastocatella	Chemo-organotrophic growth on very few complex substrate, acid-tolerant	aerobic	Foesel et al., 2013	Decrease in the upper levels
Mucilaginibacter	Chemo-organotrophic, degrading pectin, xylan, laminarin and some other polysaccharides, acid-tolerant	facultatively aerobic (fermentation des sucres possible)	Pankratov et al., 2007	Decrease in the upper levels
Parafilimonas	Chemo-organotrophic	aerobic	Kim et al., 2014	Decrease in surface and in saturated levels
Flavisolibacter	Chemo-organotrophic	aerobic	Yoon and Im, 2007	Decrease in all

				levels
Chryseolinea	Chemo-organotrophic	Strict aerobic	Kim et al., 2013	Decrease in all levels
Cytophaga	Chemo-organotrophic, degrading a large range of organic compounds	Facultatively aerobic	Manz et al., 1996	Decrease in all levels
Pirellula	Chemo-organotrophic, degradation of complex macromolecules	aerobic	Schlesner 1994	Increase in the always saturated level, decrease in the surface level
Undibacterium	Chemo-organotrophic	aerobic	Kampfer et al., 2007	Decrease in all levels
Nitrospira	Obligate chemolithotrophs that oxidize nitrite to nitrate and use carbon dioxide as sole carbon source, no organotrophic growth	aerobic	Ehrich et al., 1995	Decrease in the bottom saturated layer, increase in the 2 top layers
Roseiflexus	Grows photoheterotrophically under anaerobic light conditions and also Chemo-heterotrophically under aerobic dark conditions	Facultative anaerobic	Hanada et al., 2002	Decrease in the surface layer, increase in the 3 bottom layers
Steroidobacter	Reduces nitrate, growth on only a limited number of organic substrates.	Facultative anaerobic	Fahrbach et al., 2008	Increase in the 3 top layers
Devosia	Chemo-organotrophic	aerobic	Kumar et al., 2008	More in the surface level, decrease in the 3 bottom levels
Ohtaekwangia	Chemo-organotrophic	strictly aerobic	Yoon et al., 2011	More in the surface level, narrow decrease
Taibaiella	Chemo-organotrophic	strictly aerobic	Son et al., 2014	Decrease in all levels except the surface
Solibacter	Chemo-organotrophic, acid-tolerant nitrate and nitrite reduction Biofilm production by this species acts as an ecosystem engineer in the soil by enabling this species to adhere to its environment while also reducing	aerobic	GOLD CARD: Gc00446. Candidatus Solibacter usitatus Ellin6076. The Regents of the University of California. 2011	Increase in all levels of the mesocosm

	moisture and nutrient fluxuations in the soil environment under stressful environmental conditions			
Arthrobacter	Chemolithoautotrophic, oxidizes arsenite to arsenate (Prasad et al 2009) or chemoorganotrophic , degrading PAH (Kallimanis et al. 2009)versatile genus	aerobic	Prasad et al 2009, Kallimanis et al., 2009	Decrease in all the mesocosm
Haliangium	usually living in the surface layers of the soil, additionally inhabit decaying plant material, including rotting wood and bark, chemoorganotrophic, needs NaCl	strictly aerobic	Fudou et al 2002	Increase in all levels except the bottom saturated

References

- Albuquerque L., Franca L., Rainey F. A., Schumann P., Nobre M. F., Da Costa M. F., 2011. *Gaiella occulta* gen. nov., sp. nov., a novel representative of a deep branching phylogenetic lineage within the class Actinobacteria and proposal of *Gaiellaceae* fam. nov. and *Gaiellales* ord. nov.. Systematic and Applied Microbiology 34 (8), 595-599.
- Chin K.-J., Liesack W., Janssen P. H., 2001. *Opitutus terrae* gen. nov., sp. nov., to accommodate novel strains of the division ‘Verrucomicrobia’ isolated from rice paddy soil. International Journal of Systematic and Evolutionary Microbiology 51, 1965–1968
- Coates J. D., Ellis D. J., Gaw C. V., Lovley D. R., 1999. *Geothrix fermentans* gen. nov., sp. nov., a novel Fe(III)-reducing bacterium from a hydrocarbon contaminated aquifer. International Journal of Systematic Bacteriology 49, 1615-1622.
- Ehrich S., Behrens D., Lebedeva E., Ludwig W. and Bock E., A new obligately chemolithoautotrophic, nitrite-oxidizing bacterium, *Nitrospira moscoviensis* sp. nov. and its phylogenetic relationship, Archives of Microbiology. 164 (1), 1995, 16–23.

Escalante, G., Campos, V.L., Valenzuela, C., Yañez, J., Zaror, C., Mondaca, M.A., 2009. Arsenic Resistant Bacteria Isolated from Arsenic Contaminated River in the Atacama Desert (Chile). *Bulletin of Environmental Contamination and Toxicology* 83 (5), 657–661.

Fahrbach M., Kuever J., Remesch M., Huber B. E., Kämpfer P., Dott W., Hollender J., 2008. *Steroidobacter denitrificans* gen. nov., sp. nov., a steroidal hormone-degrading gammaproteobacterium. *International Journal of Systematic and Evolutionary Microbiology* 58, 2215–2223.

Foesel B. U., Rohde M., Overmann J., 2013. *Blastocatella fastidiosa* gen. nov., sp. nov., isolated from semiarid savanna soil – The first described species of *Acidobacteria* subdivision 4. *Systematic and Applied Microbiology* 36 (2), 82-89.

Fredrickson, J.K., Balkwill, D.L., Drake, G.R., Romine, M.F., Ringelberg, D.B., White, D.C., 1995. Aromatic-degrading *Sphingomonas* isolates from the deep subsurface. *Applied and Environmental Microbiology* 61 (5), 1917–1922.

Friedrich, M.M., Lipski, A., 2008. *Alkanibacter difficilis* gen. nov., sp. nov. and *Singularimonas variicoloris* gen. nov., sp. nov., hexane-degrading bacteria isolated from a hexane-treated biofilter. *International Journal of Systematic and Evolutionary Microbiology* 58, 2324–2329.

Fudou R., Jojima Y., Iizuka T., Yamanaka S., 2002. *Haliangium ochraceum* gen. nov., sp. nov. and *Haliangium tepidum* sp. nov.: Novel moderately halophilic myxobacteria isolated from coastal saline environments. *Journal of General and Applied Microbiology* 48 (2), 109–115.

Garcia-Moyano A., Gonzales-Toril E., Moreno-Paz M., Parro V., Amils R., 2008. Evaluation of *Leptospirillum* spp. In the Rio Tinto, a model of interest to biohydrometallurgy. *Hydrometallurgy* 94 (1-4), 155-161.

Hallberg, K.B., Hedrich, S., and Johnson, D.B., 2011. *Acidiferrobacter thiooxydans*, gen. nov. sp. nov.; an acidophilic, thermo-tolerant, facultatively anaerobic iron- and sulfur-oxidizer of the family *Ectothiorhodospiraceae*. *Extremophiles* 15 (2), 271–279.

Hanada S., Takaichi S., Matsuura K., Nakamura K., 2002. *Roseiflexus castenholzii* gen. nov., sp. nov., a thermophilic, filamentous, photosynthetic bacterium that lacks chlorosomes. *International Journal of Systematic and Evolutionary Microbiology* 52, 187–193.

Jeon C. O., Park W., Ghiorse W. C., Madsen E. L., 2004. *Polaromonas naphthalenivorans* sp. nov., a naphthalene-degrading bacterium from naphthalene-contaminated sediment. International Journal of Systematic and Evolutionary Microbiology 54, 93–97.

Jordan D. C., 1982. Transfer of *Rhizobium japonicum* Buchanan 1980 to *Bradyrhizobium* gen. nov., a Genus of Slow-Growing, Root Nodule Bacteria from Leguminous Plants. International Journal of Systematic Bacteriology 32, 136–139.

Kallimanis, A., Kavakiotis, K., Perisynakis, A., Sproer, C., Pukall, R., Drainas, C., and Koukkou, A.I., 2009. *Arthrobacter phenanthrenivorans* sp. nov., to accommodate the phenanthrene-degrading bacterium *Arthrobacter* sp. strain Sphe3. International Journal of Systematic and Evolutionary Microbiology 59, 275–279.

Kämpfer, P., Rossello-Mora R., Hermansson M., Persson F., Huber B., Falsen E., Busse H.-J. (2007). *Undibacterium pigrum* gen. nov., sp. nov., isolated from drinking water. International Journal of Systematic and Evolutionary Microbiology, 57, 1510–1515.

Kim K. K., Lee K. H., Eom M. K., Kim J. S., Kim D. S., Ko S. H., Kim B. H., Lee J. S., 2014. *Variibacter gotjawalensis* gen. nov., sp. nov., isolated from soil of a lava forest. Antonie van Leeuwenhoek 105 (5), 915–924.

Kim S.-J., Park J. H., Lim J. M., Ahn J. H., Anandham R., Weon H. Y., Kwon S. W., 2014. *Parafilimonas terrae* gen. nov., sp. nov., isolated from greenhouse soil. International Journal of Systematic and Evolutionary Microbiology 64, 3040–3045.

Kim J. J., Alkawally M., Brady A. L., Rijpstra W. I. C., Damste J. S. S., Dunfield P. F., 2013. *Chryseolinea serpens* gen. nov., sp. nov., a member of the phylum *Bacteroidetes* isolated from soil. International Journal of Systematic and Evolutionary Microbiology 63, 654–660.

Kinegam, S., Yingprasertchai, T., Tanasupawat, S., Leepipatpiboon, N., Akaracharanya, A., Kim, K.-W., 2008. Isolation and characterization of arsenite-oxidizing bacteria from arsenic-contaminated soils in Thailand. World Journal of Microbiology and Biotechnology 24 (12), 3091–3096.

Kodama Y., Watanabe K., 2011. *Rhizomicrobium electricum* sp. nov., a facultatively anaerobic, fermentative, prosthecate bacterium isolated from a cellulose-fed microbial fuel cell. International Journal of Systematic and Evolutionary Microbiology 61, 1781–1785.

Kulichevskaya, I.S., Suzina, N.E., Liesack, W., Dedysh, S.N., 2010. *Bryobacter aggregatus* gen. nov., sp. nov., a peat-inhabiting, aerobic chemo-organotroph from subdivision 3 of the *Acidobacteria*. International Journal of Systematic and Evolutionary Microbiology 60, 301–306.

Kumar M., Verma M., Lal R., 2008. *Devosia chinhatensis* sp. nov., isolated from a hexachlorocyclohexane (HCH) dump site in India. International Journal of Systematic and Evolutionary Microbiology 58, 861–865.

Li, B., Lin, J., Mi, S., Lin, J., 2010. Arsenic resistance operon structure in *Leptospirillum ferriphilum* and proteomic response to arsenic stress. Bioresource Technology 101, 9811-981.

Liesack W., Bak F., Kreft J.-U., Stackebrandt E., 1994. *Holophaga foetida* gen. nov., sp. nov., a new, homoacetogenic bacterium, degrading methoxylated aromatic compounds. Archives of Microbiology 162 (1-2), 85-90.

Macur, R.E., Wheeler, J.T., McDermott, T.R., and Inskeep, W.P., 2001. Microbial Populations Associated with the Reduction and Enhanced Mobilization of Arsenic in Mine Tailings. Environmental Science & Technology 35 (18), 3676–3682.

Manz, W., Amann, R., Ludwig, W., Vancanneyt, M., Schleifer, K. H., 1996. Application of a suite of 16S rRNA-specific oligonucleotide probes designed to investigate bacteria of the phylum *cytophaga-flavobacter-bacteroides* in the natural environment. Microbiology 142 (5), 1097-1106.

Pankratov T. A., Tindall B. J., Liesack W., Dedysh S. N., 2007. *Muciluginibacter paludis* gen. nov., sp. nov. and *Muciluginibacter gracilis* sp. nov., pectin-, xylanand laminarin-degrading members of the family *Sphingobacteriaceae* from acidic Sphagnum peat bog. International Journal of Systematic and Evolutionary Microbiology 57, 2349–2354.

Prasad, K.S., Subramanian, V., and Paul, J. (2009). Purification and characterization of arsenite oxidase from Arthrobacter sp. BioMetals 22, 711–721.

Qu J.-H., Yuan H.-L., 2008. *Sediminibacterium salmonicum* gen. nov., sp. nov., a member of the phylum *Bacteroidetes* isolated from sediment of a eutrophic reservoir. International Journal of Systematic and Evolutionary Microbiology 58, 2191–2194.

Qu J.-H., Yuan H.-L., Yang J. S., Li H. F., Chen N., 2009. *Lacibacter cauensis* gen. nov., sp. nov., a novel member of the phylum *Bacteroidetes* isolated from sediment of a eutrophic lake. International Journal of Systematic and Evolutionary Microbiology 59, 1153–1157.

Schlesner H., 1994. The Development of Media Suitable for the Microorganisms Morphologically Resembling *Planctomyces* spp., *Pirellula* spp., and other Planctomycetales from Various Aquatic Habitats Using Dilute Media. Systematic and Applied Microbiology 17, 135-145.

Son H. M., Kook M. C., Kim H. J., Yi T. H., 2014. *Taibaiella koreensis* sp. nov., isolated from soil of a ginseng field. International Journal of Systematic and Evolutionary Microbiology 64, 1018–1023.

Yoon M. H., Im W.-T., 2007. *Flavisolibacter ginsengiterrae* gen. nov., sp. nov. and *Flavisolibacter ginsengisoli* sp. nov., isolated from ginseng cultivating soil. International Journal of Systematic and Evolutionary Microbiology, 57, 1834–1839.

Yoon J.-H., Kang S. J., Lee S. Y., Lee J. S., Park S., 2011. *Ohtaekwangia koreensis* gen. nov., sp. nov. and *Ohtaekwangia kribbensis* sp. nov., isolated from marine sand, deep-branching members of the phylum Bacteroidetes. International Journal of Systematic and Evolutionary Microbiology 61, 1066–1072.

Zhang H., Sekiguchi Y., Hanada S., Hugenholtz P., Kim H., Kamagata Y., Nakamura K., 2003. *Gemmatimonas aurantiaca* gen. nov., sp. nov., a Gram-negative, aerobic, polyphosphateaccumulating micro-organism, the first cultured representative of the new bacterial phylum *Gemmatimonadetes* phyl. nov.. International Journal of Systematic and Evolutionary Microbiology 53, 1155–1163.

Zylstra, G.J., Kim, E., 1997. Aromatic hydrocarbon degradation by *Sphingomonas yanoikuyae* B1. J Industrial Microbiology and Biotechnology 19 (5-6), 408–414.