

Discovery of a silicate rock-boring organism and macrobioerosion in fresh water

Ivan N. Bolotov, Olga V. Aksenova, Torkild Bakken, Christopher J. Glasby, Mikhail Yu. Gofarov, Alexander V. Kondakov, Ekaterina S. Konopleva, Manuel Lopes-Lima, Artyom A. Lyubas, Yu Wang, et al.

▶ To cite this version:

Ivan N. Bolotov, Olga V. Aksenova, Torkild Bakken, Christopher J. Glasby, Mikhail Yu. Gofarov, et al.. Discovery of a silicate rock-boring organism and macrobioerosion in fresh water. Nature Communications, 2018, 9, 234, p. 1-23. 10.1038/s41467-018-05133-4 . insu-03661396

HAL Id: insu-03661396 https://insu.hal.science/insu-03661396

Submitted on 6 May 2022

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License



ARTICLE

DOI: 10.1038/s41467-018-05133-4

OPEN

Discovery of a silicate rock-boring organism and macrobioerosion in fresh water

Ivan N. Bolotov ^{1,2}, Olga V. Aksenova^{1,2}, Torkild Bakken³, Christopher J. Glasby⁴, Mikhail Yu. Gofarov^{1,2}, Alexander V. Kondakov^{1,2}, Ekaterina S. Konopleva^{1,2}, Manuel Lopes-Lima ^{5,6}, Artyom A. Lyubas^{1,2}, Yu Wang^{7,8}, Andrey Yu. Bychkov^{9,10}, Agniya M. Sokolova^{11,12}, Kitti Tanmuangpak¹³, Sakboworn Tumpeesuwan¹⁴, Ilya V. Vikhrev^{1,2}, J. Bruce H. Shyu⁸, Than Win ¹⁵ & Oleg S. Pokrovsky ^{16,17}

Macrobioerosion is a common process in marine ecosystems. Many types of rock-boring organisms break down hard substrates, particularly carbonate rocks and calcareous structures such as dead corals and shells. In paleontology, the presence of rocks with boreholes and fossil macroboring assemblage members is one of the primary diagnostic features of shallow marine paleo-environments. Here we describe a silicate rock-boring organism and an associated community in submerged siltstone rock outcrops in Kaladan River, Myanmar. The rock-boring mussel *Lignopholas fluminalis* is a close relative of the marine piddocks, and its borings belong to the ichnospecies *Gastrochaenolites anauchen*. The neotectonic uplift of the area leading to gradual decrease of the sea level with subsequent shift from estuarine to freshwater environment was the most likely driver for the origin of this community. Our findings highlight that rocks with macroborings are not an exclusive indicator of marine paleo-ecosystems, but may also reflect freshwater habitats.

¹ Northern Arctic Federal University, Arkhangelsk, Russia 163002. ² Federal Center for Integrated Arctic Research, Russian Academy of Sciences, Arkhangelsk, Russia 163000. ³ Norwegian University of Science and Technology, NTNU University Museum, 7491 Trondheim, Norway. ⁴ Museum and Art Gallery of the Northern Territory, Darwin, NT 0820, Australia. ⁵ CIIMAR—Interdisciplinary Centre of Marine and Environmental Research, University of Porto, 4450-208 Matosinhos, Portugal. ⁶ CIBIO/InBIO—Research Center in Biodiversity and Genetic Resources, University of Porto, 4485-661 Vairão, Portugal. ⁷ Earth Observatory of Singapore, Nanyang Technological University, Singapore 639798, Singapore. ⁸ Department of Geosciences, National Taiwan University, Taipei 10617, Taiwan. ⁹ Faculty of Geology, Lomonosov Moscow State University, Moscow, Russia 119991. ¹⁰ V. I. Vernadsky Institute of Geochemistry and Analytical Chemistry, Russian Academy of Sciences, Moscow, Russia 119991. ¹¹ A. N. Severtsov Institute of Ecology and Evolution, Russian Academy of Sciences, Moscow, Russia 119071. ¹² N. K. Koltzov Institute of Developmental Biology, Russian Academy of Sciences, Moscow, Russia 119991. ¹³ Department of Science, Faculty of Science and Technology, Loei Rajabhat University, Loei 42000, Thailand. ¹⁴ Department of Biology, Faculty of Science, Mahasarakham University, Maha Sarakham 44150, Thailand. ¹⁵ Department of Zoology, Hpa-An University, Hpa-An, Kayin State 13017, Myanmar. ¹⁶ Geosciences and Environment Toulouse, UMR 5563 CNRS, 31400 Toulouse, France. ¹⁷ BIO-GEO-CLIM Laboratory, Tomsk State University, Tomsk, Russia 634050. Correspondence and requests for materials should be addressed to I.N.B. (email: inepras@yandex.ru)

B ioerosion is a process by which a living organism incises or bores different hard substrates (e.g. rocks, shells, corals, and bones) by mechanical disruption and/or chemical decomposition¹. The eroding activities of living organisms are important factors in marine sedimentation and benthic ecology². Bioerosion increases species diversity of marine hard substrate communities by increasing habitat complexity or as a result of the increase in accessible surface area for colonization³. Additionally, this process influences the evolution of coastal profiles over long timescales^{4,5}. With respect to their environmental impact, erosional organisms represent an important group of ecosystem engineers^{6,7}.

The investigation of bioerosion patterns supplies significant background information to other research areas (e.g., zoology, paleoecology, biogeochemistry, sedimentology, and biostratigraphy)^{1,8}. Macrobioerosion trace fossils are important indicators of intertidal and shallow subtidal marine paleo-environments^{9–11}, e.g., boring bivalves may be used as biological sea level indicators marking ancient shorelines^{2,11,12}. Boring organisms have a great economic impact because they damage ships, fishing equipment, archeological heritage, various infrastructures in marine environments, and the shells of aquaculture molluscs^{12–14}.

Rock borers occur in a broad range of taxonomic groups, including bivalves, gastropods, polychaetes and sipunculans (Annelida), sea urchins, sponges, and others^{8-10,15}. The piddocks, or rock-boring mussels (Mollusca: Bivalvia: Pholadidae), are predominantly marine animals specialized for boring into a plethora of available substrates such as soft silicate and carbonate rocks, clay, corals, wood, and peat^{3,14,16}. The genera Lignopholas Turner, 1955 and Martesia Sowerby 1824 contain several estuarine and marine wood-boring bivalve species, including a single species that can bore into living mangrove trees¹⁷. There is a unique record of clavate borings and fossil specimens of Martesia sp. from Cretaceous Burmese amber¹¹. However, Lignopholas fluminalis was also collected from soft argillite rocks and brickworks in brackish sections of rivers in India and Myanmar¹⁷⁻¹⁹. The piddocks have developed a variety of adaptations to accommodate their rock-boring behavior and make their borings mechanically, by scraping at the substratum 20 .

In the marine environment, almost all macroborings are recorded in calcareous substrates, e.g., corals, shells, and carbonate rocks^{10,21}. There are far fewer examples of fossil and recent marine borings found in non-calcareous rocks and minerals such as siltstones, quartzites, basalts, andesites, dolerites, gneisses, and others^{17,21,22}. From an ichnological point of view, clavate macroborings in hardgrounds and in fossil wood belong to two ichnogenera, i.e., *Gastrochaenolites* Leymerie 1842 and *Teredolites* Leymerie 1842 for borings in lithic and lignic substrates, respectively²³. The *Gastrochaenolites* borings are known from the Early Ordovician to Recent and they are mainly produced by marine endolithic bivalves^{8,9}. However, several recent gastropods and sipunculan worms may also produce *Gastrochaenolites*⁹.

In general, macrobioerosion was considered an exclusively marine and, to a lesser degree, an estuarine process^{8,10}. However, we discovered the macrobioerosion of silicate siltstone rocks caused by a rock-boring mussel species in a freshwater section of the Kaladan River, Myanmar, which greatly expands our knowledge of bioerosion. Our phylogenetic and biogeographic modeling shows that at least three members of this rock-boring assemblage are relict marine-derived lineages, which emphasizes a broad-scale expansion of saltwater taxa into freshwater environment in Southeast Asia.

Results

Discovery of freshwater macrobioerosion. The site is located in the middle reaches of Kaladan River: 21.0094° N, 92.9813° E, altitude of 11 m a.s.l., Rakhine State, western Myanmar (Fig. 1a, b).

The rock-boring assemblage is associated with submerged black siltstone rocks located in the lower part and bottom of the river valley, with an above-water outcrop near the bioerosion site (Fig. 1b–d). The submerged rocks are located within a riverine section of ~500 m in length and of ~300 m in width, with depth values of 1-2 m and more.

Freshwater environment. The model of tidal influence reveals that the distance between the rock-borer's site and the upper level of the Kaladan estuary is 71 km (Fig. 2). Dating of the shift from estuarine to freshwater environment at the site suggests that it was approximately 3.5–14 Kyr ago, i.e., not earlier than the Late Pleistocene (Fig. 2). The analysis of a water sample confirms that it is a freshwater section of the river, with a chloride ion concentration of 2.7 mg/L, sodium ion concentration of 11.6 mg/L, and a total salinity of 0.16‰ (Supplementary Table 1).

Rock substrate. According to our grain-size analyses (Supplementary Fig. 1), the substrate belongs to siltstone (aleurolite) rocks due to a primary grain size of $2-62 \,\mu$ m. The backscattered electron (BSE) images and X-ray diffraction (XRD) analyses indicate that the rock consists of quartz, feldspar, clay minerals, chlorite, and mica grains (Supplementary Table 2 and Supplementary Fig. 1). The X-ray fluorescence (XRF) analysis reveals the dominance of silicon dioxide ($56.9 \pm 0.5\%$) and aluminum oxide ($17.0 \pm 0.4\%$) (Supplementary Table 3). A mean microindentation hardness (Vickers test) value of the substrate rock is 0.62 GPa with a range of 0.50–0.72 GPa (mean HV value = $61.9 \,\text{kgf} \times \text{mm}^{-2}$, range = $52.1-73.5 \,\text{kgf} \times \text{mm}^{-2}$).

Ichnotaxonomy. The macroborings from the Kaladan River are clavate, circular in cross-section throughout the length, expanded gradually below the aperture, with a greatest diameter about three-fourths of the depth; bases are rounded; sides are smooth, without clear circular or spiral bioglyph; a distinguishable neck is lacking (Fig. 3f). With respect to the lithic substrate and morphological features²⁴, these borings correspond well to the ichnospecies *Gastrochaenolites anauchen* Wilson & Palmer, 1998.

Rock-boring species. Freshwater rock borers heavily invaded the submerged section of siltstone rocks that have numerous piddocklike boreholes (Fig. 3a-f). Empty borings are sometimes filled with clay (Fig. 3c). This boring species was determined as Lignopholas fluminalis on the basis of morphological characters (Table 1, Fig. 4a and Supplementary Note 1). The COI gene sequences indicates that it is related to the marine piddock species Barnea davidi but with low similarity (81.4%), while the nuclear 18S rRNA gene sequences show a very close relationship with several other marine Barnea spp. (similarity of 98-99%) (Supplementary Table 4). The Bayesian and maximum likelihood phylogenies reveal that this species is a sister lineage of Barnea spp. The ancestral area reconstruction models suggest that the most recent common ancestor (MRCA) of the *Lignopholas* + *Barnea* clade was a marine mussel (Fig. 5a and Table 2). The boring mechanism of L. fluminalis is not known, but their boreholes should be made mechanically as do its marine relatives²⁰.

Associated nestling species. We recorded six species of nestling macroinvertebrates that appear to be associated with this rockborer's ecosystem (Table 1, Supplementary Table 4, and Supplementary Note 1). An integrative study of the species using morphological and molecular approaches indicates that they include two clams, *Scaphula deltae* (Arcidae) and *Novaculina gangetica* (Pharidae), a gastropod, *Clithon* cf. *reticularis* (Neritidae), two polychaetes, *Namalycastis indica* and *Neanthes*



Fig. 1 Freshwater bioerosion site, Kaladan River, Myanmar. **a** Map of Myanmar with location of the downstream area of Kaladan River (red frame). **b** Map of the study site. The violet asterisk indicates the freshwater rock-borer's site, and white circles indicate the levels of river water above sea level. The yellow circles indicate main towns. The maps were created using ESRI ArcGIS 10 software (www.esri.com/arcgis); the topographic base of the maps was created with Natural Earth Free Vector and Raster Map Data (www.naturalearthdata.com), Vector Map (VMap) Level 0 (http://earth-info.nga.mil/publications/vmap0.html), and ASTER GDEM (https://lpdaac.usgs.gov/node/1079) (Maps: Mikhail Yu. Gofarov). **c** River site with freshwater rock-borer's ecosystem. The red arrow indicates siltstone rocks, a substrate of rock-boring bivalves. **d** Fragment of the black siltstone outcrop, the submerged part of which forms the basis of freshwater rock-borer ecosystem. (Photos: Olga V. Aksenova)



Fig. 2 Model of neotectonic evolution of the lower course of the Kaladan River. The long-term uplift rate *R* of 0.5–2 mm/year was applied (see Methods section). The model assumes a gradual shift from estuarine to freshwater environment at the rock-borer's site due to the tectonic uplift with subsequent adaptation of the population of the brackish rock-boring species *Lignopholas fluminalis* to freshwater habitat. (Model: Ivan N. Bolotov, Mikhail Yu. Gofarov, Yu Wang, & J. Bruce H. Shyu)

meggitti (Nereididae), and a sponge, *Corvospongilla ultima* (Spongillidae) (Fig. 4b–g). The results of phylogenetic and biogeographic modeling reveal that both clam taxa, *S. deltae* and *N. gangetica*, most likely represent the relict marine-derived freshwater lineages (Fig. 5b, c and Table 2). *N. gangetica* is a close relative of the *Sinonovacula* + *Pharella* clade (Fig. 5b and Supplementary Table 4). *S. deltae* clusters together with several members of the Noetiidae but not with the Arcidae (Fig. 5c, Supplementary Fig. 2, and Supplementary Table 4).

Discussion

Here we present the discovery of a rock-boring bivalve species and associated hard substrate community in fresh water. Our record is of exceptional paleontological, archeological, and environmental importance, because, to our knowledge, until now rock-borers were known only among marine and estuarine groups of macroinvertebrates^{8–11,14,15}. From freshwater habitats, only a few records of carbonate microbioerosion structures have been described, i.e., micro-borings in mollusc shells associated with a group of minute shell-boring polychaetes^{25,26}. To the best of our knowledge, silicate macrobioerosion in fresh water was previously unknown.

Our findings highlight that rocks with boreholes and fossil members of rock-borer's assemblages are not necessarily an absolute indicator of shallow marine paleo-environments, but may also reflect past freshwater riverine ecosystems. Additionally, we found that the three most important members of the assemblage, i.e., *L. fluminalis, S. deltae*, and *N. gangetica*, are marine-derived, secondary freshwater taxa. This finding is in full agreement with the hypothesis of Annandale²⁷, who assumed that *Scaphula* and *Novaculina* are freshwater genera of relict marine origin. In contrast, at least one member of the rockborer's assemblage, the sponge *C. ultima*, is an entirely freshwater species that is not known from brackish water bodies²⁸. The freshwater fauna of the lower and middle reaches of

the Kaladan and Lemro River systems appears to be rather a derivative of the Indian fauna (e.g., indicator taxa such as *N. gangetica*^{29,30}, *Lamellidens* aff. *marginalis*^{31,32}, and *C. ultima*²⁸), but the lack of comparable molecular data from India precludes any final biogeographic conclusion. Our modeling suggests that the shift from an estuarine to a freshwater environment at the rock-borer's site was a relatively recent event that occurred after the Late Pleistocene. These results support the hypothesis that a few marine or brackish species appear to have the unusual ability to invade freshwater over rapid time scales³³. However, we show that the invasion of a rock-boring species to a freshwater environment may lead to the origin of an associated hard substrate community representing a mix of primary estuarine and freshwater taxa.

The Gastrochaenolites has been considered a marine ichnotaxon produced by endolithic bivalves^{8,23} and, to a lesser degree, by gastropods and sipunculan worms9. Our discovery of a freshwater silicate rock-boring organism indicates that fossil macroborings in non-calcareous substrates may have been produced not only by marine invertebrates but also by freshwater lineages of endolithic bivalves. From this perspective, the discovery of fossil pholadid bivalves in Cretaceous Burmese amber may also indicate rather a freshwater paleo-environment than the proximity of resin-producing forests to brackish waters¹¹. The ichnospecies G. anauchen was described from the Early Pennsylvanian deposits of the USA²⁴. Our new record supports the hypothesis of Wilson and Palmer²⁴ that the Pennsylvanian borings were formed by endolithic bivalves, because the recent Kaladan borings are morphologically identical to those from the Paleozoic coarse grainstone cobbles. Wilson and Palmer²⁴ associated such smooth-sided clavate borings with lithophagid bivalves, but the borings of the pholadid species L. fluminalis are also characterized by smooth sides. The empty borings in the freshwater rock-boring site at the Kaladan River are often inhabited by nestling bivalve species, i.e., S. deltae and N. gangetica. This observation is in agreement with paleontological records that fossil bivalve associations are complicated by frequent records of nestling bivalves, e.g., mytilids and Hiatella sp. (Hiatellidae), in the vacated borings 34 .

Human-mediated biological invasions of various species outside their native ranges are a global-scale process³⁵, which are accompanied by climatic niche shifts³⁶ and rapid evolution of dispersal ability³⁷ in invaders. Among the piddocks, alien populations of Martesia striata were recorded from the Mediterranean Sea, Hawaii, Islands of the British Isles, and Florida³⁸. There is a possibility of further invasion of the freshwater rock-borer's lineage outside the Kaladan River, because of the Kaladan Multi-Modal Transit Transport Project that is being implemented by the Government of India and the Government of Myanmar. The project involves the development of the transit transport system to Mizoram through Myanmar, with a 158 km inland water transport segment via the Kaladan River³⁹, which may facilitate a broad expansion of the rock-borer. The larval development of L. fluminalis is unknown, but it is most likely that this species has a planktonic larva as the marine pholadids, e.g., Barnea and Martesia (Supplementary Note 1). A successful invasion of this freshwater lineage into surrounding larger Asian river basins, including the Ganges, Salween, and Mekong, can lead to unpredictable consequences for native ecosystems that can be comparable to those from other invaders with planktonic larvae^{40,41}. In contrast, a freshwater population of the rockboring mussel is of exceptional importance for scientific research, as it probably represents a lineage with unique physiological adaptations to survive in freshwater environments. Taking into account its possible local range and ancient origin, this lineage should be a focus of special research and conservation efforts.



Fig. 3 Blocks of siltstone rocks with borings and living macroinvertebrates from the freshwater bioerosion site at the Kaladan River, Myanmar. **a**, **b** Rockboring mussels, *Lignopholas fluminalis*, in their clavate borings (ichnospecies: *Gastrochaenolites anauchen*). **c** Arc clams, *Scaphula deltae*, and empty borings filled with clay, a habitat of polychaetes *Neanthes meggitti* and *Namalycastis indica* (Nereididae). **d** Arc clam, *Scaphula deltae*, in an empty boring. **e** Jackknife clam, *Novaculina gangetica*, and sponge, *Corvospongilla ultima*. Scale bars = 10 mm (Photos: Olga V. Aksenova). **f** Longitudinal cross-section of the boring, *Gastrochaenolites anauchen*. Scale bar = 5 mm. (Photo: Ilya V. Vikhrev)

Our discovery of a freshwater rock-boring assemblage in the Kaladan is in agreement with numerous occurrences of marinederived groups in rivers of the Oriental Region, which have attracted the attention of scientists for many decades^{27,30,42-45}. Fossil freshwater shark assemblages were discovered from the Late Jurassic and Early Cretaceous deposits of the Khorat Plateau in Thailand 46,47 indicating that the colonization of Asian river systems by marine animals started as early as the mid-Mesozoic. Molecular studies generally support the primary marine origin of most recent marine-derived taxa in tropical fresh waters. There were at least three separate freshwater expansions by stingrays in Southeast Asia, and brackish water habitats may have played an important role for these events as evolutionary bottlenecks⁴⁸. Two freshwater colonization events were recorded in ariid catfishes⁴⁹ and at least one event occurs in Southeast Asian freshwater pufferfishes⁵⁰. The phylogeography of a killifish species, Aplocheilus panchax, indicates multiple freshwater colonization events during the Pleistocene⁵¹. Freshwater prawns of the genus Macrobrachium independently colonized fresh waters at least five times⁵² and freshwater polychaetes of the genus Namanereis show two distinct invasions⁵³.

In summary, Southeast Asia seems to be a long-term arena for wide-scale expansion of marine taxa into fresh water, examples of which are known within different animal families. This pattern may be associated with the presence of a plethora of huge estuarine areas, which may facilitate the adaptation of saltwater organisms into freshwater environments. The land-level change resulting in the combination of progressive tectonic uplift of the mainland⁵⁴ and a gradual decrease of the sea level⁵⁵ may also have contributed to this process through a slow rise of river catchment elevation with a subsequent decrease in the level of tidal saltwater influence in the lower sections of the rivers (Fig. 2). Such a neotectonic evolutionary model seems to be the most appropriate explanation for origin of the freshwater rock-borer's community in Kaladan River. Several more ancient freshwater colonization events are consistent with the collision of the Asian and Indian plates and the subsequent Tethyan regression, which advanced the cohesive zone shift from shoal sea waters to freshwater environments⁴⁴.

The freshwater lineage of *L. fluminalis* from the Kaladan River is an example of a rock-boring freshwater organism that provides the reliable evidence for freshwater macrobioerosion in silicate rock. Phylogenetically, the rock-boring species is a close relative

Phylum: Class:	Species	Role in the	Origin of	Acc. numbers of reference sequences				
Family		community, preferred environment	the family	СОІ	16S rRNA	28S rRNA	18S rRNA	
Mollusca: Bivalvia:	Lignopholas fluminalis	Rock-boring	Marine ¹⁴	MF958974	n/a	n/a	n/a	
Pholadidae	(Blanford, 1867)	species, brackish		MF958975	n/a	n/a	MF959022	
				MF958976	n/a	n/a	MF959023	
				MF958977	n/a	n/a	MF959024	
				MF958978	n/a	n/a	MF959025	
				MF958979	n/a	n/a	MF959026	
Mollusca: Bivalvia:	Scaphula deltae	Nestling species,	Marine ⁷³	MF958980	n/a	n/a	n/a	
Arcidae	(Blanford, 1867)	from brackish to freshwater		MF958981	n/a	MF959007	n/a	
				MF958982	n/a	n/a	n/a	
				MF958983	n/a	MF959008	n/a	
				MF958984	n/a	MF959009	n/a	
				MF958985	n/a	MF959010	n/a	
Mollusca: Bivalvia:	Novaculina gangetica	Nestling species,	Marine ⁷⁴	MF958986 *	MF958997 *	MF959011 *	n/a	
Pharidae	(Benson, 1830)	from brackish to freshwater		MF958987 *	MF958998 *	MF959012 *	n/a	
				MF958988 *	MF958999 *	MF959013 *	n/a	
				MF958989	MF959000	MF959000x	n/a	
				MF958990	MF959001	MF959015	n/a	
				MF958991	MF959002	MF959016	n/a	
Mollusca: Gastropoda:	Clithon cf. reticularis	Nestling species,	Marine ⁷⁵	MF958992	MF959003	MF959017	n/a	
Neritidae	(Sowerby, 1838)	probably brackish		MF958993	MF959004	MF959018	n/a	
Annelida: Phyllodocida:	Neanthes meggitti	Nestling species,	Marine ⁷⁶	MF958994	MF959006	MF959020	n/a	
Nereididae	(Monro, 1931)	brackish						
Annelida: Phyllodocida:	Namalycastis indica	Nestling species,	Marine ⁷⁶	MF958995	MF959005	MF959019	n/a	
Nereididae	(Southern, 1921)	freshwater		MG759522	MG759523	MG759524	n/a	
Porifera: Demospongiae: Spongillidae	Corvospongilla ultima (Annandale, 1910)	Nestling species, freshwater	Freshwater ⁷⁷	MF958996	n/a	MF959021	n/a	
n/a—not available *Specimens that were collected	outside the rock-borer site, fron	n another river (Myanmar: I	emro River, 20.6150° ا	N, 93.2481° E)				

Table 1 Taxonomy, ecology, and reference sequences of freshwater rock-borer community's members from Kaladan River, western Myanmar

of the marine piddocks *Barnea* spp. The borings of this species in siltstones belong to the ichnospecies *G. anauchen* which expands the putative range of the saltwater *Gastrochaenolites* into fresh water.

The freshwater rock-borer community in Myanmar includes several nestling macroinvertebrate species, including the arc clam S. deltae and the jackknife clam N. gangetica which belong to relict marine-derived genera with brackish and freshwater species. Our tectonic modeling indicates that this unusual invertebrate community most likely originated approximately 3.5-14 Kyr ago via the neotectonic uplift of the area leading to a gradual lowering of the sea level and a subsequent shift from an estuarine to a freshwater environment. This example corresponds well to the modern data on multiple and wide-scale expansions of marine taxa into fresh water in Southeast Asia which began as early as the mid-Mesozoic. We assume that such a general biogeographic pattern may be associated with the combination of the progressive tectonic uplift of the mainland and with the subsequent decrease in the level of tidal saltwater influence in the lower sections of the rivers occurring in huge estuarine areas around the mainland. This neotectonic evolutionary model predicts the gradual shift in adaptive zone from shallow marine and brackish waters to freshwater habitats.

Our findings highlight that the rocks with macroborings and fossilized members of rock-boring communities are not a direct indicator of shallow marine paleo-environments, but may also reflect freshwater habitats. A rock-borer lineage from Kaladan River adapted to life in a freshwater environment and having putative planktonic larvae represents a potential international threat in the case it manages to invade the surrounding larger Asian river basins, including the Ganges, Mekong, and Salween Rivers.

Methods

Data sampling. The samples were collected from a site in the middle reach of Kaladan River. Several blocks with numerous borings and live representatives of the rock-boring community were obtained from submerged siltstone rocks at depths between 1.0–1.5 m using a large tommy-bar. A water sample was collected with a plastic bottle, filtered (0.45 µm) and stored in the dark before analysis by atomic absorption spectroscopy and ionic chromatography. Macroinvertebrate specimens were collected using forceps and immediately preserved in 96% ethanol. Additionally, four samples of siltstone fragments with rock-boring mussels and accessory taxa were conserved in containers with 96% ethanol.

Geographic and tectonic modeling. The model of the estuarine and freshwater sections of the Kaladan River was created using ESRI ArcGIS 10 software (www.esri.com/arcgis). The topographic base of the map was created with Natural Earth Free Vector and Raster Map Data (www.naturalearthdata.com), Vector Map (VMap) Level 0 (http://earth-info.nga.mil/publications/vmap0.html), and ASTER GDEM (https://lpdaac.usgs.gov/node/1079). The water levels were obtained from a topographic map (scale 1:500,000; The General Staff of the USSR, map nos. F46-3 and F46-4). The water level at the rock-borer's site was estimated using linear interpolation. We calculated the length of brackish section of the river based on the maximum tide height that was obtained from the Joint Archive for Sea Level⁵⁶. The land-level change rate (i.e. uplift and sea-level change rate) is needed to reconstruct neotectonic evolution of the lower course of the Kaladan River during the Late Holocene. Unfortunately, data for this part of Myanmar are extremely scarce and uneven. South of our research location, Than Tin Aung et al.⁵⁷, and Wang et al.⁵⁹ have attempted to obtain such rates on the basis of field surveys of uplifted sea-level indicators. Their observations pointed out that the long-term land-level change rates in this area contain significant spatial variations. For example, late Holocene average uplift rates at southwestern Cheduba Island can be as high as 3.5-5.2 mm/year, whereas the land-level change rate landward

ARTICLE



Fig. 4 Rock-boring and nestling macroinvertebrates associated with freshwater bioerosion site at the Kaladan River, western Myanmar. **a** *Lignopholas fluminalis*, a rock-boring species: (a1) Lateral view of an adult specimen with fully developed callum, (a2) Ventral view of apposed valves, (a3) Dorsal view of apposed valves, (a4) Anterior view, (a5) Frontal view of siphons, (a6) Lateral view of siphons, (a7) Lateral view of a young specimen, (a8) Ventral view of the anterior end, (a9) Dorsal view, (a10) Internal morphology of soft body, (a11) Frontal view of siphons (scale bars = 2 mm). **b** *Scaphula deltae*, an ark clam species: (b1) Lateral and (b2) dorsal view of a young specimen, (b3) Live clam attached by byssus in borehole, (b4) Lateral and (b5) frontal view of an adult specimen, (b8) Cardinal teeth of left valve, (b9) Cardinal teeth of right valve (scale bars = 2 mm). **c** *Novaculina gangetica*, a jackknife clam species: (c1) Lateral view of right and left shell valves (outside), (c2) Lateral view of right and left shell valves (inside), (c3) Shell variability, (c4) Live specimen with protruding foot and siphons (scale bars = 10 mm). **d** *Clithon* cf. *reticularis*, a gastropod species: (d1) Apertural view, (d2) Dorsal view, (d3) Apical view (scale bars = 2 mm). (e) *Corvospongilla ultima*, a sponge species: (e1) General view of a sponge body fragment (scale bar = 2 mm), (e2) Three adherent gemmules; gemmular cage covering the left gemmule is removed (scale bar = 500 µm). **f** *Neanthes meggitti*, a polychaete species: (f1) Dorsal view of complete specimen (scale bar = 2 mm), (f2) Dorsal view of the anterior end (scale bar = 0.5 mm). (Photos: Olga V. Aksenova [a, b, d, e1, f, and g], Ekaterina S. Konopleva and llya V. Vikhrev [**c**], and Agniya M. Sokolova [e2])



Fig. 5 Ultrametric Bayesian (BEAST 2) phylogenies showing the primary marine origin of rock-boring and nestling bivalve species from the bioerosion site in the Kaladan River, including **a** *Lignopholas fluminalis*, Pholadidae (*18S rRNA* + *28S rRNA*, 3033 bp), **b** *Novaculina gangetica*, Pharidae (*COI* + *16S rRNA* + *28S rRNA*, 2602 bp), and **b** *Scaphula deltae*, Arcidae (*28S rRNA* + *18S rRNA*, 2450 bp, full tree includes 44 taxa but the other clade of Arcidae is not shown here: see Supplementary Fig. 2). Black numbers near nodes are BPP values inferred from BEAST/BPP values inferred from MrBayes/BS values inferred from RAXML ("—" indicates a topological difference). Pie chaps on the nodes indicate the probabilities of certain ancestral areas with respect to combined results under three different models (S-DIVA, DEC, and S-DEC) inferred from RASP v. 3.2

along the eastern coast of Ramree Island may be less than 0.5 mm/year⁵⁹. These two islands are located further south of the Kaladan River, but within a ~500-km-long seismic patch that includes the downstream section of the Kaladan River⁶⁰. Closer to the Kaladan River, Than Tin Aung et al.⁵⁸ also reports similar variations just south of the town of Sittwe, where the land-level change rate changes from more than 4.5 mm/year at the seaward sites to ~2 mm/year at the landward sites. Since the Kaladan River is located near the landward sites in these studies, we conservatively applied a long-term land-level change rate *R* of 0.5–2 mm/year in our tectonic modeling.

Mineralogical analyses of rock substrate. The grain size analysis was carried out on an Analysette 22 MicroTec Plus Laser Particle Sizer (Fritsch GmbH-Milling and Sizing, Germany) using a rock sample dispersed in distilled water. An XRD approach was applied to estimate the mineral composition of the substrate using a DRON-3M diffractometer (Bourevestnik Inc., Russia). The BSE images were obtained on an electronic scanning microscope Jeol JSM-6480LV (Jeol Ltd., Japan) with energy-dispersive Oxford X-Max^N Silicon Drift Detector and crystaldiffractive INCA Wave-500 WDS spectrometer (Oxford Instrument Ltd., UK). The chemical composition of the rock samples was determined by an XRF analysis using a wavelength-dispersive XRF spectrometer PW 2400 (Philips Analytical, the Netherlands). The microindentation hardness (Vickers test) of the rocks was measured using a PMT-3M Vickers Microhardness Tester (LOMO, Russia) with 100 g load. The tester was calibrated using NaCl crystal with 10 g load. Several fragments of the substrate were placed into briquette and were fixed with epoxy glue with subsequent polishing of the surface. Five indentations were performed on each rock fragment, and both diagonals of indentation mark were measured. A mean microhardness value was calculated based on 20 measurements.

Ichnological identification of borings. The bivalve borings were identified based on morphological patterns and the type of substrate using the appropriate ichnotaxonomic works^{23,24}. Longitudinal cross-sections of the borings were used for morphological investigation.

Morphological identification of invertebrates. For the morphological study of invertebrate specimens, we used a stereomicroscope (Leica M165C, Leica Microsystems, Germany). The comparative analysis of bivalve taxa was carried out according to the shell shape, structure of the hinge, muscle attachment scars, and umbo position. A gastropod species was identified based on shell shape and marking patterns. The polychaete taxa were identified in accordance with morphology of the external body, everted pharynx, and structures of the parapodia and chaetae. Identification of the sponge species was carried out through analysis of skeleton elements using a light microscope (Olympus CX21, Olympus Corporation, Japan) and scanning electron microscope, SEM (Tescan Vega TS5130MM, Tescan Orsay Holding, Czech Republic). Spicules of each type were purified with potassium dichromate solution (2.5% CrO₃ in 50% H₂S), washed and mounted on a slide and specimen stub according to standard methods⁶¹. Measurements (n = 25) were performed under light microscope using an ocular micrometer; spine size was measured under SEM; spicule size is presented as minimum-mean-maximum dimensions.

DNA extraction and molecular analyses. New sequences were obtained from 24 invertebrate specimens belonging to seven species that were collected from the Kaladan and Lemro rivers, western Myanmar (Table 1). Total genomic DNA was extracted from 96% ethanol-preserved tissue samples using the NucleoSpin[®] Tissue Kit (Macherey-Nagel GmbH & Co. KG, Germany), according to the

Table 2 Evolutionary origin of bivalve lineages from the freshwater rock-borer community in western Myanmar on the basis of ancestral area reconstruction analyses

MRCA	The most likely ancestral area	Biogeographic event (origin of	Probability of the most likely ancestral area			
		estuarine/freshwater lineages)	S-DIVA	DEC	S-DEC	Combined results
Lignopholas + Barnea	Marine	Dispersal	1.00	0.52	0.52	0.68
Novaculina + (Pharella + Sinonovacula)	Marine	Dispersal	1.00	0.64	0.79	0.81
Scaphula + Noetia	Marine	Dispersal	1.00	0.87	0.90	0.92

manufacturer's protocol. For molecular analyses, we obtained partial sequences of the following markers: the mitochondrial *cytochrome c oxidase subunit I gene (COI)* and *16S ribosomal RNA (16S rRNA)*, and the nuclear *28S ribosomal RNA (28S rRNA)*. We were unable to obtain the *28S rRNA* gene sequences from the rockboring species, and we therefore sequenced another nuclear marker, i.e., the *18S ribosomal RNA (18S rRNA)*. Additionally, sequencing of the *16S rRNA* gene from samples of *L. fluminalis* and *S. deltae* was also unsuccessful (Table 1).

PCR primers are shown in Supplementary Table 5. We applied four marker-specific PCR algorithms as follows: (i) COI: 95 °C (4 min), 94 °C (35–37 repeats, 50 s), 50 °C (50 s), 72 °C (50 s), and 72 °C (5 min); (ii) 16S rRNA: 95 °C (4 min), 94 °C (35–37 repeats, 50 s), 50 °C (50 s), 72 °C (50 s), and 72 °C (5 min); (iii) 28S rRNA: 95 °C (4 min), 94 °C (36 repeats, 50 s), 62 °C (50 s), 72 °C (50 s), and 72 °C

Searching for the nearest neighbors. Each molecular sequence was checked via the basic local alignment search tool (BLAST; blast.ncbi.nlm.nih.gov) to search for the most similar sequences in NCBI GenBank. Additionally, we used the nearest-neighbor search algorithm implemented in the Barcoding of Life Data System (BOLD; www.boldsystems.org) with COI gene sequences of our specimens.

Sequence alignment. To estimate the phylogenetic position of three bivalve species, i.e., *L. fluminalis, S. deltae*, and *N. gangetica*, we sampled three sequence data sets, namely, "Pholadidae & relatives" (two partitions: *18S rRNA* + *28S rRNA*), "Arcidae & Noetiidae" (two partitions: *28S rRNA* + *18S rRNA*), and "Pharidae" (five partitions: *3* codons of *COI* + *16S rRNA* + *28S rRNA*), respectively (see Supplementary Table 6 for additional sequences obtained from GenBank). Sequences of the marine clam *Cavatidens omissa* Iredale, 1930 (Lucinidae) were used as an outgroup (Supplementary Table 6). The number of partitions in each data set was selected in accordance with the presence of available sequences for most closely related taxa in GenBank. The multiple sequence alignment was produced for each gene in each data set separately in MEGA66⁶³, with the Muscle ("Arcidae & Noetiidae" and "Pholadidae & relatives" data sets) and ClustalW ("Pharidae" data set) algorithms. The aligned sequence data sets were inspected with GBlocks v. 0.91b (Supplementary Table 7) to exclude gaps and variable sections from the alignments using a less strict set of options. Lacking sites were coded as missing positions.

Saturation analyses and congruence of phylogenetic signals. To estimate each partition in each data set for evidence of substitution saturation, we used the test of Xia et al. with DAMBE v. 5.3.108⁶⁴. The test revealed very little saturation even under the hypothesis of an asymmetrical tree. A partition homogeneity test was calculated in PAUP* v. 4.0a150 to check the congruence of phylogenetic signals among sequence partitions⁶⁵. This test returned the conformity of the phylogenetic signals among the partitions in the "Arcidae & Noetiidae" data set (P = 0.99). For the "Pharidae" data set, signals were also congruent (P > 0.3), with exception of those between the COI and 28S rRNA genes (P = 0.01) although there was no significant discordance between the 16S rRNA and 28S rRNA genes (P = 0.34). We concluded that the incongruence between these data partitions does not affect the model because it appears to be the effect of copious homoplasy³¹. We therefore used the combined data set (3 codons of COI + 16S rRNA + 28S rRNA) in phylogenetic analyses. In contrast, the test indicated significant incongruence between the 18S and 28S rRNA genes in the "Pholadidae & relatives" data set (P = 0.01). However, testing of a restricted data set, which includes only the members of this family and an outgroup taxon, returned no conflicts among partitions (P = 1.00). We therefore used this "Pholadidae" data set in subsequent reconstructions.

Phylogenetic analyses. Phylogenetic analyses were carried out with RAxML v. 8.2.6 HPC Black Box⁶⁶ and MrBayes v. $3.2.6^{67}$ at the San Diego Supercomputer Center through the CIPRES Science Gateway⁶⁸. In the RAxML analyses, a GTR+G model was selected for each partition. Nodal support values were calculated with a standard bootstrapping approach⁶⁶. The evolutionary models for each partition according to the corrected Akaike Information Criterion (AICc) of MEGA6⁶³ that were used under Bayesian inference framework are presented in Supplementary

Table 8. The MrBayes analyses were carried out for 25,000,000 generations with the following options: two runs, four Markov chains (three cold and one heated with temperature of 0.1), and sampling every 1000th cycle; 10% of the samples were excluded as an appropriate burn-in, and the convergence of the Markov chains was inspected with Tracer v. 1.6^{69} .

Ancestral area reconstructions. To estimate the ancestral area patterns, we calculated the sets of uncalibrated phylogenetic trees for each data set, i.e., "Pholadidae", "Arcidae & Noetiidae", and "Pharidae", using BEAST 2 v. 2.4.6 with a lognormal relaxed clock algorithm and the Yule speciation process as the tree prior . Models were calculated at the San Diego Supercomputer Center through the CIPRES Science Gateway⁶⁸. We applied the same options to the data sets as in the MrBayes analyses (see above). However, we selected simplified substitution models (see Supplementary Table 8). In each case, two independent runs with 25,000,000 generations were carried out. The phylogenies were reconstructed every 1000th cycle. The primary log data sets were inspected visually with Tracer v. 1.6 for the congruence of the Markov chains and the effective sample size of parameters (ESS)⁶⁹. The ESS values for all parameters were found as >350, and the posterior and prior distributions were congruent. For each data set, the phylogenies from two separate runs were joined with LogCombiner v. 1.8.3 (burn-in = 10%), and the maximum clade credibility tree was reconstructed with TreeAnnotator v. 1.8.371.

For each sequence data set, a sample of post-burn-in binary trees (N = 45,002) and a user-specified consensus tree obtained from BEAST runs were used to calculate ancestral area patterns on the basis of three algorithms, i.e., Statistical Dispersal-Vicariance Analysis (S-DIVA), Dispersal-Extinction Cladogenesis (DEC), and Statistical Dispersal-Extinction Cladogenesis (S-DEC) implemented in RASP v. 3.2^{72} . The out-group taxa were discarded from the phylogenies with RASP v. 3.2. Additionally, only one representative of each species was used in RASP analyses to avoid uncertainty of the biogeographic models. We assigned two possible ancestral areas of the in-group species, i.e., (a) marine and (ab) estuarine to freshwater. The S-DIVA scenarios were computed with two allowed areas, allowing 100 reconstructions with maximum reconstruction for final tree = 1000, and allowing extinctions. The DEC and S-DEC scenarios were reconstructed with default options and two allowed areas. Additionally, we calculated a combined model (S-DEC + DEC + S-DIVA) joining the independent scenarios inferred from three different algorithms in RASP v. 3.2.

Data availability. The sequences generated under this study are available from GenBank. Accession numbers for each specimen are presented in Table 1. The voucher specimens and samples of siltstone blocks with boreholes are available in the RMBH, Russian Museum of Biodiversity Hotspots, the Federal Center for Integrated Arctic Research, Russian Academy of Sciences (Arkhangelsk, Russia).

Received: 29 January 2018 Accepted: 15 June 2018 Published online: 23 July 2018

References

- Cáceres, L. M., Muñiz, F., Rodríguez-Vidal, J., Vargas, J. M. & Donaire, T. Marine bioerosion in rocks of the prehistoric tholos of La Pastora (Valencina de la Concepción, Seville, Spain): archaeological and palaeoenvironmental implications. J. Archaeol. Sci. 41, 435–446 (2014).
- Warme, J. E. in *The Study Of Trace Fossils* (ed Frey, R. W.) 181–227 (Springer, Berlin Heidelberg, 1975).
- Pinn, E. H., Thompson, R. C. & Hawkins, S. J. Piddocks (Mollusca: Bivalvia: Pholadidae) increase topographical complexity and species diversity in the intertidal. *Mar. Ecol. Prog. Ser.* 355, 173–182 (2008).
- Naylor, L. A., Coombes, M. A. & Viles, H. A. Reconceptualising the role of organisms in the erosion of rock coasts: a new model. *Geomorphology* 157, 17–30 (2012).
- Moura, D. et al. Integrated assessment of bioerosion, biocover and downwearing rates of carbonate rock shore platforms in southern Portugal. *Cont. Shelf Res.* 38, 79–88 (2012).

ARTICLE

- Glynn, P. W. & Manzello, D. P. in *Coral Reefs in the Anthropocene* (ed Birkeland, C.) 67–97 (Springer, Netherlands, 2015).
- Jones, C. G., Lawton, J. H. & Shachak, M. Organisms as ecosystem engineers. Oikos 69, 373–386 (1994).
- Wilson, M. A. in *Trace Fossils* (ed. Miller, W.) 356–367 New York, USA, (Elsevier, 2007).
- Bromley, R. G. A stratigraphy of marine bioerosion. *Geol. Soc. Spec. Publ.* 228, 455–479 (2004).
- Taylor, P. D. & Wilson, M. A. Palaeoecology and evolution of marine hard substrate communities. *Earth-Sci. Rev.* 62, 1–103 (2003).
- Smith, R. D. & Ross, A. J. Amberground pholadid bivalve borings and inclusions in Burmese amber: implications for proximity of resin-producing forests to brackish waters, and the age of the amber. *Earth Environ. Sci. Trans. R. Soc. Edinb.* 107, 239–247 (2018).
- Ricci, S., Perasso, C. S., Antonelli, F. & Petriaggi, B. D. Marine bivalves colonizing Roman artefacts recovered in the Gulf of Pozzuoli and in the Blue Grotto in Capri (Naples, Italy): boring and nestling species. *Int. Biodeterior. Biodegradation* 98, 89–100 (2015).
- Alagarswami, K. & Chellam, A. On fouling and boring organisms and mortality of pearl oysters in the farm at Veppalodai, Gulf of Mannar. *Indian J. Fish.* 23, 10–22 (1976).
- Distel, D. L. et al. Molecular phylogeny of Pholadoidea Lamarck, 1809 supports a single origin for xylotrophy (wood feeding) and xylotrophic bacterial endosymbiosis in Bivalvia. *Mol. Phylogenet. Evol.* 61, 245–254 (2011).
- Bromley, R. G., Hanken, N. M. & Asgaard, U. Shallow marine bioerosion: preliminary results of an experimental study. *Bull. Geol. Soc. Den.* 38, 85–99 (1990).
- Monari, S. Phylogeny and biogeography of pholadid bivalve *Barnea* (Anchomasa) with considerations on the phylogeny of Pholadoidea. *Acta Palaeontol. Pol.* 54, 315–335 (2009).
- Turner, R. D. & Santhakumaran, L. N. The genera Martesia and Lignopholas in the Indo-Pacific (Mollusca: Bivalvia: Pholadidae). Ophelia 30, 155–186 (1989).
- Annandale, N. Bivalve molluscs injuring brickwork in Calcutta docks. J. Asiat. Soc. Bengal 18, 555–557 (1922).
- Blanford, W. T. Contributions of Indian Malacology, No. VIII. List of estuary shells collected in the delta of the Irawady, in Pegu, with descriptions of new species. J. Asiat. Soc. Bengal 36, 51–72 (1867).
- Nederlof, R. & Muller, M. A biomechanical model of rock drilling in the piddock *Barnea candida* (Bivalvia; Mollusca). *J. R. Soc. Interface* 9, 2947–2958 (2012).
- Johnson, M. E., Wilson, M. A. & Redden, J. A. Borings in quartzite surf boulders from the Upper Cambrian basal Deadwood Formation, Black Hills of South Dakota. *Ichnos* 17, 48–55 (2010).
- Rodríguez-Tovar, F. J., Uchman, A. & Puga-Bernabéu, Á. Borings in gneiss boulders in the Miocene (Upper Tortonian) of the Sorbas Basin, SE Spain. *Geol. Mag.* 152, 287–297 (2015).
- Kelly, S. R. & Bromley, R. G. Ichnological nomenclature of clavate borings. Palaeontology 27, 793–807 (1984).
- Wilson, M. A. & Palmer, T. J. The earliest Gastrochaenolites (Early Pennsylvanian, Arkansas, USA): an upper Paleozoic bivalve boring? J. Paleontol. 72, 769–772 (1998).
- Hagan, T. H., Coniglio, M. & Edwards, T. W. Subfossil bioerosion of mollusc shells from a freshwater lake, Ontario, Canada. *Ichnos* 6, 117–127 (1998).
- Jones, M. L. On the Caobangiidae, a new family of the Polychaeta, with a redescription of *Caobangia billeti* Giard. *Smithson. Contrib. Zool.* 175, 1–55 (1974).
- Annandale, N. The marine element in the fauna of the Ganges. *Bijdr. tot De. Dierkd.* 22, 143–154 (1922a).
- Jakhalekar, S. S. & Ghate, H. V. Taxonomy of freshwater sponges of Maharashtra, India, with illustrated descriptions and notes on ecology and habitats (Porifera: Spongillida: Spongillidae). Zootaxa 4173, 501–529 (2016).
- Nesemann, H. A., Sharma, S. U., Sharma, G. O. & Sinha, R. K. Illustrated checklist of large freshwater bivalves of the Ganga river system (Mollusca: Bivalvia: Solecurtidae, Unionidae, Amblemidae). *Nachrichchtenblatt der Ersten Vorarlberg. Malakol. Ges.* 13, 1–51 (2005).
- Nesemann, H., Sharma, G. & Sinha, R. Benthic macro-invertebrate fauna and "marine elements" sensu Annandale (1922) highlight the valuable dolphin habitat of river Ganga in Bihar-India. *Taprobanica* 3, 18–30 (2011).
- Bolotov, I. N. et al. Ancient river inference explains exceptional Oriental freshwater mussel radiations. Sci. Rep. 7, 2135 (2017).
- 32. Bolotov, I. N. et al. New taxa of freshwater mussels (Unionidae) from a species-rich but overlooked evolutionary hotspot in Southeast Asia. *Sci. Rep.* **7**, 11573 (2017a).
- Lee, C. E. Rapid and repeated invasions of fresh water by the copepod Eurytemora affinis. Evolution 53, 1423–1434 (1999).
- Wilson, M. A., Ozanne, C. R. & Palmer, T. J. Origin and paleoecology of freerolling oyster accumulations (ostreoliths) in the Middle Jurassic of southwestern Utah, USA. *Palaios* 13, 70–78 (1998).

- Isbell, F. et al. Linking the influence and dependence of people on biodiversity across scales. *Nature* 546, 65–72 (2017).
- Guisan, A., Petitpierre, B., Broennimann, O., Daehler, C. & Kueffer, C. Unifying niche shift studies: insights from biological invasions. *Trends Ecol. Evol.* 29, 260–269 (2014).
- Ochocki, B. M. & Miller, T. E. Rapid evolution of dispersal ability makes biological invasions faster and more variable. *Nat. Commun.* 8, 14315 (2017).
- Çevik, C., Özcan, T. & Gündoğdu, S. First record of the striate piddock Martesia striata (Linnaeus, 1758) (Mollusca: Bivalvia: Pholadidae) in the Mediterranean Sea. Bioinvasions Rec. 4, 277–280 (2015).
- Kaladan Multi-Modal Transit Transport Project. The Government of India, Ministry of Development of North Eastern Region. http://www.mdoner.gov.in/ content/introduction-1 (2014).
- Karatayev, A. Y., Boltovskoy, D., Padilla, D. K. & Burlakova, L. E. The invasive bivalves *Dreissena polymorpha* and *Limnoperna fortunei*: parallels, contrasts, potential spread and invasion impacts. J. Shellfish Res. 26, 205–213 (2007).
- Pimentel, D., Zuniga, R. & Morrison, D. Update on the environmental and economic costs associated with alien-invasive species in the United States. *Ecol. Econ.* 52, 273–288 (2005).
- Brandt, R. A. M. The non-marine aquatic Mollusca of Thailand. Arch. Molluskenkd. 105, 1–423 (1974).
- Morton, B. & Dinesen, G. E. Colonization of Asian freshwaters by the Mytilidae (Bivalvia): a comparison of *Sinomytilus harmandi* from the Tonle-Sap River, Phnom Penh, Cambodia, with *Limnoperna fortunei*. *Molluscan Res.* 30, 57–72 (2010).
- Hou, Z. & Li, S. Tethyan changes shaped aquatic diversification. *Biol. Rev.* 10.1111/brv.12376 (2017).
- Seehausen, O. & Wagner, C. E. Speciation in freshwater fishes. Annu. Rev. Ecol. Evol. Syst. 45, 621–651 (2014).
- Cuny, G. et al. Shark faunas from the Late Jurassic—Early Cretaceous of northeastern Thailand. *Paläontol. Z.* 88, 309–328 (2014).
- Cuny, G., Suteethorn, V., Kamha, S., Buffetaut, E. & Philippe, M. A new hybodont shark assemblage from the Lower Cretaceous of Thailand. *Hist. Biol.* 18, 21–31 (2006).
- Kirchhoff, K. N., Hauffe, T., Stelbrink, B., Albrecht, C. & Wilke, T. Evolutionary bottlenecks in brackish water habitats drive the colonization of fresh water by stingrays. *J. Evol. Biol.* **30**, 1576–1591 (2017).
- Betancur-R, R. Molecular phylogenetics supports multiple evolutionary transitions from marine to freshwater habitats in ariid catfishes. *Mol. Phylogenet. Evol.* 55, 249–258 (2010).
- Yamanoue, Y. et al. Multiple invasions into freshwater by pufferfishes (Teleostei: Tetraodontidae): a mitogenomic perspective. *PLoS ONE* 6, e17410 (2011).
- Beck, S. V. et al. Plio-Pleistocene phylogeography of the Southeast Asian Blue Panchax killifish. *Aplocheilus panchax*. PLoS ONE 12, e0179557 (2017).
- Wowor, D. et al. Evolution of life history traits in Asian freshwater prawns of the genus *Macrobrachium* (Crustacea: Decapoda: Palaemonidae) based on multilocus molecular phylogenetic analysis. *Mol. Phylogenet. Evol.* 52, 340–350 (2009).
- Glasby, C. J., Fiege, D. & Van Damme, K. Stygobiont polychaetes: notes on the morphology and the origins of groundwater *Namanereis* (Annelida: Nereididae: Namanereidinae), with a description of two new species. *Zool. J. Linn. Soc.* **171**, 22–37 (2014).
- Licht, A., Reisberg, L., France-Lanord, C., Naing Soe, A. & Jaeger, J. J. Cenozoic evolution of the central Myanmar drainage system: insights from sediment provenance in the Minbu Sub-Basin. *Basin Res.* 28, 237–251 (2016).
- Voris, H. K. Maps of Pleistocene sea levels in Southeast Asia: shorelines, river systems and time durations. J. Biogeogr. 27, 1153–1167 (2000).
- Caldwell, P. C., Merrifield, M. A. & Thompson, P. R. Sea level measured by tide gauges from global oceans — the Joint Archive for Sea Level holdings (NCEI Accession 0019568), Version 5.5, NOAA National Centers for Environmental Information. *Dataset*. https://doi.org/10.7289/V5V40S7W (2015).
- 57. Than Tin, Aung et al. Geologic evidence for three great earthquakes in the past 3400 years off Myanmar. *J. Earthq. Tsunami* **2**, 259–265 (2008).
- Than Tin, Aung et al. Paleoseismological field survey along the western coast of Myanmar. Annual Report on Active Fault and Paleoearthquake Researches. *Geol. Surv. Jpn.* 6, 171–188 (2006).
- Wang, Y. et al. Permanent upper plate deformation in western Myanmar during the great 1762 earthquake: Implications for neotectonic behavior of the northern Sunda megathrust. *J. Geophys. Res.* 118, 1277–1303 (2013).
- Garzanti, E. et al. Sediment recycling at convergent plate margins (Indo-Burman ranges and Andaman–Nicobar Ridge). *Earth-Sci. Rev.* 123, 113–132 (2013).
- Manconi, R. & Pronzato, R. Rediscovery of the type material of *Spongilla lacustris* (L., 1759) in the Linnean herbarium. *Ital. J. Zool.* 67, 89–92 (2000).

- Hall, T. A. BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symp. Ser.* 41, 95–98 (1999).
- Tamura, K., Stecher, G., Peterson, D., Filipski, A. & Kumar, S. MEGA6: molecular evolutionary genetics analysis version 6.0. *Mol. Biol. Evol.* 30, 2725–2729 (2013).
- Xia, X. & Lemey, P. in *The Phylogenetic Handbook: A Practical Approach to* DNA and Protein Phylogeny 2nd edn (eds Lemey, P., Salemi, M. & Vandamme, A.) 615–630 (Cambridge University Press, Cambridge, UK, 2009).
- Swofford, D.L. PAUP*. Phylogenetic Analysis Using Parsimony (*and Other Methods). V. 4.0b10 (Sinauer Associates, Sunderland, Massachusetts, 2002).
- Stamatakis, A. RAxML-VI-HPC: maximum likelihood-based phylogenetic analyses with thousands of taxa and mixed models. *Bioinformatics* 22, 2688–2690 (2006).
- 67. Ronquist, F. et al. MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. *Syst. Biol.* **61**, 539–542 (2012).
- Miller, M., Pfeiffer, W. & Schwartz, T. Creating the CIPRES Science Gateway for inference of large phylogenetic trees. In *Gateway Computing Environments Workshop (GCE)* 1–8 (New Orleans Convention Center New Orleans, LA, USA, 2010).
- Rambaut, A., Suchard, M. & Drummond, A. J. Tracer v1.6. http://beast.bio.ed. ac.uk/software/tracer/ (2013).
- Bouckaert, R. et al. BEAST 2: a software platform for bayesian evolutionary analysis. *PLoS Comput. Biol.* 10, e1003537 (2014).
- Drummond, A. J., Suchard, M. A., Xie, D. & Rambaut, A. Bayesian phylogenetics with BEAUti and the BEAST 1.7. *Mol. Biol. Evol.* 29, 1969–1973 (2012).
- Yu, Y., Harris, A. J., Blair, C. & He, X. J. RASP (Reconstruct Ancestral State in Phylogenies): a tool for historical biogeography. *Mol. Phylogenet. Evol.* 87, 46–49 (2015).
- Combosch, D. J. & Giribet, G. Clarifying phylogenetic relationships and the evolutionary history of the bivalve order Arcida (Mollusca: Bivalvia: Pteriomorphia). *Mol. Phylogenet. Evol.* 94, 298–312 (2016).
- 74. Vierna, J. et al. Species delimitation and DNA barcoding of Atlantic *Ensis* (Bivalvia, Pharidae). *Zool. Scripta* **43**, 161–171 (2014).
- Quintero-Galvis, J. & Raquel-Castro, L. Molecular phylogeny of the Neritidae (Gastropoda: Neritimorpha) based on the mitochondrial genes cytochrome oxidase I (COI) and 16S rRNA. Acta Biol. Colomb. 18, 307–318 (2013).
- Glasby, C. J. & Timm, T. Global diversity of polychaetes (Polychaeta; Annelida) in freshwater. *Hydrobiologia* 595, 107–115 (2008).
- Meixner, M. J. et al. Phylogenetic analysis of freshwater sponges provide evidence for endemism and radiation in ancient lakes. *Mol. Phylogenet. Evol.* 45, 875–886 (2007).

Acknowledgements

This work was partly funded by grants from the Russian Ministry of Education and Science (project no. 6.2343.2017/4.6), Federal Agency for Scientific Organizations (project nos. 0409-2015-0143 and 0409-2016-0022), National Geographic Society (project no. NGS-274R-18), Russian Foundation for Basic Research (project nos. 16-34-00638, 17-45-290066, 17-44-290016, and 16-34-00125), and Northern Arctic Federal University. The geochemical study of rock substrate was supported by the Russian Science Foundation (project no. 14-50-00029) and Lomonosov Moscow State University Program of Development. The SEM study of sponge samples was conducted using the Joint Usage Center 'Instrumental Methods in Ecology' at A. N. Severtsov Institute of Ecology and Evolution, Russian Academy of Sciences. The work of A.M.S. was conducted under IDB RAS GBRP (project no. 0108-2018-0002). We are grateful to the late Dr. Tony Whitten (Fauna & Flora International—Asia-Pacific), Mr. Frank Momberg, Mr. Zau Lunn, and Mr. Nyein Chan (Fauna & Flora International—Myanmar Program), Prof. Dr. Myin Zu Minn and Prof. Dr. Thida Lay Thwe (University of Yangon, Myanmar), and local villagers living along the Kaladan and Lemro rivers for their great help during this study. We would like to express our sincerest gratitude to the Department of Fisheries of the Ministry of Agriculture, Livestock and Irrigation (Myanmar) for the permission for the field work and sampling in Myanmar (survey permission no. 5/6000/MOLFRD-3103/2016 and export permission no. Nga La/Nga Tha Hta-Phont Thu/2016-5856).

Author contributions

I.N.B. developed the concept of the study. I.N.B., I.V.V., E.S.K., O.V.A., and M.Y.G. coordinated the samples collection. A.V.K. designed and carried out the molecular analyses, with contribution from E.S.K. I.N.B., O.V.A., and E.S.K. identified the mollusc taxa. I.N.B. identified the ichnospecies. C.J.G. and T.B. identified the polychaete taxa. A.M.S. identified the freshwater sponge species. O.S.P. and A.Y.B. carried out geological and mineralogical analyses of the rock substrate. I.N.B., O.V.A., E.S.K., C.J.G., T.B., and A.M.S. wrote the corresponding taxonomic accounts. M.Y.G. created the maps. I.N.B. and M.Y.G. created the model of neotectonic evolution of the lower course of the Kaladan River, with contribution from J.B.H.S. and Y.W. I.N.B. performed the phylogenetic and biogeographic modeling. O.V.A., E.S.K., I.V.V., and A.M.S. pictured the samples. I.N.B. wrote the paper, with inputs from O.S.P., O.V.A., T.B., C.J.G., M.Y.G., A.V.K., E.S.K., M.L.-L., A.Y.B., A.M.S., K.T., S.T., I.V.V., and T.W. All authors approved the final version of the manuscript.

Additional information

Supplementary Information accompanies this paper at https://doi.org/10.1038/s41467-018-05133-4.

Competing interests: The authors declare no competing interests.

Reprints and permission information is available online at http://npg.nature.com/ reprintsandpermissions/

Publisher's note: Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit http://creativecommons.org/ licenses/by/4.0/.

© The Author(s) 2018