

Global synthesis for the scaling of soil microbial nitrogen to phosphorus in terrestrial ecosystems

Zhiqiang Wang, Mingcheng Wang, Kailiang Yu, Huifeng Hu, Yuanhe Yang, Philippe Ciais, Ashley P. Ballantyne, Karl J. Niklas, Heng Huang, Buqing

Yao, et al.

► To cite this version:

Zhiqiang Wang, Mingcheng Wang, Kailiang Yu, Huifeng Hu, Yuanhe Yang, et al.. Global synthesis for the scaling of soil microbial nitrogen to phosphorus in terrestrial ecosystems. Environmental Research Letters, 2021, 16, 10.1088/1748-9326/abed78. insu-03660171

HAL Id: insu-03660171 https://insu.hal.science/insu-03660171

Submitted on 5 May 2022

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License

ENVIRONMENTAL RESEARCH _ETTERS

LETTER • OPEN ACCESS

Global synthesis for the scaling of soil microbial nitrogen to phosphorus in terrestrial ecosystems

To cite this article: Zhiqiang Wang et al 2021 Environ. Res. Lett. 16 044034

View the article online for updates and enhancements.

You may also like

- Typhoon enhancement of N and P release from litter and changes in the litter N:P ratio in a subtropical tidal wetland Weiqi Wang, Jordi Sardans, Chuan Tong et al.
- The U.S. consumer phosphorus footprint: where do nitrogen and phosphorus diverge? Geneviève S Metson, Graham K MacDonald, Allison M Leach et al.
- Nitrogen -addition accelerates phosphorus cycling and changes phosphorus use strategy in a subtropical Moso bamboo forest

Quan Li, Jianhua Lv, Changhui Peng et al.

ENVIRONMENTAL RESEARCH

CrossMark

OPEN ACCESS

RECEIVED 30 September 2020

REVISED 27 December 2020

ACCEPTED FOR PUBLICATION

10 March 2021

PUBLISHED 25 March 2021

Original content from this work may be used under the terms of the Creative Commons Attribution 4.0 licence.

Any further distribution of this work must maintain attribution to the author(s) and the title of the work, journal citation and DOI.



Global synthesis for the scaling of soil microbial nitrogen to phosphorus in terrestrial ecosystems

Zhiqiang Wang^{1,*}^(D), Mingcheng Wang^{1,*}, Kailiang Yu², Huifeng Hu³, Yuanhe Yang³, Philippe Ciais², Ashley P Ballantyne^{2,4}, Karl J Niklas⁵, Heng Huang^{6,**}, Buqing Yao^{7,**}

and S Joseph Wright⁸

LETTER

- ¹ Institute for Advanced Study, Chengdu University, Chengdu, Sichuan, People's Republic of China
- ² Le Laboratoire des Sciences du Climat et de l'Environnement, IPSL-LSCECEA/CNRS/UVSQ Saclay, Gif-sur-Yvette, France
- ³ State Key Laboratory of Vegetation and Environmental Change, Institute of Botany, Chinese Academy of Sciences, Beijing, People's Republic of China
- Department of Ecosystem and Conservation Sciences, WA Franke College of Forestry and Conservation, University of Montana, Missoula, MT, United States of America
- ⁵ Plant Biology Section, School of Integrative Plant Science, Cornell University, Ithaca, NY, United States of America
- ⁶ Department of Environmental Science, Policy, and Management, University of California, Berkeley, United States of America
- ⁷ Key Laboratory of Cold Regions Restoration Ecology, Qinghai Province, and Key Laboratory of Adaptation and Evolution of Plateau Biota, Northwest Institute of Plateau Biology, Chinese Academy of Sciences, Xining, People's Republic of China
- Smithsonian Tropical Research Institute Apartado, 0843-03092 Balboa, Ancon, Panama
- These authors contributed equally to this paper.
 - Authors to whom any correspondence should be addressed.

E-mail: henghuang@berkeley.edu, bqyao@nwipb.cas.cn

Keywords: nitrogen, nitrogen:phosphorus ratio, phosphorus, scaling exponent, soil microbial biomass Supplementary material for this article is available online

Abstract

Nitrogen (N) and phosphorus (P) are associated with the life history of all organisms. Soil microbes play essential roles in nutrient cycling and ecosystem dynamics. As compared to plants, however, our understanding of the soil microbial N versus P scaling relationship remains limited. In this study, we used a comprehensive dataset comprising 2210 observations from 422 sites worldwide to examine microbial C, N, and P concentrations and C:N:P ratios, and to determine the scaling of microbial N versus P in different ecosystems and spatial scales (i.e. soil depths, latitudinal zones, and local sites). The global mean values of microbial C, N, and P concentrations were 721.5 mg kg⁻¹, 84.7 mg kg⁻¹, 37.6 mg kg⁻¹, respectively, whereas C:N, C:P, and N:P ratios were 9.5, 30, and 3.4, respectively. Microbial C, N, and P concentrations varied within and across different ecosystems and spatial scales, but C:N:P ratios varied surprisingly little. The numerical value of the scaling exponent (i.e. the slope of the log-log linear relationship) of microbial N versus P was 0.89 across the entire dataset and for different ecosystems. However, the numerical value of the exponent varied significantly across different spatial scales. Soil total P was the largest contributor to the variation observed in the scaling of microbial N versus P at different local sites. These findings have important implications for predicting soil microbial growth rates and improving our understanding of nutrient cycling in plant-soil-microbe systems.

1. Introduction

Nitrogen (N) and phosphorus (P) are important growth-limiting elements in terrestrial ecosystems and regulate ecosystem functions and biogeochemical cycles (Chapin 1980, Güsewell 2004, Reich and Oleksyn 2004). It is critical for organisms to maintain a balance of N and P for their basic metabolic requirements and to sustain the availability of N and P in the environment (Sterner and Elser 2002). Therefore, a quantitative evaluation-1pc of the N versus P scaling relationship across different organisms can improve our mechanistic understanding of nutrient cycling and ecosystem dynamics (Elser *et al* 2000, Hedin 2004, Kerkhoff *et al* 2006, Cleveland and Liptzin 2007, Reich *et al* 2010).

It is widely known that N and P concentrations in organisms are tightly correlated and can be quantified via a general log-log stoichiometric scaling relationship, taking the form $N = \beta P^{\alpha}$ (or, when logtransformed, $\log N = \log \beta + \alpha \log P$), where β and α are the scaling constant and scaling exponent (i.e. the Y-intercept and slope of the log-log linear relationship), respectively (Sterner and Elser 2002, Niklas et al 2005, Kerkhoff et al 2006). This simple N versus P scaling relationship provides a useful empirical model to predict the N and P limitation of organisms and ecosystem functioning (Elser et al 2000, Díaz et al 2004). For example, the empirical stoichiometric growth model (also known as the growth rate hypothesis, GRH) states that rapidly growing organisms require disproportionately more P than N to ensure rapid protein synthesis (Elser et al 2003), thereby resulting in a N versus P scaling exponent below unity $(\alpha < 1)$ (Sterner and Elser 2002).

During the past several years, numerous studies have explored N versus P relationship in plant tissues, such as leaves (Wright et al 2004, Han et al 2005, Niklas et al 2005, Reich et al 2010), stems (Niklas and Cobb 2005, Wang et al 2018, Zhang et al 2018), roots (Yuan et al 2011, Wang et al 2019), and seeds (Wang et al 2020). These studies have revealed that the strategic allocation of N and P in plant tissues appears to follow fundamental stoichiometric rules (Niklas et al 2005, Niklas 2006, Ågren 2008) that regulate the stoichiometry of elements in plant tissues. Previous work has also shown that the scaling exponent for the N versus P relationship is apparently constant, as a result of environmental convergence and evolutionary history (McGroddy et al 2004, Wright et al 2004). For example, based on a large and comprehensive dataset, the exponent of the N versus P scaling relationship is reported to be 0.66 for leaves (Reich et al 2010) and 0.82 for fine roots (Wang et al 2019). Nevertheless, several studies have reported significant interspecific variation in the N versus P scaling exponent in leaves (Niklas and Cobb 2005, McGroddy et al 2004, Tian et al 2017), roots (Yuan et al 2011, Geng et al 2014, Zhao et al 2016), and among geographical locations due to environmental differences.

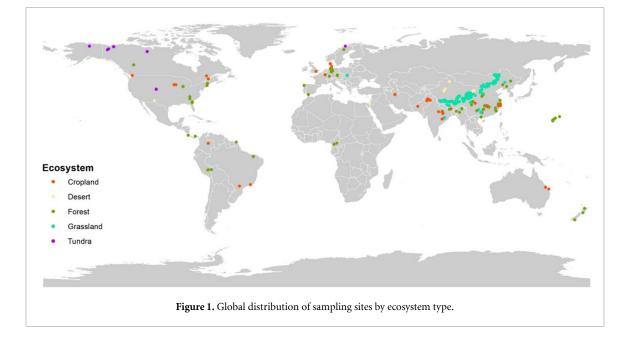
Despite extensive studies of plant organs, our quantitative understanding of the covariation of these two nutrients in soil microbes remains extremely limited in comparison (Cleveland and Liptzin 2007, Hartman and Richardson 2013). As an important indicator of soil fertility, soil microbes play fundamental roles in driving and regulating fundamental terrestrial ecosystem processes such as soil carbon (C) sequestration, organic matter decomposition, and nutrient recycling (Li *et al* 2012, Mooshammer *et al* 2014a, Heuck *et al* 2015, Chen *et al* 2016, Spohn 2016), and thus affect ecosystem productivity (Van Der Heijden et al 2008). For example, soil microbial biomass accounts for approximately 1%–5% of the soil organic C (Jenkinson et al 1990), and soil microbial dynamics largely determine the release of immobilized labile nutrients (Singh and Gupta 2018). Soil microbes also contribute to soil N mineralization and soil respiration by recycling C and N as carbon dioxide and ammonium (Moorhead et al 2012). Equally important, soil microbes can transform soil N and P into a labile pool of plantavailable nutrients. In turn, important resources are supplied to soil microbes through the input of plant necromass, root exudation, and other soil organic matter (Mooshammer et al 2014b). Given that the stoichiometry of N and P is tightly coupled within plant-soil-microbe systems (McGroddy et al 2004), a scaling relationship of N versus P might also exist in soil microbial systems in a manner similar to that of plants.

In this study, we compiled a comprehensive global dataset of paired soil microbial biomass C, N, and P to explore whether a general N versus P scaling relationship exists. Specifically, we (a) estimated microbial C, N, and P concentrations and C:N:P ratios across different ecosystems, soil depths, and latitudinal zones; (b) quantified the microbial N versus P scaling exponent across ecosystems, soil depths, latitudinal zones, and local sites; and (c) explored the relationships among environmental factors and the scaling exponent across sites.

2. Materials and methods

2.1. Data sources

We compiled a dataset of soil microbial biomass carbon (MBC), microbial biomass nitrogen (MBN), and microbial biomass phosphorus (MBP) from the primary published literature using the phrase 'soil microbial biomass' in Google Scholar, Web of Science (http://apps.webofknowledge.com), and the China National Knowledge Infrastructure Database (http://cnki.net) from 1980 to 2019. Four criteria were used to avoid bias in selecting publications. First, microbial C, N, and P had to be reported simultaneously for the same samples. Based on this criterion, we excluded some publications used in previous studies (Cleveland and Liptzin 2007, Xu et al 2013, Li et al 2014). Second, microbial C, N, and P data had to be for mineral soil depths between 0 and 30 cm. Third, only data from control plots that reported the results of manipulation experiments were used. Finally, fourth, to avoid variation among methods, soil microbial biomass had to be evaluated using the chloroform fumigation-extraction (CFE) method (Vance et al 1987). CFE is the most commonly used method and provides an index of total soil microbial biomass, including both bacteria and fungi (Fierer et al 2009).



We recorded additional site-specific data from the original publications, including geographic location (i.e. latitude and longitude), climatic variables (i.e. mean annual temperature MAT, and mean annual precipitation, MAP), and soil physical and chemical properties (i.e. soil pH, soil organic C, and total N and P). When data were presented graphically, they were extracted from figures, tables, and appendices of the original publications using GetData Graph Digitizer (version 2.22). When the geographical coordinates and climatic factors were not specified in the original articles, they were obtained from the Global Gazetteer Version 2.2 (www.fallingrain.com/world/index.html) and the WorldClim global climatic database (http://worldclim.org/) with a grid precision of $0.5^{\circ} \times 0.5^{\circ}$ according to geographical coordinates, respectively. To standardize the data, all microbial biomass N and P values were converted into units of mg kg⁻¹ dry soil. The final dataset consisted of 2210 observations from 422 sites worldwide, reported in 131 published studies (figure 1; a list of the data sources is provided in the supplementary materials). The dataset encompassed five types of terrestrial ecosystems, including croplands (1030 observations), forests (657 observations), grasslands (366 observations), deserts (81 observations), and tundra (76 observations).

2.2. Data analysis

One-way ANOVA was used to evaluate possible differences in microbial C, N, P concentrations, and C:N, C:P, and N:P ratios among ecosystems, latitudinal zones, and soil depths, followed by Tukey's *post-hoc* comparisons when effects were significant (P < 0.05). All of the data were log-transformed to achieve a normal distribution for robust statistical analysis (figure S1 (available online at stacks.iop.org/ERL/16/044034/mmedia)). Standard major axis regression (SMA, a form of model II regression) was used to determine the microbial N versus P scaling relationship for ecosystems, soil depths, latitudinal zones, and the sites using logtransformed values of microbial N and P. Each site was sorted into one of five ecosystems (croplands, forests, grasslands, deserts, or tundra). Given the influence of climate and soil properties on soil microbial biomass (Li et al 2012, Xu et al 2013, Tischer et al 2014, Liu et al 2020), sites were also sorted into one of three latitudinal zones (tropical 0-25°, temperate $25-50^\circ$, and boreal $>50^\circ$) and into one of three soil depths (0-10 cm, 10-20 cm, and 20-30 cm). Data for other depth thresholds (e.g. 0-15 cm, 0-20 cm or 15-30 cm) were excluded from soil depth analyses. Finally, we examined the microbial N versus P scaling relationships for each of the 26 sites with 10 or more records. These 26 sites included croplands (6 sites), forests (8 sites), grasslands (2 sites), deserts (2 sites), tundra (3 sites), and croplands + forests (5 sites). Heterogeneity of the numerical values of scaling exponents within the different categories of ecosystems, soil depths, and latitudinal zones was tested using likelihood ratio tests (Warton et al 2006). 'Funnel' graph analyses were used to evaluate whether scaling exponents varied with sample size for the 26 sites with 10 or more samples (Wright et al 2005, Reich et al 2010).

We further analyzed the influence of environmental factors on variation in the numerical values of the microbial N versus P scaling exponents for the 26 sites with 10 or more samples. Stepwise multiple regression (SMR) was used to identify the contributions of six environmental factors (i.e. MAT, MAP, soil organic C, total N and P, and pH). Environmental factors that did not significantly contribute to the variation in scaling exponent were excluded from subsequent analyses. Redundancy analysis (RDA) was used to determine the proportion of variation in scaling exponents explained by each significant environmental factor. All statistical analyses were performed using R 3.4.0 (R Core Team 2017).

3. Results

3.1. C, N, and P concentrations and C:N:P stoichiometry in soil microbial biomass

The overall mean values of microbial C, N, and P concentrations were 721.5 mg kg⁻¹, 84.7 mg kg⁻¹, and 37.6 mg kg $^{-1}$, respectively. The overall mean values of C:N, C:P, and N:P ratios were 9.5, 30, and 3.4, respectively (table 1). Microbial C, N, and P concentrations varied among ecosystems, soil depths, and latitudinal zones. Across ecosystems, tundra had the highest concentrations of microbial C, N, and P. In contrast, the microbial C was lowest in deserts, whereas the microbial N and P were lowest in grasslands. Across soil depths, microbial C, N, and P concentrations were largest in the 0-10 cm soil layer, and microbial C and P were lower in the 20-30 cm layer, whereas microbial N was lowest in the 10-20 cm soil layer. Across latitudinal zones, the boreal and temperate zones had the highest and lowest concentrations of microbial C, N, and P, respectively. The global averaged microbial C:N:P ratio was 32:3:1. Compared to microbial C, N, and P concentrations, microbial C:N:P ratios were confined to a relatively modest range in different ecosystems, soil depths, and latitudinal zones (table 1). For example, microbial C:N ratios ranged from 5.0 for deserts to 10.9 for croplands. Microbial C:P ratios ranged from 24.8 in forests to 42.6 in grasslands, whereas microbial N:P ratios only ranged from 2.1 for deserts to 4.3 for grasslands.

3.2. Microbial N versus P scaling exponents across ecosystems, soil depths, and latitudinal zones

Across the entire dataset, the numerical value of the microbial N versus P scaling exponent was 0.89 (P < 0.001, $r^2 = 0.62$; figure 2(a), table 2). The numerical values of the scaling exponent showed no significant differences among ecosystems, with all ecosystems having values close to 0.89 (figures 2(b)–(f), table 2). In contrast, the numerical values of the scaling exponent differed significantly among soil depths, ranging from 0.89 in the 0–10 cm soil layer to 0.63 in the 20–30 cm soil layer (figure 3(a), table 2). The numerical values of the scaling exponent also differed significantly among latitudinal zones, with larger values in the tropical (0.96) and boreal (0.98) zones than in the temperate (0.85) zone (figure 3(b), table 2).

3.3. Microbial N versus P scaling exponents across local sites

The numerical values of the scaling exponent of the microbial N versus P scaling relationship varied significantly among sites with 10 or more values (figure 4(a)). A funnel-plot assessment of the scaling exponent in relation to the sample size for the 26 sites showed that the numerical values of the scaling exponent ranged from 0.45 to 1.41 with a mean value of 0.92. The numerical values converged around the mean value with increasing sample size (approx. equal to 0.89) (figure 4(b)).

Preliminary SMR analysis demonstrated that MAT, soil organic C, and total N did not significantly affect the numerical values of the microbial N versus P scaling exponent. The SMR analysis showed that MAP, soil pH, and soil total P explained 74% of the variation in the numerical values of the scaling exponents across sites ($r^2 = 0.74$, P < 0.001). The values of scaling exponents were positively correlated with MAP (a standardized coefficient = 0.30, P < 0.05), and negatively correlated with soil pH (a standardized coefficient of = -0.30, P < 0.05) and soil total P (a standardized coefficient of = -0.69, P < 0.001) (figure S2). RDA analyses demonstrated that MAP, soil pH, total P, and their interactions collectively explained 80.2% of the observed variation in the numerical values of the microbial N versus P scaling exponents (figure 5). When these factors were decomposed, soil total P was the largest contributor (49.1%) to the numerical values of the scaling exponent.

4. Discussion

4.1. Variation in microbial C, N, and P concentrations and C:N:P ratios across ecosystems, soil depths, and latitudinal zones

The results reported here indicate that global mean microbial C, N, and P concentrations are 721.5 mg kg⁻¹, 84.7 mg kg⁻¹, and 37.6 mg kg⁻¹, respectively, which are lower than those global means reported by Xu *et al* (2013) (748.4 mg kg⁻¹, 131.4 mg kg⁻¹, and 67.5 mg kg⁻¹, respectively). The discrepancy between our study and that of Xu et al (2013) can be attributed to using different datasets to calculate mean values. For example, the global average of microbial C, N, and P concentrations reported here are based on paired microbial C, N, and P values, whereas comparatively few paired microbial C, N, and P values were used in the study of Xu et al (2013). Moreover, the dataset used by Xu et al (2013) included a significant proportion of data drawn from manipulated plots (e.g. fertilizer and temperature treatments), organic layers or unspecified soil depths. However, our data were obtained from control plots and from specific surface soil horizons (depths of 0–30 cm). These differences probably also contribute to the lower microbial C, N, and P concentrations reported in our study. In addition, it is important to point out that the majority of microbial C, N, and P in our dataset were obtained from croplands and forest ecosystems. This inequality

		Table 1. MBC, MBN and MBP	concentrations and their ratios fi	Table 1. MBC, MBN and MBP concentrations and their ratios for different ecosystems, soil depths, and latitudinal zones.	s, and latitudinal zones.		
Group	и	$MBC (mg kg^{-1})$	$MBN (mg kg^{-1})$	$MBP (mg kg^{-1})$	MBC:MBN	MBC:MBP	MBN:MBP
All	2210	721.5 (678.9–764)	84.7 (79.9–89.5)	37.6 (34.9–40.4)	9.5 (9.3–9.8)	30 (27.9–32.1)	3.4 (3.2-3.6)
Ecosystem							
Cropland	1030	897.8 ^b (846.8-048.8)	75.8 ^b (77.8-78.8)	30.9 ^c (76 3 37 4)	10.9^{a}	30.3 ^b (70.3-31.4)	3.1 ^b
Forest	657	491.9° (455.8–528.1)	78.9 ^b 72.4–85.5)	(29.4–36.7) (29.4–36.7)	(10.3–111.2) 7.5 ^b (7.1–8)	24.8 ^b (22.1–27.4)	()) 3.7 ^a (3.4–4)
Grassland	366	301.8° (274.6–329.0)	39.0 ^c (35.3–42.6)	16.1 ^d (14–18.2)	9.8^a (9.0–10.6)	42.6^{a} (31.5–53.7)	4.3^{a} (3.5–5)
Desert	81	193.3^{d} (142.4–244.2)	69.5 ^b (56.6–82.4)	58.8 ^b (48.8–68.7)	5 ^b (2.1–8)	25.6^{b} (13.6–37.6)	$2.1^{\rm b}$ (1.4–2.7)
Tundra	76	4158.9^{a} (3067.2–5250.7)	492.1^{a} (481.7–500)	248.6 ^a (199.2–298)	10.6^{a} (9.3–11.8)	38.8 ^a (25.4–52.2)	3.1 ^b (2.4–3.8)
Soil depth							
0-10 cm	697	762.8 ^a (653 7–872 0)	114.3 ^a (101 3–127 4)	54.2 ^a (46 4–61 9)	9.4 ^a (8 7–10 1)	32.3 ^a (78 0–36 6)	4.0 ^a (3.6–4.4)
10–20 cm	124	204.8 ^b	41.2 ^b	20.5 ^c	(5.5 ^b	32.5 ^a	4.0^{a}
20–30 cm	48	(177.2-232.4) 114.8 ^b (80.8-148.8)	(34.7-47.7) 35^{b} (24.2-45.7)	(15.8-25.3) 30.9^{b} (21.3-40.4)	$(5.6^{\rm b}$ (3.4-7.9)	(14.7-50.3) 52.5 ^a (5.7-110)	(2.8-5.1) 4.1 ^a (0.9-7.2)
Latitudinal zone							
Tropical (0–25°)	660	1030.6 ^b (954.0–1107.2)	89.6 ^b (84. 5–94.7)	33.2 ^b (30.4–35.9)	10.8^{a} (10.3–11.3)	32.6^{a} (30.7–34.5)	3.4^{a} (3.2-3.7)
Temperate $(25-50^{\circ})$	1418	466.9° (446.2–487.5)	60.2° (57.0–63.3)	27.1° (25.5–28.6)	$9.0^{ m b}$ (8.7–9.4)	29.3^{a} (26.2–32.4)	3.5^{a} (3.2-3.7)
Boreal (>50°)	132	2273.6^{a} (1678.2–2869.1)	323.6 ^a (270.8–376.5)	173.4^{a} (140.1–206.8)	$8.8^{\rm b}$ (8.1–9.6)	$23.4^{\rm a}$ (17.0–29.8)	2.5 ^b (2.1–2.9)
Values are means with 95% cor	ufidence intervals in pa	arentheses. <i>n</i> is the number of sau	mples. Different letters mean sign	Values are means with 95% confidence intervals in parentheses. n is the number of samples. Different letters mean significant differences at the 0.05 level	_		

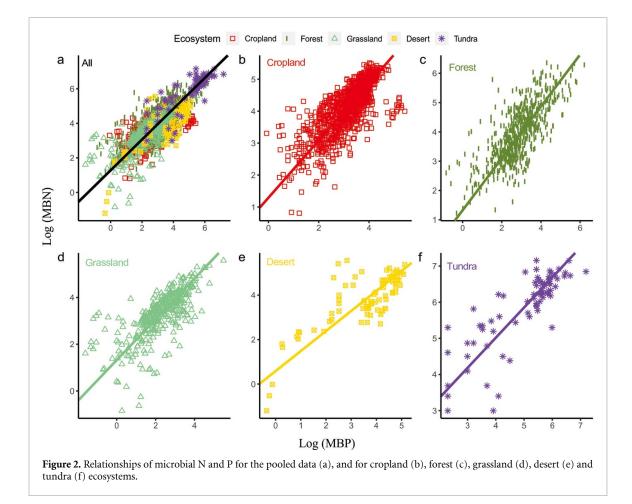


Table 2. Summary of standard major axis (SMA) regression analyses for the relationship between microbial N versus P for different ecosystems, soil depths and latitudinal zones.

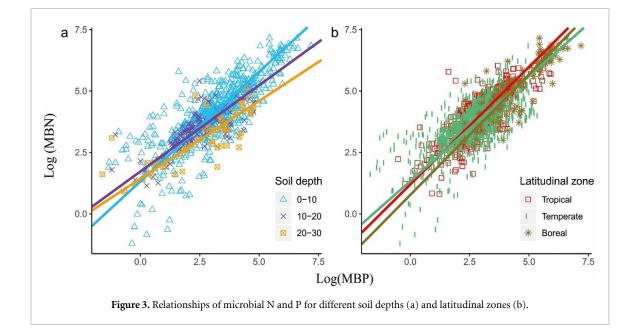
Group	п	Intercept	Exponent	Low CI	High CI	r^2
All	2210	1.28	0.89	0.87	0.91	0.62
Ecosystem						
Cropland	1030	1.29	0.90 ^a	0.86	0.93	0.55
Forest	657	1.32	0.89 ^a	0.84	0.93	0.54
Grassland	366	1.32	0.87^{a}	0.81	0.94	0.51
Desert	81	0.57	0.90 ^a	0.79	1.04	0.62
Tundra	76	1.68	0.83 ^a	0.73	0.95	0.66
Soil depth						
0–10 cm	697	1.29	0.89 ^a	0.85	0.93	0.66
10–20 cm	124	1.73	0.70^{b}	0.62	0.79	0.54
20–30 cm	48	1.43	0.63 ^c	0.52	0.76	0.59
Latitudinal zone						
Tropical (0–25°)	660	1.20	0.96 ^a	0.92	1.00	0.68
Temperate $(25-50^{\circ})$	1418	1.33	0.85^{b}	0.82	0.88	0.52
Boreal (>50°)	132	0.77	0.98 ^a	0.90	1.07	0.74

Different letters denote significant differences in exponents based on a likelihood ratio test. Each regression relationship is statistically significant at P < 0.001 (all data were log₁₀-transformed before analysis).

in the data drawn from different ecosystems could lead to the differences in the global values reported here and elsewhere. Clearly, there is a need for more data on microbial C, N, and P from other

ecosystems before any canonical conclusions can be drawn.

Indeed, large differences in microbial C, N, and P concentrations across different ecosystems, soil



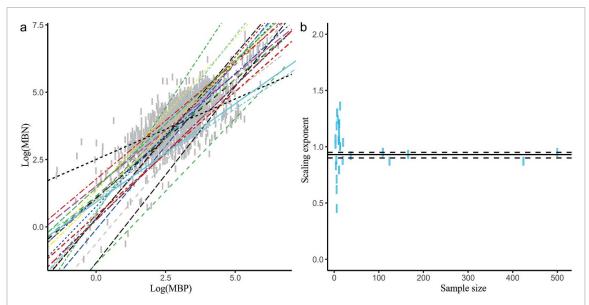
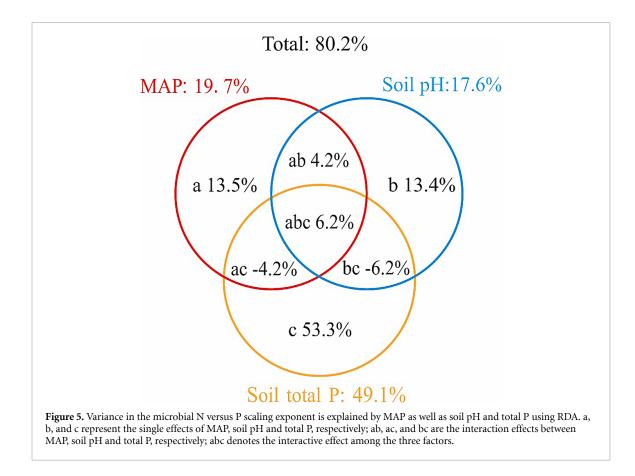


Figure 4. Relationships of microbial N and P at 26 sites with more than ten records (a); relationship between the scaling exponent of microbial N and P and the sample size for 26 sites (b). The mean (0.92) and 95% CI for the scaling exponent are shown in the solid and dashed lines, respectively.

depths, and latitudinal zones are evident in our global dataset (table 1). The microbial C, N, and P concentrations in the tundra sites in our dataset are significantly higher than in other ecosystems, which is consistent with the results reported previously by Xu et al (2013). We found that the microbial C, N, and P concentrations are significantly higher in the 0-10 cm soil layer than in deeper soil layers. This finding probably reflects surface C, N, and P inputs from plant residues, including litterfall and necromass (Sistla and Schimel 2012). We also observed significantly higher microbial C, N, and P concentrations in the boreal zone compared to the other latitudinal zones. This finding is consistent with the hypothesis that soil microbes at high latitudes with low temperatures tend to have higher C, N, and P concentrations, most probably in

order to compensate for their lower metabolic activity (Li *et al* 2014).

The global average microbial C:N:P ratio (i.e. 32:3:1) in our study is considerably lower than that reported by Xu *et al* (2013) (42:6:1). This difference is likely attributable to the factors previously discussed, e.g. a large proportion of microbial C, N, and P data (30%) used by Xu *et al* (2013) was drawn from manipulated plots, which can result in differences in microbial C, N, and P concentrations and thus their ratios. In addition, all of the microbial C, N, and P data used in our study are based on the FE determination method, whereas the microbial C, N, and P data used in previous datasets are based on only 55% of studies using this protocol. Thus, a 'method discrepancy' may also lead to higher C:N:P ratios in previous



studies (e.g. Jenkinson *et al* 2004) compared to ours. Microbial C, N, and P concentrations tend to covary across ecosystems, soil depths, and latitudinal zones so that C:N:P ratios vary less than do C, N, and P concentrations (table 1). This result supports the existence of 'Redfield-like' ratios in soil microbes (Redfield 1958, Cleveland and Liptzin 2007). In addition, our results further support the contention that the growth of soil microbes is limited by C, rather than by N or P, indicating the soil microbial biomass N:P ratios should not be used as an indicator of ecosystem limitation (Allison *et al* 2010, Xu *et al* 2013).

4.2. The numerical values of microbial N versus P scaling exponents across different ecosystems, soil depths and latitudinal zones

The numerical value of the scaling exponent governing the microbial N versus P scaling relationship is approximately 0.89 across the different ecosystems (figure 2, table 2). This value is slightly lower than that reported by Hartman and Richardson (2013) at the global scale, i.e. 1.04. This difference is likely a result of the different communities sampled. For example, a large proportion of data (34%) used in the Hartman and Richardson (2013) study comes from landuse regimes (e.g. pastures and grazed lands). The 0.89 scaling exponent for the microbial N versus P scaling relationship reported here indicates that microbial P concentrations increase faster than microbial N concentrations (i.e. the microbial N versus P scaling exponent is below unity), which supports the growth rate hypothesis of Sterner and Elser (2002).

The numerical value of the microbial N versus P scaling exponent declines from the 0-10 cm to the 20–30 cm soil layer (figure 3(a)), which may reflect an adaptive response to changes in the availability of soil resources. In deeper soil layers, studies have shown that microbial metabolic processes are limited by C availability as a reflection of denitrification and P mineralization processes (Fierer et al 2003, Camenzind et al 2018). Like other organisms, soil microbes must optimize their resource allocation in response to nutrient limitation, which can lead to lower scaling exponent values. Moreover, soil microbes can regulate their nutrient-use efficiencies within different soil depths to adjust N and P investments in growth (Manzoni et al 2012, Mooshammer et al 2014b). Thus, soil microbes are flexible and very efficient in acquiring nutrients from different soil depths to maintain their metabolic functions (Bardgett et al 2003).

We found that the numerical values of the microbial N versus P scaling exponents in boreal and tropical latitudes are similar and close to 1.0. Due to the N and P limitations in boreal and tropical ecosystems, microbial communities can also mobilize N and P elements to meet their demands by producing specific extracellular enzymes to alleviate limitations (Zechmeister-Boltenstern *et al* 2015). Moreover, in boreal areas, low temperatures may strongly restrain microbial growth by reducing metabolic activity. Our SMR analysis shows that the numerical value of the microbial N versus P scaling exponent is negatively correlated with soil pH, such that the scaling exponent in acid soils is numerically greater than in alkaline soils, perhaps greater N and P use efficiency in more acidic soils with less diversity (Fierer and Jackson 2006). This result supports the notion that in tropical areas, soil microbes in acid soils redirect their metabolic energy from growth to cell maintenance (Anderson and Domsch 1993). Therefore, it is not particularly surprising that the numerical value of the microbial N versus P scaling exponent is close to 1.0 in boreal and tropical areas compared to temperate areas with more equitable climates.

4.3. Variant numerical values of microbial N versus P scaling exponents across local sites

Large differences in the numerical values of microbial N versus P scaling exponents are observed across local sites (figure 4). MAP, soil pH, and soil total P play key roles in determining these numerical values across local sites, indicating the observation that soil microbial growth is influenced significantly by climate and soil conditions (Maria et al 2014). RDA indicates that soil total P is an important driver in determining the numerical value of microbial N versus P scaling exponents compared to MAP and soil pH (figure 5). This result supports the idea that soil P regulates microbial N and P stoichiometry and nutrient use strategies (Vitousek et al 2010, Camenzind et al 2018). In addition, our analyses provide empirical evidence that soils with high P concentrations (which support a high microbial composition and diversity) may favor fast growing microorganisms such as bacteria (Delgado-Baquerizo et al 2016). However, our understanding of the N versus P scaling relationship in bacteria and fungi is still very limited. Hence, future studies on bacterial and fungal N and P stoichiometry are clearly required.

5. Conclusions

To the best of our knowledge, this study is the first to comprehensively document pairwise soil MBC, MBN, and MBP concentrations and to provide detailed information on microbial N versus P scaling relationships for different ecosystems, soil depths, latitude zones, and local sites at the global scale. Based on the pooled data, the numerical value of the microbial N versus P scaling exponent is 0.89 as well as for different ecosystems, supporting the growth rate hypothesis. However, the data also indicate that the numerical value of the N versus P scaling exponent differs as a function of soil depth, latitudinal zones, and across local sites. Soil total P appears to be the most important factor explaining differences in the numerical values of the microbial N versus P scaling exponent. These findings highlight

the importance of climate and soil factors in shaping soil microbial investment strategies and improve our understanding of nutrient cycling in plant-soilmicrobe networks. However, it is clear that additional research is required to understand the influence of climate and soil conditions on microbial N versus P scaling relationships, especially from low- and high-latitude ecosystems. This need is particularly important in the context of temperature-controlled studies to address how global climate change may affect soil temperatures and microbial metabolic and growth rates.

Data availability statement

All data that support the findings of this study are included within the article (and any supplementary files).

Acknowledgments

This study was supported by the National Key Research and Development Program of China (2016YFC0501903), and the Natural Science Foundation of Qinghai Province of China (2020-ZJ-925).

Conflict of interest

The authors declare no competing financial interests.

Author contributions

Z Wang, H Huang and B Yao conceived the study with M Wang and J Wright wrote the first draft of the paper. All authors contributed to the writing of the paper.

ORCID iDs

Zhiqiang Wang lo https://orcid.org/0000-0002-2627-7062

Huifeng Hu is https://orcid.org/0000-0003-2223-170X

Yuanhe Yang ^(b) https://orcid.org/0000-0002-5399-4606

Philippe Ciais (a) https://orcid.org/0000-0001-8560-4943

Ashley P Ballantyne bhttps://orcid.org/0000-0003-1532-5126

Heng Huang bhttps://orcid.org/0000-0001-5552-3333

Buqing Yao (1) https://orcid.org/0000-0001-7111-7394

References

Ågren G I 2008 Stoichiometry and nutrition of plant growth in natural communities *Annu. Rev. Ecol. Evol. Syst.* **39** 153–70

- Allison S D, Wallenstein M D and Bradford M A 2010 Soil-carbon response to warming dependent on microbial physiology *Nat. Geosci.* **3** 336–40
- Anderson T H and Domsch K H 1993 The metabolic quotient for CO₂ (qCO₂) as a specific activity parameter to assess the effects of environmental conditions, such as pH, on the microbial biomass of forest soils *Soil Biol. Biochem.*25 393–5
- Bardgett R D, Streeter T C and Bol R 2003 Soil microbes compete effectively with plants for organic-nitrogen inputs to temperate grasslands *Ecology* **84** 1277–87
- Camenzind T, Hättenschwiler S, Treseder K K, Lehmann A and Rillig M C 2018 Nutrient limitation of soil microbial processes in tropical forests *Ecol. Monogr.* **88** 4–21
- Chapin F S III 1980 The mineral-nutrition of wild plants *Annu*. *Rev. Ecol. Evol. Syst.* **11** 233–60
- Chen Y L *et al* 2016 Linking microbial C:N:P stoichiometry to microbial community and abiotic factors along a 3500-km grassland transect on the Tibetan Plateau *Glob. Ecol. Biogeogr.* 25 1416–27
- Cleveland C C and Liptzin D 2007 C:N:P stoichiometry in soil: is there a 'Redfield ratio' for the microbial biomass? *Biogeochemistry* **85** 235–52
- Delgado-Baquerizo M, Reich P B, Khachane A N, Campbell C D, Thomas N, Freitag T E, AlSoud W A, Sorensen S J, Bardgett R D and Singh B K 2016 It is elemental: soil nutrient stoichiometry drives bacterial diversity *Environ. Microbiol.* 19 1176
- Díaz S, Hodgson J G, Thompson K, Cabido M and Zak M R 2004 The plant traits that drive ecosystems: evidence from three continents *J. Veg. Sci.* **15** 295–304
- Elser J J et al 2003 Growth rate-stoichiometry couplings in diverse biota Ecol. Lett. 6 936–43
- Elser J J, Sterner R W, Gorokhava E, Fagan W F, Markow T A, Cotner J B, Harrison J F, Hobbie S E, Odell G M and Weider L J 2000 Biological stoichiometry from genes to ecosystems *Ecol. Lett.* 3 540–50
- Fierer N and Jackson R B 2006 The diversity and biogeography of soil bacterial communities *Proc. Natl Acad. Sci. USA* 103 626–31
- Fierer N, Schimel J P and Holden P A 2003 Variations in microbial community composition through two soil depth profiles *Soil Biol. Biochem.* **35** 167–76
- Fierer N, Strickland M S, Liptzin D, Bradford M A and Cleveland C C 2009 Global patterns in belowground communities *Ecol. Lett.* **12** 1238–49
- Geng Y, Wang L, Jin D M, Liu H Y and He J S 2014 Alpine climate alters the relationships between leaf and root morphological traits but not chemical traits *Oecologia* **175** 445–55
- Güsewell S 2004 N:P ratios in terrestrial plants: variation and functional significance *New Phytol.* **164** 243–66
- Han W X, Fang J Y, Guo D L and Zhang Y 2005 leaf nitrogen and phosphorus stoichiometry across 753 terrestrial plant species in China *New Phytol.* **168** 377–85
- Hartman W H and Richardson C J 2013 Differential nutrient limitation of soil microbial biomass and metabolic quotients (qCO₂): is there a biological stoichiometry of soil microbes? *PLoS One* **8** e57127
- Hedin L O 2004 Global organization of terrestrial plant-nutrient interactions *Proc. Natl Acad. Sci. USA* **101** 10849–50
- Heuck C, Weig A and Spohn M 2015 Soil microbial biomass C:N:P stoichiometry and microbial use of organic phosphorus Soil Biol. Biochem. 85 119–29
- Jenkinson D S, Andrew S P S, Lynch J M, Goss M J and Tinker P B 1990 The turnover of the organic carbon and nitrogen in soil *Phil. Trans. R. Soc.* B **329** 361–8
- Jenkinson D S, Brookes P C and Powlson D S 2004 Measuring soil microbial biomass *Soil Biol. Biochem.* **36** 5–7
- Kerkhoff A J, Fagan W F, Elser J J and Enquist B J 2006 Phylogenetic and growth form variation in the scaling of nitrogen and phosphorus in the seed plants Am. Nat. 168 E103–22

- Li P, Yang Y H, Han W X and Fang J Y 2014 Global patterns of soil microbial nitrogen and phosphorus stoichiometry in forest ecosystems *Glob. Ecol. Biogeogr.* **23** 979–87
- Li Y, Wu J S, Liu S L, Shen J L, Huang D Y, Su Y R, Wei W X and Syers J K 2012 Is the C:N:P stoichiometry in soil and soil microbial biomass related to the landscape and land use in southern subtropical China? *Glob. Biogeochem. Cycles* **26** GB4002
- Liu Y Q *et al* 2020 Comparison of patterns of microbial C:N:P stoichiometry between topsoil and subsoil along an aridity gradient *Biogeosciences* 17 2009–19
- Manzoni S, Taylor P, Richter A, Porporato A and Ågren G I 2012 Environ- mental and stoichiometric controls on microbial carbon-use efficiency in soils *New Phytol.* **196** 79–91
- Maria M, Wolfgang W, Sophie Z B and Andreas R 2014 Stoichiometric imbalances between terrestrial decomposer communities and their resources: mechanisms and implications of microbial adaptations to their resources *Front. Microbiol.* **5** 22
- McGroddy M E, Daufresne T and Hedin L O 2004 Scaling of C:N:P stoichiometry in forests worldwide: implications of terrestrial Redfield-type ratios *Ecology* **85** 2390–401
- Moorhead D L, Lashermes G and Sinsabaugh R L 2012 Atheoretical model of C- and N-acquiring exoenzyme activities,which balances micro-bial demands during decomposition *Soil Biol. Biochem.* **53** 133–41
- Mooshammer M *et al* 2014b Adjustment of microbial nitrogen use efficiency to carbon: nitrogen imbalances regulates soil N cycling *Nat. Commun.* **5** 3694
- Mooshammer M, Wanek W, Zechmeister-Boltenstern S and Richter A 2014a Stoichiometric imbalances between terrestrial decomposer communities and their resources: mechanisms and implications of microbial adaptations to their resources *Front. Microbiol.* **4** 22
- Niklas K J 2006 A phyletic perspective on the allometry of plant biomass-partitioning patterns and functionally equivalent organ-categories *New Phytol.* **171** 27–40
- Niklas K J and Cobb E D 2005 N, P, and C stoichiometry of *Eranthis hyemalis* (Ranunculaceae) and the allometry of plant growth *Am. J. Bot.* **92** 1256–63
- Niklas K J, Owens T, Reich P B and Cobb E D 2005 Nitrogen/phosphorus leaf stoichiometry and the scaling of plant growth *Ecol. Lett.* **8** 636–42
- R Development Core Team 2017 R: a language and environment for statistical computing (Vienna: R Foundation for Statistical Computing)
- Redfield A C 1958 The biological control of chemical factors in the environment *Am. Sci.* **46** 205–21
- Reich P B and Oleksyn J 2004 Global patterns of plant leaf N and P in relation to temperature and latitude *Proc. Natl Acad. Sci. USA* **101** 11001–6
- Reich P B, Oleksyn J, Wright I J, Niklas K J, Hedin O L and Elser J J 2010 Evidence of a general 2/3-power law of scaling leaf nitrogen to phosphorus among major groups and biomes *Proc. R. Soc.* B 277 877–83
- Singh J S and Gupta V K 2018 Soil microbial biomass: a key soil driver in management of ecosystem functioning *Sci. Total Environ.* **634** 497–500
- Sistla S A and Schimel J P 2012 Stoichiometric flexibility as a regulator of carbon and nutrient cycling in terrestrial ecosystems under change *New Phytol.* **196** 68–78
- Spohn M 2016 Element cycling as driven by stoichiometric homeostasis of soil microorganisms *Basic Appl. Ecol.* 17 471–78
- Sterner R W and Elser J J 2002 Ecological Stoichiometry: The Biology of Elements from Molecules to the Biosphere (Princeton, NJ: Princeton University Press)
- Tian D *et al* 2017 Global leaf nitrogen and phosphorus stoichiometry and their scaling exponent *Natl Sci. Rev.* **5** 728–39
- Tischer A, Potthast K and Hamer U 2014 Land-use and soil depth affect resource and microbial stoichiometry in a tropical

mountain rainforest region of southern Ecuador Oecologia 175 375–93

- Van Der Heijden M G A, Bardgett R D and Van Straalen N M 2008 The unseen majority: soil microbes as drivers of plant diversity and productivity in terrestrial ecosystems *Ecol. Lett.* 11 296–310
- Vance E D, Brookes P C and Jenkinson D S 1987 An extraction method for measuring soil microbial C Soil Biol. Biochem. 19 703–7
- Vitousek P M, Porder S, Houlton B Z and Chadwick O A 2010 Terrestrial phosphorus limitation: mechanisms, implications, and nitrogen-phosphorus interactions *Ecol. Appl.* **20** 5–15
- Wang Z Q, Bu H Y, Wang M C, Huang H and Niklas K J 2020 Allocation strategies for seed nitrogen and phosphorus in an alpine meadow along an altitudinal gradient on the Tibetan Plateau Front. Plant Sci. 11 614644
- Wang Z Q, Huang H, Li X W, Mao K S, Ran J Z and Deng J M 2018 Allocation of nitrogen and phosphorus within and between needles, stems and roots of Picea seedlings *Nord. J. Bot.* 36 e01952
- Wang Z Q, Yu K L, Lv S Q, Niklas K J, Mipam T D, Crowther T W, Umaña M N, Zhao Q, Huang H and Reich P B 2019 The scaling of fine root nitrogen versus phosphorus in terrestrial plants: a global synthesis *Funct. Ecol.* 33 2081–94

- Warton D I, Wright I J, Falster D D and Westoby M 2006 Bivariate line-fitting methods for allometry *Biol. Rev.* 81 259–91
- Wright I J et al 2004 The worldwide leaf economics spectrum Nature 428 821–7
- Wright I J *et al* 2005 Assessing the generality of global leaf trait relationships *New Phytol.* **166** 485–96
- Xu X F, Thornton P E and Post W M 2013 A global analysis of soil microbial biomass carbon, nitrogen and phosphorus in terrestrial ecosystems *Glob. Ecol. Biogeogr.* 22 737–49
- Yuan Z Y, Chen H Y H and Reich P B 2011 Global-scale latitudinal patterns of plant fine-root nitrogen and phosphorus *Nat. Commun.* **2** 344
- Zechmeister-Boltenstern S, Keiblinger K M, Mooshammer M, Peñuelas J, Richter A, Sardans J and Wanek W 2015 The application of ecological stoichiometry to plant-microbial-soil organic matter transformations *Ecol. Monogr.* 85 133–55
- Zhang J H, He N P, Liu C C, Xu L, Yu Q and Yu G R 2018 Allocation strategies for nitrogen and phosphorus in forest plants *Oikos* 127 1506–14
- Zhao N, Yu G R, He N P, Xia F C, Wang Q F, Wang R L, Xu Z W and Jia Y L 2016 Invariant allometric scaling of nitrogen and phosphorus in leaves, stem, and fine roots of woody plants along an altitudinal gradient *J. Plant Res.* 129 647–57